



## Effect of Colchicine and Bio-catharantin on the DNA Relative Content and Stomatal Structure of Black Rice (*Oryza sativa* L. var. Jeliteng)

Dwi Setyati<sup>1</sup>, Mukhamad Su'udi<sup>1</sup>, Dyah Retno Wulandari<sup>2</sup>, Tri Handoyo<sup>3</sup> and Fuad Bahrul Ulum<sup>1\*</sup>

<sup>1</sup>Department of Biology, Faculty of Mathematics and Sciences, Universitas Jember, Jember, Indonesia;

<sup>2</sup>Research Center for Applied Botany, National Research and Innovation Agency (BRIN), Indonesia;

<sup>3</sup>Department of Agronomy, Faculty of Agriculture, Universitas Jember, Jember, Indonesia

\*Corresponding author: [fuad.fmipa@unej.ac.id](mailto:fuad.fmipa@unej.ac.id)

### Abstract

Black rice (*Oryza sativa* L. var. Jeliteng), known for its health benefits compared to white rice, faces challenges in productivity. Among varieties, this black rice is popular in Indonesia but shows low yield. Research on improving black rice through genetic manipulation with antimetabolic substances is limited. Therefore, this study aims to compare the effects of colchicine and Bio-catharantin on the germination rate, DNA relative content, and stomatal structure of *O. sativa* L. var. Jeliteng. Seeds were treated with colchicine (0.1%, 0.2%, and 0.3%) and Bio-catharantin (0.1%, 0.2%, 0.3%, 0.4%, and 0.5%) at soaking durations of 12, 24, and 48 hours. Germination was assayed, ploidy was determined using flow cytometry, and stomatal traits, including size and density, were examined microscopically. The results showed that Bio-catharantin did not exhibit any toxic effects on germination rates, whereas colchicine reduced germination starting at 0.2% concentration. Both chemical agents modified the DNA relative content of Jeliteng black rice. Colchicine generally increased stomatal length and width while decreasing stomatal density, with significant changes at 0.3% concentration for 24 hours. Bio-catharantin also altered stomatal traits, enhancing length and width in most cases but significantly reducing density under certain conditions. Bio-catharantin emerged as a promising alternative to colchicine for inducing chromosomal mutations in plants, offering benefits in altered stomatal structures without the toxic effects on germination, compared to colchicine.

**Keywords:** antimetabolic; crop improvement; flow cytometry; germination

**Cite this as:** Setyati, D., Su'udi, M., Wulandari, D. R., Handoyo, T., & Ulum, F. B. (2024). Effect of Colchicine and Bio-catharantin on the DNA Relative Content and Stomatal Structure of Black Rice (*Oryza sativa* L. var. Jeliteng). *Caraka Tani: Journal of Sustainable Agriculture*, 39(2), 465-478. doi: <http://dx.doi.org/10.20961/carakatani.v39i2.88279>

### INTRODUCTION

Rice serves as the primary staple food for 95% of the Indonesian population (Paiman et al., 2020). Based on its pigment content, rice is categorized as either pigmented or non-pigmented. Pigmented rice contains bioactive compounds in the pericarp layer of the rice that produce color pigments of brown, black, purple, or red (Xia et al., 2021; Nabilah et al., 2022). Certain bioactive compounds, such as polyphenols, flavonoids, and antioxidants

are present in black rice (Suryanti et al., 2020; Tyagi et al., 2022). The consumption of black rice as a functional food was increasingly popular as people became more health-conscious (Suryanti et al., 2020; Nabilah et al., 2022). Black rice offers various health benefits, such as cancer prevention, free radical defense, improved digestion, cholesterol reduction, and anti-inflammatory effects (Luo et al., 2014; Thepthanee et al., 2021).

\* Received for publication June 14, 2024

Accepted after corrections September 12, 2024

The black rice *Oryza sativa* L. var Jeliteng is a superior variety with a fluffy texture and an amylose content of 19.6% (Nandariyah et al., 2023; Nurhidajah et al., 2024). This black rice variety originated from the hybridization of black sticky rice (*ketan hitam*) and commercial rice seed (Pandan Wangi Cianjur) (Sitaresmi et al., 2023). It contained high levels of phenolic content ( $29 \pm 3.6$  mg FAE  $g^{-1}$ ) (Wijayanti et al., 2023). The harvest age of Jeliteng was 113 days with a productivity of 6.18 tons (Nandariyah et al., 2022). The agronomic advantages of this rice were susceptibility to rice tungro spherical virus (RTSV) and rice tungro bacilliform virus (RTBV) and moderate susceptibility to *Nilaparvata lugens* or brown planthopper (Indonesian Rice Research Center, 2019).

Although Jeliteng rice was considered a superior variety, it was not extensively cultivated. Conventional farmers prefer to cultivate the common commercial white rice due to its shorter lifespan of 90 days and higher productivity of 8.99 tons (Nandariyah et al., 2022; 2023). Therefore, breeding efforts are necessary to enhance the qualities of the Jeliteng black rice. In rice breeding strategies, mutation breeding using chemical agents such as colchicine, ethyl methanesulfonate (EMS), and sodium azide (SA) induces random mutations that can enhance specific traits while minimizing undesirable changes and to some extent induced polyploid plants. Irradiation methods like fast-neutron irradiation and ion beam radiation are also employed to generate genetic variability, targeting important traits such as disease resistance and abiotic stress tolerance (Viana et al., 2019). Genetic manipulation through cell treatment with antimetabolic induction could be performed to enhance the qualities of rice (Chen et al., 2021) for sustainable agriculture in Indonesia (Hafeez et al., 2023).

Genetic manipulation can be induced either physically, biologically or using antimetabolic chemical agents, where the latter is the most widely applied method (Chen et al., 2021). Colchicine is an antimetabolic agent that impedes anaphase, hence preventing the separation and transfer of chromosomes to the opposite pole. This mechanism produces individuals that contain multiple sets of chromosomes or in another case altered variations in phenotypes (Eng and Ho, 2018). The alternation of phenotypic features such as the enlargement of roots, stems, leaves, fruits, and flowers (Ermayanti et al., 2018), as well as stomatal size (Rohmah et al., 2017;

Wu et al., 2022). The application of chemical compounds was desirable due to their effectiveness, lower detrimental characteristics, and long-lasting characteristic changes (Chen et al., 2021).

The application of colchicine in plant breeding induced genetic mutation resulted in metabolite alternation (Samadi et al., 2022), and enhanced stomatal size by increasing chloroplasts in guard cells, leading to a larger stomatal size (Yao et al., 2023). Stomatal size and density have an inverse relationship, as larger stomatal size results in decreased stomatal density (Tossi et al., 2022). Nonetheless, the use of colchicine as a means to reduce stomatal density is limited due to its toxicity, which can also negatively affect plant germination (Kurniawan et al., 2023). Moreover, these compounds possess toxic properties for humans, particularly when used in high concentrations (Eng and Ho, 2018). Natural antimetabolic compounds, such as Bio-catharantin extracted from *Catharanthus roseus*, have the potential to serve as a polyploid inducer and avoid these toxic effects (Muarifin et al., 2021; Billa et al., 2022; Kurniawan et al., 2023; Kasim et al., 2024). Bio-catharantin was successfully used to enhance the chromosome number of *Allium cepa* L. var. *ascalonicum* 'Tajuk' from diploid (2n) into triploid (3n) and tetraploid (4n) at concentrations of 0.2% and 0.4%, respectively (Billa et al., 2022). Bio-catharantin was also reported to improve the growth performance of *Alternanthera amoena* Voss., even in the absence of polyploid individuals (Shafura et al., 2022). Another study reported that Bio-catharantin altered the stomata and chromosome structure of *O. sativa* var. Cempo Ireng without negative effect on the seed germination (Kurniawan et al., 2023).

The research on the improvement of black rice through genetic modification using antimetabolic compounds in Indonesia remains limited (Dwiningsih and Alkahtani, 2023). By employing a natural compound, Bio-catharantin, which has no toxic impact, this research has paved the way for the development of effective antimetabolic compounds in the genetic modification of black rice. This study aims to compare the impact of genetic modification by using antimetabolic compounds, specifically colchicine and Bio-catharantin, on the germination rates, DNA relative, and stomatal structure of the black rice *O. sativa* L. var Jeliteng. This study investigated the toxic effects of combined treatments of colchicine and Bio-catharantin at various

concentrations and soaking durations on seed germination rates. Subsequently, the paddy leaves were subjected to a flow cytometric analysis to ascertain the impact of the treatments on the chromosomes of the black rice's somatic cells. Further observations of the stomatal structure were conducted to compare the changes in size and density of the induced rice.

## MATERIALS AND METHOD

### Materials

The black rice *O. sativa* L. var. Jeliteng seeds were obtained from the Center for Rice Agriculture Research and Development, Ministry of Agriculture with registration number 167/HK.540/C/01/2019. The Bio-catharantin was a commercial product from the research group of the Faculty of Biology at Universitas Gadjah Mada, Indonesia. Colchicine was the commercial product of C9754 Sigma-Aldrich  $\geq 95\%$  (High performance liquid chromatography/HPLC) powder.

### Polyplloid induction and germination rate

The Jeliteng rice seeds were peeled and sterilized in 70% ethanol for 5 minutes, then diluted in 0.1% NaCl solution for 10 minutes. Rice seeds were subsequently rinsed with sterile distilled water 4 times and dried with sterile filter paper for 2 hours (Prasetyo et al., 2018). The polyploidy induction was carried out following the method proposed by Gaafar et al. (2017). A total of 100 seeds were soaked in each concentration and duration series using 2 antimitotic substances, colchicine and Bio-catharantin. Colchicine is a highly toxic chemical compound (Chen et al., 2021); therefore, the lower concentration was applied. The colchicine solution concentrations were 0.1%, 0.2%, and 0.3%, and Bio-catharantin solution concentrations were 0.1%, 0.2%, 0.3%, 0.4%, and 0.5%. The soaking times for colchicine were 24 and 48 hours and for Bio-catharantin were 12, 24, and 48 hours. The seeds were then germinated in a tray containing clay soil in the Biology Department greenhouse (Universitas Jember) for 14 days. Observations of germination were made by counting the number of seeds that germinated normally within 7 to 14 days. The normal shoots were indicated by the presence of a root system with primary and seminal roots, well-developed hypocotyl without tissue damage, perfect plumula growth with green leaves emerging from the coleoptile, and epicotyl growth with normal buds. The germination rate was calculated based on

the proportion of germinated seeds among the total seeds of each treatment (Kurniawan et al., 2023). All treatments were replicated a minimum of 3 times.

### Flow cytometry test

Fresh rice leaves were cut into 0.5 cm<sup>2</sup> pieces and placed on a petri dish. Next, 250  $\mu$ l of CyStain PI Absolute (nuclei extraction buffer) was dripped onto the leaves. Using a razor blade, the leaves were finely chopped and filtered with a 30  $\mu$ m Millipore sieve. The filtered filtrate was transferred to a cuvette tube and mixed with 350  $\mu$ l staining solution, consisting of propidium iodine and RNase. Finally, the DNA relative content was measured using the BD Accuri™ C6 Plus Flow cytometer for ploidy analysis (Hodač et al., 2016).

### Stomatal size and density

The abaxial leaves were cleaned and coated with transparent nail polish. After the polish had dried, a strip of tape was applied to the polished area. The tape was slowly removed, causing the epidermis to peel off and adhere to a slide. Stomatal characteristics were observed using a Nikon Eclipse E100 LED MV R microscope connected to Optilab Advance by Miconos at 400x magnification. Stomatal length and width were analyzed utilizing Image Raster software. Stomatal density was computed using Equation 1 (Chatterjee et al., 2020).

$$\text{Stomatal density} = \frac{\text{Number of stomata}}{\text{Field of view area in mm}^2} \quad (1)$$

### Statistical analysis

Data were analyzed using R version 4.1.2 for Windows (R Foundation for Statistical Computing). Statistical data were visualized with ggplot2 (Wickham et al., 2016). Significant differences were assessed using either parametric (ANOVA) or non-parametric (Kruskal Wallis) tests. Post-hoc analysis was conducted using either the Student's T-test, Wilcox test, or Duncan's test, with the 'agricolae' package (de Mendiburu and de Mendiburu, 2019).

## RESULTS AND DISCUSSION

### Colchicine and Bio-catharantin effect on seed germination

Colchicine at a higher concentration (more than 0.2%) reduced the germination rate of black rice seeds, whereas Bio-catharantin showed no adverse effects on the germination rate (Figure 1). The germination rate of Jeliteng black rice seeds

was 91.29% (Supplementary Table 1). The statistical analysis confirmed that the germination rate of the seeds under Bio-catharantin treatments remained within the range of the control treatment (83.33 to 93.33%). In contrast, the colchicine toxicity to the seeds became apparent at a concentration of 0.2% with a 24-hour soaking duration, resulting in a noticeable decline in germination. The negative effect of colchicine increased with higher concentrations and soaking durations (Figure 1).

Colchicine is one of the most common chemical mutagens for inducing polyploidy in plants (Chen et al., 2021). Polyploidization with colchicine has been utilized in horticultural crops for more than 8 decades and continues to be a leading antimitotic agent for polyploid production. A total of 46 plant mutants with colchicine had been registered in FAO in 2018. As research on the use of colchicine in the polyploidization of horticultural crops grows, its effectiveness remains prominent. The success of polyploidization with this mutagenic was related to the concentration and duration of treatment (Eng and Ho, 2018). The consideration of concentration affected the seedling viability. Application of colchicine in a higher concentration reduced the seed germination in related family Poace, i.e. *Dendrocalamus brandisii* (Munro) Kurz by inhibiting cell

division, DNA synthesis, and sucrose degradation (Lv et al., 2021). Another report also confirmed the negative effect of colchicine in high concentration that reduced the seed germination and plant growth of the medicinal plant *Salvia hains* (Grouh et al., 2011). Toxic effects can be circumvented by employing natural antimitotic compounds, for instance, Bio-catharantin from *Catharanthus roseus* extract. Bio-catharantin has the potential to serve as a polyploid inducer (Kurniawan et al., 2023). It was reported successfully inducing polyploidization with no toxicity effect on *Arachis hypogaea* (Muarifin et al., 2021), *A. cepa* (Billa et al., 2022), and *A. amoena* (Shafura et al., 2022).

### Colchicine and Bio-catharantin effect on DNA relative

The DNA relative of the black rice was measured using flow cytometry analysis as presented in Figure 2. The flow cytometer data revealed that none of the treatments exhibited polyploid in the leaf samples. However, the antimitotic compounds affected the chromosome structure, resulting in a slight shift in the peak index (PI). The relative DNA content of the control was 50.95, while the Bio-catharantin-treated seeds exhibited the greatest alteration in PI at 55.85 (Figure 2e), followed by colchicine at 54.85 (Figure 2i). The flow cytometer's X axis

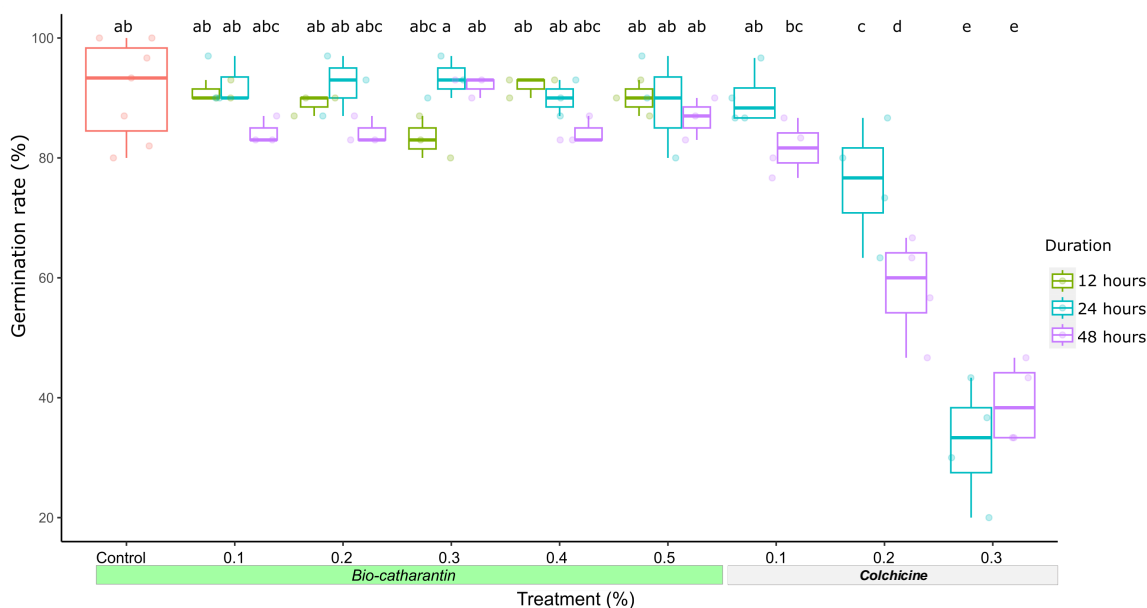


Figure 1. Germination rates for *O. sativa* L. var. Jeliteng seeds under Bio-catharantin and colchicine treatment

Note: The box and whisker plots display the distribution across the 25<sup>th</sup> and 75<sup>th</sup> percentile range and median. The jitter plot illustrates the distribution of the data. Significantly different values of the Duncan test ( $p < 0.005$ ) were indicated by different letters. Detailed data were provided in Supplementary Table 1

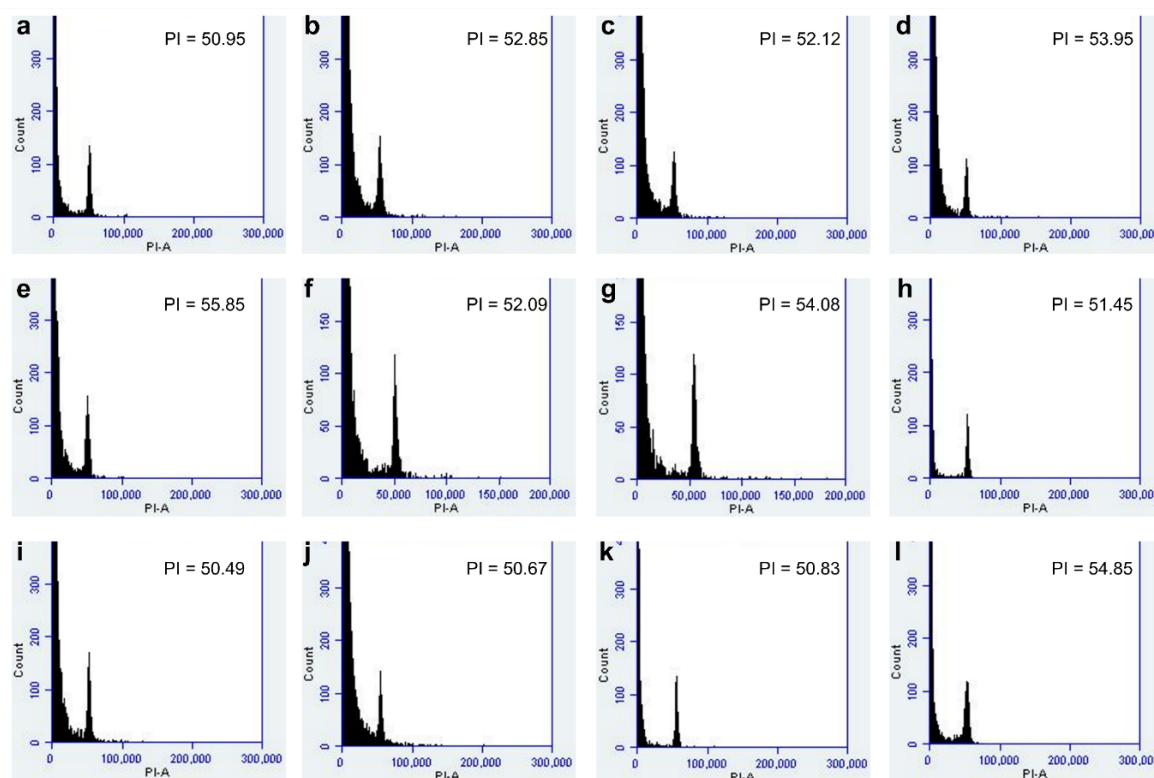


Figure 2. The ploidy level of leaf samples of *O. sativa* L. var. Jeliteng using a flow cytometer. Samples were categorized as a) Control; b-f = Bio-catharantin with shocking duration 48 hours, b) 0.1 %; c) 0.2 %; d) 0.3 %; e) 0.4 %; and f) 0.5 %; g-j = colchicine: g) 0.1% for 24 hours, h) 0.1 % for 48 hours; i) 0.2% for 48 hours; j) 0.2% for 48 hours; k) 0.3% for 48 hours; l) control. DNA relative to the mean value PI was recorded for each sample

displays the sample's relative DNA content, while the Y axis indicates the number of cells detected by the device.

The flow cytometer is essential for analyzing the DNA relative content. This high-throughput machine can assess a large number of nuclei (100 to 10,000 cells per second) and evaluate multiple samples in a single run (Hodač et al., 2016). Polyploid plants exhibit an increase in total DNA content in the cell nucleus while doubling chromosomes, and thus total DNA content serves as an indicator for ploidy determination in plants (Moghbel et al., 2015). Each plant type has a distinct optimal time and concentration for genetic mutation (Eng and Ho, 2018). Polyploid induction on diploid *Mentha spicata* L. with 40  $\mu$ M oryzalin for 48 hours produced a stable hexaploid genotype with a higher essential oil yield (Bharati et al., 2023). In *Colocasia esculenta*, soaking with 0.1% and 0.2% colchicine for 24 hours resulted in mixoploid and tetraploid plants, but longer exposure (48 hours) did not lead to polyploidy (Ermayanti et al., 2018). Another study reported

the genotype mutation of *Passiflora foetida*, where treatment of Bio-catharantin on the concentration of 1 to 1.5% for 24 to 72 hours only produced mixoploid, while a significantly lower concentration of colchicine led to the formation of tetraploid (Kasim et al., 2024). If the mutagenic solution concentration is too low, the polyploidy properties cannot be obtained. Even so, chromosomes will still polymerize into microtubules, resulting in spindle thread formation and diploid properties (Eng and Ho, 2018). On the other hand, if the concentration is too high or the soaking time is too long, the cell differentiation process will be hindered (Trojak-Goluch and Skomra, 2013), and chromosome doubling cannot occur. Cells undergo plasmolysis, which prevents the formation of polyploid properties (Eng and Ho, 2018). Additionally, the Jeliteng variety of black rice contains high levels of phenolic compounds (Nandariyah et al., 2022; Nurhidajah et al., 2024), which have antimutagenic properties that might inhibit colchicine-induced mutations in chromosomes.

### Colchicine and Bio-catharantin effect on stomatal size and densities

The stomata of *O. sativa* var. Jeliteng exhibits typical gramineae characteristics, with 2 dumbbell-shaped guard cells flanked by 2 lateral subsidiary cells (Figure 3). The experiment showed that altering the concentration of colchicine and soaking time affected the length, width, and density of stomata (Figure 4). In almost all colchicine-treated seeds, stomata were significantly wider and longer compared to the control group (Figure 4a and 4b). The control group exhibited an average length of 20.6  $\mu\text{m}$  and a width of 13.9  $\mu\text{m}$ . In treated seeds, the longest and widest stomata were observed at a concentration of 0.3% for 24 hours, measuring 24.7  $\mu\text{m}$  in length and 16.5  $\mu\text{m}$  in width. Conversely, the shortest stomatal length was at 0.2% for 24 hours, with a length of 21  $\mu\text{m}$ , and the shortest width was at a concentration of 0.2%, measuring 14.5  $\mu\text{m}$  (Supplementary Table 2). The stomatal density of the control group was higher than some colchicine-treated plants (Figure 4c). The control group had an average stomatal

density of 330  $\text{mm}^2$ , whereas the treated group had values ranging from 159 to 281  $\text{mm}^2$ . The treatment with 0.2% concentration for 48 hours showed the highest stomatal density value of 281  $\text{mm}^2$ , while the treatment with 0.3% concentration for 24 hours showed the lowest value of 159  $\text{mm}^2$  (Supplementary Table 2). The reduction of stomatal density under colchicine treatments was significantly observed in 3 treatments, i.e., at the concentrations of 0.1% for 48 hours, 0.3% for 24 hours, and 0.3% for 48 hours.

On the other hand, the Bio-catharantin altered the stomatal length, width, and density of black rice in several treatments (Figure 4). The average stomatal length treated with Bio-catharantin was within the range of 15.3 to 23.7  $\mu\text{m}$ , while the average stomatal width was within the range of 13.6 to 20.3  $\mu\text{m}$  (Supplementary Table 2). The stomatal length under Bio-catharantin of 0.1% varied based on different soaking durations. A 12-hour treatment reduced the stomatal length, a 24-hour treatment resulted in a length similar to the control group, and the 48-hour treatment significantly increased the stomatal length

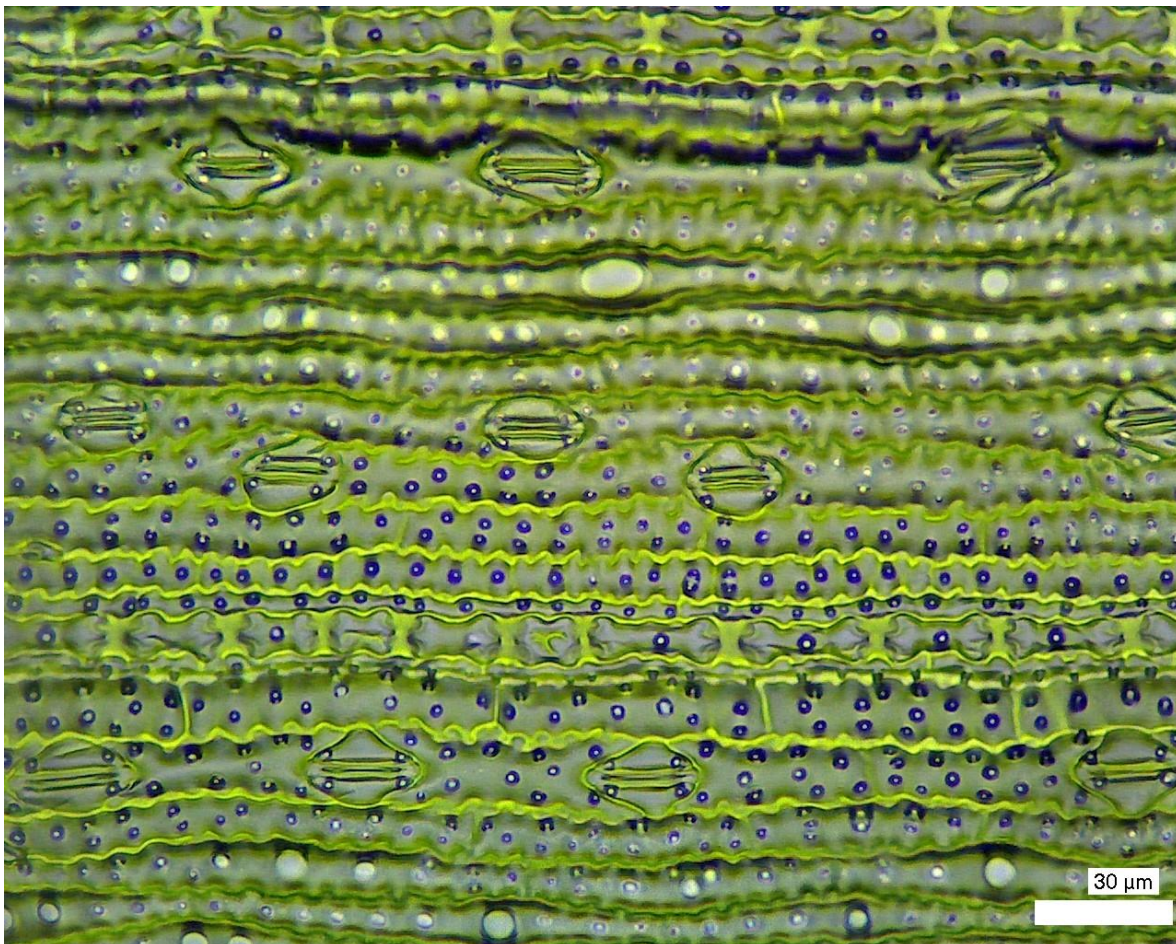


Figure 3. Stomatal structure of *O. sativa* L. var. Jeliteng under control treatment

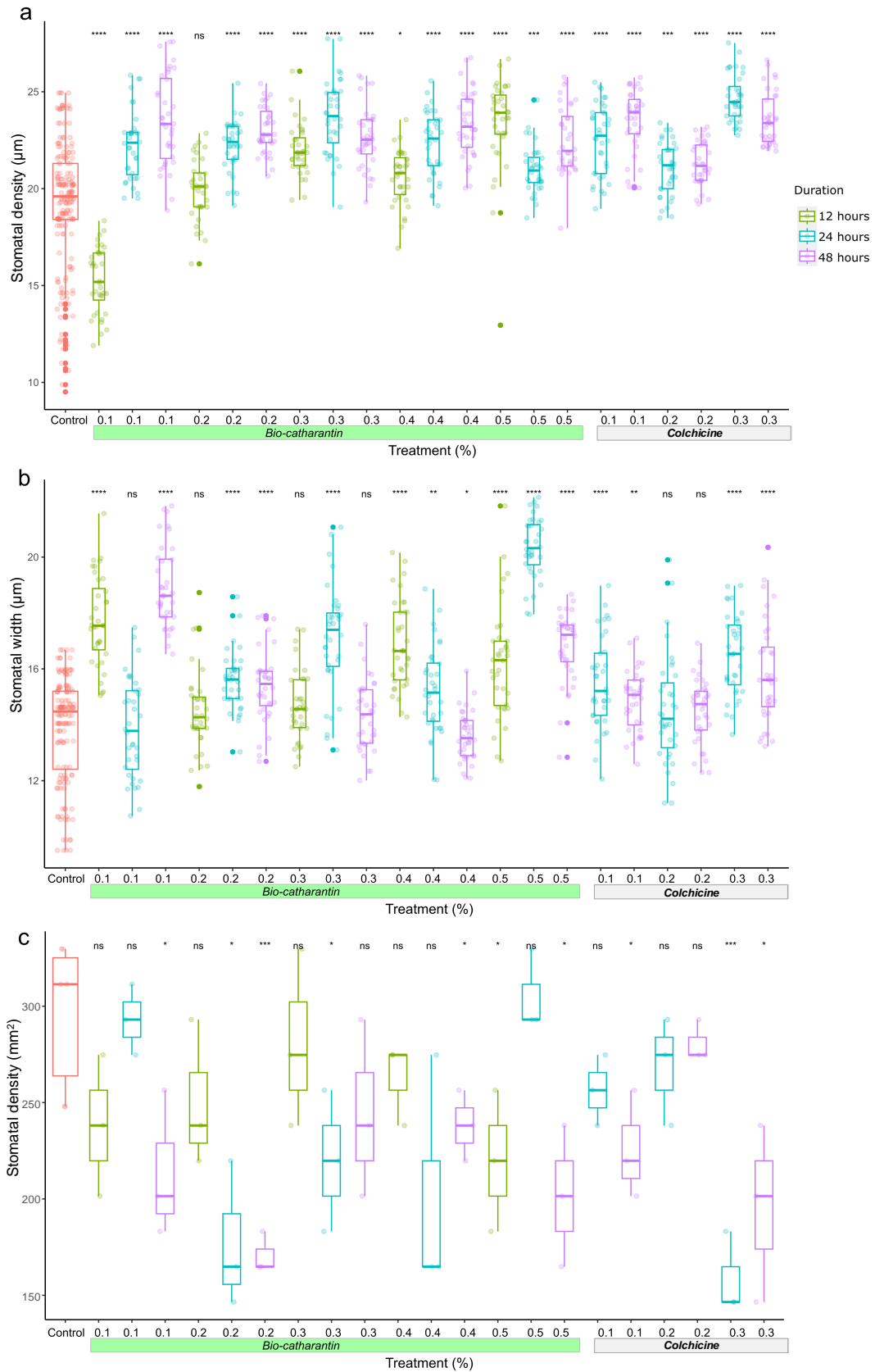


Figure 4. Stomatal structure of *O. sativa* L. var. Jeliteng under Bio-catharantin and colchicine treatment. a) Stomatal length; b) Stomatal width; c) Stomatal density

Note: The box and whisker plots display the distribution across the 25<sup>th</sup> and 75<sup>th</sup> percentile range and median. The jitter plot illustrates the distribution of the data. Significantly different values of the Duncan test ( $p < 0.005$ ) were indicated by different letters. Detailed data were provided in Supplementary Table 2

(Figure 4a). The rest of the higher concentrations in all soaking durations extended the stomatal size (Figure 4a). The majority of treatments also led to an increase in stomatal width, except for 4 cases: 0.1% concentration for 24 hours, 0.2% for 12 hours, and 0.3% for both 12 and 48 hours. The reduction of stomatal density under Bio-catharantin treatments was significantly observed in 8 treatments, i.e., at the concentrations of 0.1% for 24 and 48 hours, 0.2% for 24 and 48 hours, 0.3% for 24 hours, 0.4% for 48 hours, 0.5% for 12 and 48 hours.

Based on research conducted by Wardana et al. (2019), the treatment of 0.1% colchicine for 24 hours in *Zephyranthes rosea* increased leaf area from 9.6 cm<sup>2</sup> in the control group to 13.92 cm<sup>2</sup>. Similarly, a study by Manzoor et al. (2019) on *Gladiolus grandiflorus* Andrews found that immersion in 0.3% colchicine for 24 hours increased the length of stomata from 22.6 to 25.6 µm and width from 18 to 21 µm. There is a variation in the stomatal length and width in the control and treatment combination of the black rice variety Jeliteng. As detailed in Figure 4a and 4b, the treated black rice Jeliteng exhibited longer and wider stomata than the control group. Stomatal size typically influences density, as tissues with smaller cell sizes exhibit higher stomatal density in a given area compared to tissues with larger cells (Bharati et al., 2023; Kurniawan et al., 2023). The stomatal size and density are critical factors of water conductivity (Rathnasamy et al., 2023; Phunthong et al., 2024). The cultivation of rice is water-intensive and vulnerable to drought and high temperatures, both of which are expected to increase in frequency due to climate change. Despite reduced photosynthesis in some cases, the low-stomatal-density rice produced equivalent or even higher yields (Caine et al., 2019). The data of lower stomatal densities suggested that *O. sativa* L. var. Jeliteng induced by the 2 antimetabolic compounds would become the genetic resource for mitigation of the negative impacts of climate change on food security by improving water-use efficiency and drought tolerance.

Genetic changes induced by antimetabolic compounds led to alterations in cellular structure, regardless of whether polyploid individuals were formed. In this study, none of the polyploids occurred, although the antimetabolic compound altered the stomatal size and density, which might relate to the genetic mutation. The stomatal size and density alternation with the absence of

polyploid were reported solely by a genetic mutation, such as in sugarcane with gamma irradiation (Yasmeen et al., 2020), and in rice with overexpression of OsEPF1 (Caine et al., 2019) and CRISPR/Cas9 approach (Rathnasamy et al., 2023). The high genetic diversity in local Indonesian black rice still requires efforts to develop short-lived, high-yielding varieties (Muhamad et al., 2017; Sitaresmi et al., 2023). The research on Cempo Ireng provides a reference for the potential use of non-toxic antimetabolic compounds, which are safe for direct application by farmers in black rice breeding. The stomatal data in this study might contribute to the stomatal diversity and development among rice (Trojak-Goluch and Skomra, 2013; Nofitahesti and Daryono, 2016; Chatterjee et al., 2020; Kurnianingsih et al., 2024). Plant breeding using antimetabolic compounds requires optimization of concentration and soaking duration (Viana et al., 2019; Bhuvaneswari et al., 2020; Chen et al., 2021; Kurniawan et al., 2023). Another approach to enhance the success rate of genetic modification with antimetabolic compounds was *in vitro* methods. With tissue culture technology, somatic cells can be more effectively exposed to antimetabolic compounds (Udall and Wendel, 2006; Eng and Ho, 2018; Chen et al., 2021; Bharati et al., 2023). Bio-catharantin has greater potential compared to colchicine due to its lower toxicity, which does not harm the cells in rice calluses. Moreover, somatic embryo growth tests have been widely reported (Nabilah et al., 2022) so it has potential for further studies with the application of Bio-catharantin.

## CONCLUSIONS

The present study highlights that Bio-catharantin and colchicine have distinct impacts on the germination and genetic characteristics of Jeliteng black rice. While Bio-catharantin did not affect the germination rate, colchicine at a concentration of 0.2% significantly reduced it. Both agents induced changes in the DNA relative content, suggesting their effectiveness in genetic manipulation. Variations in the treatment concentrations and soaking durations influenced the stomatal size and density, with Bio-catharantin uniquely affecting stomatal size without altering density. Importantly, Bio-catharantin emerged as a promising agent for chromosome doubling due to its lack of toxic effects, highlighting its potential for use in plant



breeding programs. This study underscores the viability of Bio-catharantin as a safer alternative to traditional agents like colchicine for inducing polyploidy in crops.

## ACKNOWLEDGEMENT

This research was funded by the Ministry of Education, Culture, Research, and Technology of the Republic of Indonesia through post-doctoral scheme Universitas Jember 2023 (Contract number: 3490/UN25.3/LT/2023).

## REFERENCES

- Bharati, R., Fernández-Cusimamani, E., Gupta, A., Novy, P., Moses, O., Severová, L., Svoboda, R., & Šrédli, K. (2023). Oryzalin induces polyploids with superior morphology and increased levels of essential oil production in *Mentha spicata* L. *Industrial Crops and Products*, 198, 116683. <https://doi.org/10.1016/J.INDCROP.2023.116683>
- Bhuvanewari, G., Thirugnanasampandan, R., & Gogulramnath, M. (2020). Effect of colchicine induced tetraploidy on morphology, cytology, essential oil composition, gene expression and antioxidant activity of *Citrus limon* (L.) Osbeck. *Physiology and Molecular Biology of Plants*, 26(2), 271. <https://doi.org/10.1007/S12298-019-00718-9>
- Billa, A. T., Lestari, S. S., Daryono, B. S., & Subiastuti, A. S. (2022). Bio-catharantin effects on phenotypic traits and chromosome number of shallots (*Allium cepa* L. var. *ascalonicum* 'Tajuk'). *SABRAO Journal of Breeding and Genetics*, 54(2), 350–358. <https://doi.org/10.54910/sabrao2022.54.2.11>
- Caine, R. S., Yin, X., Sloan, J., Harrison, E. L., Mohammed, U., Fulton, T., ... & Gray, J. E. (2019). Rice with reduced stomatal density conserves water and has improved drought tolerance under future climate conditions. *New Phytologist*, 221(1), 371–384. <https://doi.org/10.1111/NPH.15344>
- Chatterjee, J., Thakur, V., Nepomuceno, R., Coe, R. A., Dionora, J., Elmido-Mabilangan, A., ... & Quick, W. P. (2020). Natural diversity in stomatal features of cultivated and wild oryza species. *Rice*, 13(1), 1–20. <https://doi.org/10.1186/S12284-020-00417-0>
- Chen, R., Feng, Z., Zhang, X., Song, Z., & Cai, D. (2021). A new way of rice breeding: Polyploid rice breeding. *Plants*, 10(3), 422. <https://doi.org/10.3390/PLANTS10030422>
- de Mendiburu, F., & de Mendiburu, M. F. (2019). Package 'agricolae'. *R Package, version*, 1(3), 1143–49. Retrieved from [https://scholar.google.co.id/scholar?cites=4519011155554250915&as\\_sdt=2005&scioldt=0,5&hl=id&authuser=3](https://scholar.google.co.id/scholar?cites=4519011155554250915&as_sdt=2005&scioldt=0,5&hl=id&authuser=3)
- Dwiningsih, Y., & Alkahtani, J. (2023). Potential of pigmented rice variety Cempo Ireng in rice breeding program for improving food sustainability. *Journal of Biomedical Research & Environmental Sciences*, 4(3), 426–433. <https://doi.org/10.37871/JBRES1691>
- Eng, W.-H., & Ho, W.-S. (2018). Polyploidization using colchicine in horticultural plants: A review. *Scientia Horticulturae*, 246, 604–617. <https://doi.org/10.1016/j.scienta.2018.11.010>
- Ermayanti, T. M., Wijayanta, A. N., & Ratnadewi, D. (2018). Induksi poliploidi pada tanaman talas (*Colocasia esculenta* (L.) Schott) kultivar Kaliurang dengan perlakuan kolkisin secara *in vitro*. *Jurnal Biologi Indonesia*, 14(1), 91–102. <https://doi.org/10.14203/JBI.V14I1.3667>
- Gaafar, R. M., El Shanshoury, A. R., El Hisseiwy, A. A., AbdAlhak, M. A., Omar, A. F., Abd El Wahab, M. M., & Nofal, R. S. (2017). Induction of apomixis and fixation of heterosis in Egyptian rice Hybrid1 line using colchicine mutagenesis. *Annals of Agricultural Sciences*, 62(1), 51–60. <https://doi.org/10.1016/J.AOAS.2017.03.001>
- Grouh, H. S. M., Meftahizade, H., Lotfi, N., Rahimi, V., & Baniyasi, B. (2011). Doubling the chromosome number of *Salvia hains* using colchicine: Evaluation of morphological traits of recovered plants. *Journal of Medicinal Plants Research*, 5(19), 4892–4898. <https://doi.org/10.5897/JMPR.9000459>
- Hafeez, A., Ali, B., Javed, M. A., Saleem, A., Fatima, M., Fathi, A., ... & Soudy, F. A. (2023). Plant breeding for harmony between sustainable agriculture, the environment, and global food security: An era of genomics-assisted breeding. *Planta*, 258(5), 97. <https://doi.org/10.1007/S00425-023-04252-7>
- Hodač, L., Ulum, F. B., Opfermann, N., Breidenbach, N., Hojsgaard, D., Tjitrosoedirdjo, S. S., ... & Hörandl, E. (2016).

- Population genetic structure and reproductive strategy of the introduced grass *Centotheca lappacea* in tropical land-use systems in Sumatra. *PLoS ONE*, *11*(1), e0147633. <https://doi.org/10.1371/journal.pone.0147633>
- Indonesian Rice Research Center. (2019). *Jeliteng*. Retrieved from <https://padi-bsippid.pertanian.go.id/doc/64/LAKIN%20BB%20PADI%202019.pdf>
- Kasim, N., Sjahril, R., Riadi, M., Syaiful, S. A., Ngatimin, S. N. A., Pratiwi, A., & Anwar, I. (2024). Comparative analysis of colchicine and bio-catharanthine as mutagenic agents for polyploid generation in wild passion fruit (*Passiflora foetida*). *Australian Journal of Crop Science*, *18*(06), 334–341. <https://doi.org/10.21475/ajcs.24.18.06.pne53>
- Kurnianingsih, N., Safitri, A., Septianingrum, E., Ardhiyanti, S. D., & Fatchiyah, F. (2024). Nutritional profiling of Indonesian superior hybrid and biofortification rice varieties: Macronutrient and micronutrient changes under different heat temperatures. *Berkala Penelitian Hayati*, *30*(1), 15–20. <https://doi.org/10.23869/bphjbr.30.1.20243>
- Kurniawan, L., Laili, A. N., Anggraeni, D. S., Qurrotu'ain, S., Wulandari, D. R., & Ulum, F. B. (2023). Polyploidy induction of Indonesian black rice *Oryza sativa* L. Var. Cempo Ireng with Bio-catharantine. *Life Science and Biotechnology*, *1*(2), 41–47. <https://doi.org/10.19184/lb.v1i2.43753>
- Luo, L., Han, B., Yu, X., Chen, X., Zhou, J., Chen, W., ... & Li, S. (2014). Anti-metastasis activity of black rice anthocyanins against breast cancer: Analyses using an ErbB2 positive breast cancer cell line and tumoral xenograft model. *Asian Pacific Journal of Cancer Prevention*, *15*(15), 6219–6225. <https://doi.org/10.7314/APJCP.2014.15.15.6219>
- Ly, Z., Zhu, F., Jin, D., Wu, Y., & Wang, S. (2021). Seed germination and seedling growth of *Dendrocalumus brandisii* *in vitro*, and the inhibitory mechanism of colchicine. *Frontiers in Plant Science*, *12*, 784581. <https://doi.org/10.3389/fpls.2021.784581>
- Manzoor, A., Ahmad, T., Bashir, M. A., Hafiz, I. A., & Silvestri, C. (2019). Studies on colchicine induced chromosome doubling for enhancement of quality traits in ornamental plants. *Plants*, *8*(7), 194. <https://doi.org/10.3390/PLANTS8070194>
- Moghbel, N., Borujeni, M. K., & Bernard, F. (2015). Colchicine effect on the DNA content and stomata size of *Glycyrrhiza glabra* var. glandulifera and *Carthamus tinctorius* L. cultured *in vitro*. *Journal of Genetic Engineering & Biotechnology*, *13*(1), 1–6. <https://doi.org/10.1016/J.JGEB.2015.02.002>
- Muarifin, A., Perdamaian, A. B. I., Sartika, D., & Daryono, B. S. (2021). Induced polyploidy in *Arachis hypogaea* L. var. Talam using *Catharanthus roseus* phenolic extract. *Asian Journal of Plant Sciences*, *20*(2), 263–270. <https://doi.org/10.3923/AJPS.2021.263.270>
- Muhamad, K., Ebana, K., Fukuoka, S., & Okuno, K. (2017). Genetic relationships among improved varieties of rice (*Oryza sativa* L.) in Indonesia over the last 60 years as revealed by morphological traits and DNA markers. *Genetic Resources and Crop Evolution*, *64*(4), 701–715. <https://doi.org/10.1007/s10722-016-0392-1>
- Nabilah, S., Handoyo, T., Kim, K., & Ubaidillah, M. (2022). Expression analysis of *OsSERK*, *OsLECI* and *OsWOX4* genes in rice (*Oryza sativa* L.) callus during somatic embryo development. *BIOCELL*, *46*(7), 1633–1641. <https://doi.org/10.32604/BIOCELL.2022.019111>
- Nandariyah, N., Sukaya, S., Purnomo, D., Sutarno, S., Yuniastuti, E., & Az-Zahra, C. D. A. (2023). Study of black rice parents performance and the crossing ability. *Caraka Tani: Journal of Sustainable Agriculture*, *38*(1), 65–74. <https://doi.org/10.20961/carakatani.v38i1.60245>
- Nandariyah, Yuniastuti, E., Purwanto, E., & Astuti, R. D. (2022). Potential lines of black rice crossing with Jeliteng variety and their reciprocals. *IOP Conference Series: Earth and Environmental Science*, *1016*(1), 012017. <https://doi.org/10.1088/1755-1315/1016/1/012017>
- Nofitahesti, I., & Daryono, B. S. (2016). Karakter fenotip kedelai (*Glycine max* (L.) Merr.) hasil poliploidisasi dengan kolkisin. *Scientiae Educatia: Jurnal Pendidikan Sains*, *5*(2), 90–98. <https://doi.org/10.24235/sc.educatia.v5i2.957>

- Nurhidajah, N., Yonata, D., Bintanah, S., & Pranata, B. (2024). Physicochemical and structural composition of black rice (*Oryza sativa*) flour from Java, Indonesia. *Biodiversitas Journal of Biological Diversity*, 25(2), 811–818. <https://doi.org/10.13057/biodiv/d250241>
- Paiman, Ardiyanto, Ansar, M., Effendy, I., & Sumbodo, B. T. (2020). Rice cultivation of superior variety in swamps to increase food security in Indonesia. *Reviews in Agricultural Science*, 8, 300–309. [http://dx.doi.org/10.7831/ras.8.0\\_300](http://dx.doi.org/10.7831/ras.8.0_300)
- Phunthong, C., Pitaloka, M. K., Chutteang, C., Ruengphayak, S., Arikrit, S., & Vanavichit, A. (2024). Rice mutants, selected under severe drought stress, show reduced stomatal density and improved water use efficiency under restricted water conditions. *Frontiers in Plant Science*, 15, 1307653. <https://doi.org/10.3389/FPLS.2024.1307653>
- Prasetyo, F. H. H., Sugiharto, B., & Ermawati, N. (2018). Cloning, transformation and expression of cell cycle-associated protein kinase OsWeel in indica rice (*Oryza sativa* L.). *Journal of Genetic Engineering and Biotechnology*, 16(2), 573–579. <https://doi.org/10.1016/J.JGEB.2018.10.003>
- Rathnasamy, S. A., Kambale, R., Elangovan, A., Mohanavel, W., Shanmugavel, P., Ramasamy, G., ... & Vellingiri, G. (2023). Altering stomatal density for manipulating transpiration and photosynthetic traits in rice through CRISPR/Cas9 mutagenesis. *Current Issues in Molecular Biology*, 45(5), 3801–3814. <https://doi.org/10.3390/cimb45050245>
- Rohmah, A., Rahayu, T., & Hayati, A. (2017). Pengaruh pemberian kolkisin terhadap karakter stomata daun zaitun (*Olea europaeae* L.). *Jurnal Ilmiah Biosaintropis (Bioscience-Tropic)*, 2(2), 10–17. <https://doi.org/10.33474/E-JBST.V2I2.81>
- Samadi, N., Naghavi, M. R., Moratalla-López, N., Alonso, G. L., & Shokrpour, M. (2022). Morphological, molecular and phytochemical variations induced by colchicine and EMS chemical mutagens in *Crocus sativus* L. *Food Chemistry: Molecular Sciences*, 4, 100086. <https://doi.org/10.1016/J.FOCHMS.2022.100086>
- Shafura, N., Janah, L. N., Huda, M. S., & Daryono, B. S. (2022). Effectiveness of Bio-Catharantin induction to increase red spinach (*Alternanthera amoena* Voss.) production. *Proceedings of the 7th International Conference on Biological Science (ICBS 2021)*, 22, 528–532. <https://doi.org/10.2991/ABSR.K.220406.074>
- Sitairesmi, T., Hairmansis, A., Widyastuti, Y., Rachmawati, Susanto, U., Wibowo, B. P., ... & Nugraha, Y. (2023). Advances in the development of rice varieties with better nutritional quality in Indonesia. *Journal of Agriculture and Food Research*, 12, 100602. <https://doi.org/10.1016/J.JAFR.2023.100602>
- Suryanti, V., Riyatun, Suharyana, Sutarno, & Saputra, O. A. (2020). Antioxidant activity and compound constituents of gamma-irradiated black rice (*Oryza sativa* L.) var. Cempo Ireng indigenous of Indonesia. *Biodiversitas*, 21(9), 4205–4212. <https://doi.org/10.13057/biodiv/d210935>
- Thepthanee, C., Liu, C.-C., Yu, H.-S., Huang, H.-S., Yen, C.-H., Li, Y.-H., Lee, M.-R., & Liaw, E.-T. (2021). Evaluation of phytochemical contents and *in vitro* antioxidant, anti-inflammatory, and anticancer activities of black rice leaf (*Oryza sativa* L.) extract and its fractions. *Foods*, 10(12), 2987. <https://doi.org/10.3390/foods10122987>
- Tossi, V. E., Martínez Tosar, L. J., Laino, L. E., Iannicelli, J., Regalado, J. J., Escandón, A. S., ... & Pitta-Álvarez, S. I. (2022). Impact of polyploidy on plant tolerance to abiotic and biotic stresses. *Frontiers in Plant Science*, 13, 869423. <https://doi.org/10.3389/fpls.2022.869423>
- Trojak-Goluch, A., & Skomra, U. (2013). Artificially induced polyploidization in *Humulus lupulus* L. and its effect on morphological and chemical traits. *Breeding Science*, 63(4), 393–399. <https://doi.org/10.1270/JSBBS.63.393>
- Tyagi, A., Shabbir, U., Chen, X., Chelliah, R., Elahi, F., Ham, H. J., & Oh, D. H. (2022). Phytochemical profiling and cellular antioxidant efficacy of different rice varieties in colorectal adenocarcinoma cells exposed to oxidative stress. *PLoS ONE*, 17(6), e0269403.

- <https://doi.org/10.1371/JOURNAL.PONE.0269403>
- Udall, J. A., & Wendel, J. F. (2006). Polyploidy and crop improvement. *Crop Science*, *46*, S-3-S-14. <https://doi.org/10.2135/CROPSCI2006.07.0489TPG>
- Viana, V. E., Pegoraro, C., Busanello, C., & Costa de Oliveira, A. (2019). Mutagenesis in rice: The basis for breeding a new super plant. *Frontiers in Plant Science*, *10*, 419616. <https://doi.org/10.3389/FPLS.2019.01326/BIBTEX>
- Wardana, Slamet, A., Andarias, S. H., Bahrn, A. H., Mantja, K., & Darwis. (2019). Induction of Lili Hujan polyploid (*Zephyranthes rosea* Lindl.) with ethanolic extract of Tapak Dara leaf (*Catharanthus roseus* (L.) G. don.) to increase its economic value. *IOP Conference Series: Earth and Environmental Science*, *235*(1), 012102. <https://doi.org/10.1088/1755-1315/235/1/012102>
- Wickham, H., & Wickham, H. (2016). *Getting started with ggplot2*. ggplot2: Elegant graphics for data analysis, pp.11-31. [https://doi.org/10.1007/978-3-319-24277-4\\_2](https://doi.org/10.1007/978-3-319-24277-4_2)
- Wijayanti, E. D., Safitri, A., Siswanto, D., & Fatchiyah, F. (2023). Indonesian purple rice ferulic acid as a candidate for anti-aging through the inhibition of collagenase and tyrosinase activities. *Indonesian Journal of Chemistry*, *23*(2), 475–488. <https://doi.org/10.22146/ijc.79819>
- Wu, J., Cheng, X., Kong, B., Zhou, Q., Sang, Y., & Zhang, P. (2022). *In vitro* octaploid induction of *Populus hopeiensis* with colchicine. *BMC Plant Biology*, *22*(1), 176. <https://doi.org/10.1186/s12870-022-03571-3>
- Xia, D., Zhou, H., Wang, Y., Li, P., Fu, P., Wu, B., & He, Y. (2021). How rice organs are colored: The genetic basis of anthocyanin biosynthesis in rice. *The Crop Journal*, *9*(3), 598–608. <https://doi.org/10.1016/J.CJ.2021.03.013>
- Yao, P. Q., Chen, J. H., Ma, P. F., Xie, L. H., & Cheng, S. P. (2023). Stomata variation in the process of polyploidization in Chinese chive (*Allium tuberosum*). *BMC Plant Biology*, *23*(1), 595. <https://doi.org/10.1186/s12870-023-04615-y>
- Yasmeen, S., Khan, M. T., & Khan, I. A. (2020). Revisiting the physical mutagenesis for sugarcane improvement: A stomatal prospective. *Scientific Reports*, *10*(1), 1–14. <https://doi.org/10.1038/s41598-020-73087-z>

**Appendices**

Table 1. Summary statistic values of germination rate of *O. sativa* L. var. Jeliteng after colchicine and Bio-catharantin treatment

Treatment		N	Mean	SD	Median	Min	Max
Concentration (%)	Duration (Hour)						
Bio-catharantine							
0.1	12	3	91.00	1.73	90.00	90.00	93.00
	24	3	92.33	4.04	90.00	90.00	97.00
	48	3	84.33	2.31	83.00	83.00	87.00
0.2	12	3	89.00	1.73	90.00	87.00	90.00
	24	3	92.33	5.03	93.00	87.00	97.00
	48	3	84.33	2.31	83.00	83.00	87.00
0.3	12	3	83.33	3.51	83.00	80.00	87.00
	24	3	93.33	3.51	93.00	90.00	97.00
	48	3	92.00	1.73	93.00	90.00	93.00
0.4	12	3	92.00	1.73	93.00	90.00	93.00
	24	3	90.00	3.00	90.00	87.00	93.00
	48	3	84.33	2.31	83.00	83.00	87.00
0.5	12	3	90.00	3.00	90.00	87.00	93.00
	24	3	89.00	8.54	90.00	80.00	97.00
	48	3	86.67	3.51	87.00	83.00	90.00
Colchicine							
0.1	24	4	90.00	4.71	88.33	86.67	96.67
	48	4	81.67	4.30	81.67	76.67	86.67
0.2	24	4	75.83	9.95	76.67	63.33	86.67
	48	4	58.33	8.82	60.00	46.67	66.67
0.3	24	4	32.50	9.95	33.33	20.00	43.33
	48	4	39.17	6.87	38.33	33.33	46.67
Control		7	91.29	8.34	93.33	80.00	100.00

Note: N = 150 seeds

Table 2. Summary statistic values of stomatal size and density of *O. sativa* L. var. Jeliteng after colchicine and Bio-catharantin treatment

Data	Concentration (%)	Soaking duration (hr)	N	Mean	SD	Median	Min	Max	CV
Length	Control		108	18.32	3.70	19.06	9.51	24.94	20.18
	Bio-catharantin	0.1	12	36	15.29	1.69	15.19	11.90	18.34
24			36	22.26	1.77	22.37	19.50	25.86	7.96
48			36	23.52	2.35	23.34	18.88	27.58	9.99
0.2		12	36	19.98	1.55	20.11	16.12	22.86	7.75
		24	36	22.23	1.33	22.41	19.13	25.44	5.98
		48	36	23.03	1.20	22.79	20.64	25.43	5.19
0.3		12	36	21.99	1.30	21.86	19.41	26.06	5.92
		24	36	23.73	1.92	23.75	19.05	27.73	8.09
		48	36	22.62	1.60	22.53	19.31	25.83	7.06
0.4	12	36	20.59	1.42	20.81	16.92	23.56	6.87	
	24	36	22.35	1.64	22.59	19.12	25.56	7.32	
	48	36	23.37	1.70	23.20	20.03	26.76	7.27	
0.5	12	36	23.36	2.48	23.92	12.95	26.69	10.63	
	24	36	20.99	1.22	20.94	18.49	24.58	5.83	
	48	36	22.49	1.73	21.95	17.96	25.76	7.71	

	Colchicine	0.1	24	36	22.54	1.84	22.73	18.96	25.49	9.38
			48	36	23.52	1.59	23.94	20.04	25.74	8.15
		0.2	24	36	21.04	1.37	21.21	18.49	23.40	6.74
			48	36	21.29	1.10	21.18	19.21	23.19	6.50
		0.3	24	36	24.68	1.20	24.47	22.75	27.52	5.14
			48	36	23.58	1.35	23.39	21.95	26.64	4.87
Width	Control			72	13.87	1.89	14.48	9.51	16.68	5.73
	Bio-catharantin	0.1	12	36	17.71	1.59	17.55	15.05	21.56	13.61
24			36	13.87	1.78	13.78	10.74	17.48	8.96	
48			36	18.93	1.45	18.62	16.53	21.83	12.81	
0.2		12	36	14.47	1.45	14.27	11.79	18.73	7.68	
		24	36	15.59	1.06	15.62	13.03	18.58	10.00	
		48	36	15.39	1.29	15.46	12.69	17.90	6.79	
0.3		12	36	14.68	1.24	14.56	12.50	17.42	8.36	
		24	36	17.10	1.92	17.40	13.10	21.07	8.42	
		48	36	14.34	1.29	14.38	12.01	17.59	11.22	
0.4		12	36	16.86	1.55	16.64	14.28	20.16	9.02	
		24	36	15.21	1.56	15.15	12.03	18.86	9.22	
		48	36	13.60	0.87	13.52	12.09	15.92	10.28	
0.5		12	36	16.21	1.97	16.31	12.71	21.83	6.41	
		24	36	20.34	1.09	20.32	17.95	22.14	12.13	
		48	36	16.79	1.26	17.22	12.84	18.67	5.38	
Colchicine		0.1	24	36	15.52	1.63	15.21	12.06	18.98	7.52
			48	36	14.86	1.08	15.07	12.59	17.10	13.61
		0.2	24	36	14.51	1.97	14.22	11.20	19.90	10.52
	48		36	14.50	1.13	14.74	12.29	16.92	7.24	
	0.3	24	36	16.50	1.44	16.54	13.65	18.98	13.56	
		48	36	15.81	1.81	15.61	13.23	20.35	7.81	
Density	Control			3	296.34	42.86	311.36	247.99	329.67	8.73
	Bio-catharantin	0.1	12	3	238.10	36.63	238.10	201.47	274.73	11.45
24			3	293.04	18.32	293.04	274.73	311.36	14.46	
48			3	213.68	38.13	201.47	183.15	256.41	15.38	
0.2		12	3	250.31	38.13	238.10	219.78	293.04	6.25	
		24	3	177.05	38.13	164.84	146.52	219.78	17.84	
		48	3	170.94	10.57	164.84	164.84	183.15	15.23	
0.3		12	3	280.83	46.09	274.73	238.10	329.67	21.53	
		24	3	219.78	36.63	219.78	183.15	256.41	6.19	
		48	3	244.20	46.09	238.10	201.47	293.04	16.41	
0.4		12	3	262.52	21.15	274.73	238.10	274.73	16.67	
		24	3	201.47	63.45	164.84	164.84	274.73	18.87	
		48	3	238.10	18.32	238.10	219.78	256.41	8.06	
0.5		12	3	219.78	36.63	219.78	183.15	256.41	31.49	
		24	3	305.25	21.15	293.04	293.04	329.67	7.69	
		48	3	201.47	36.63	201.47	164.84	238.10	16.67	
Colchicine		0.1	24	3	256.41	18.32	256.41	238.10	274.73	6.93
			48	3	225.89	27.97	219.78	201.47	256.41	18.18
		0.2	24	3	268.62	27.97	274.73	238.10	293.04	14.46
	48		3	280.83	10.57	274.73	274.73	293.04	7.14	
	0.3	24	3	158.73	21.15	146.52	146.52	183.15	12.38	
		48	3	195.36	46.09	201.47	146.52	238.10	10.41	

Note: N = 3-5 individual