



## Assessing N<sub>2</sub>O Emissions from Tropical Crop Cultivation in Mineral and Peatland Soils: A Review

Suardi<sup>1,3\*</sup>, Darmawan<sup>1</sup>, Gunawan Djajakirana<sup>1,3</sup>, Basuki Sumawinata<sup>1</sup> and Nourma Al Viandari<sup>2</sup>

<sup>1</sup>Department of Soil Science and Land Resources, Faculty of Agriculture, IPB University, Bogor, Indonesia;

<sup>2</sup>Research Center for Food Crops, Research Organization for Agriculture and Food, National Research and Innovation Agency (BRIN), Bogor, Indonesia; <sup>3</sup>Center for Mine Reclamation Studies, Institute of Research and Community Service, IPB University, Bogor; Indonesia

\*Corresponding author: [suardi-soil@apps.ipb.ac.id](mailto:suardi-soil@apps.ipb.ac.id)

### Abstract

Nitrous oxide (N<sub>2</sub>O) emissions from agricultural activities contribute significantly to global warming. Understanding the factors influencing N<sub>2</sub>O emissions is crucial for developing effective mitigation strategies. This review assesses N<sub>2</sub>O emissions from various crops cultivated in tropical mineral and peatland soils, providing insights into the impact of land use, fertilization practices and rainfall on N<sub>2</sub>O fluxes. Field measurements of N<sub>2</sub>O fluxes were conducted in agricultural fields growing corn, peanuts, and cassava in Bogor Regency, West Java Province, as well as in peatland areas with Acacia plantations and natural primary forests in Bengkalis Regency, Riau Province. The study assesses the total N<sub>2</sub>O fluxes for each crop and land type, revealing significant variations in N<sub>2</sub>O emissions among different crops and land uses. Peatland areas exhibit higher emissions compared to mineral soils, emphasizing the need for targeted mitigation measures in these ecosystems. The findings highlight the importance of considering the type and age of land use when evaluating N<sub>2</sub>O emissions. Land management practices, such as fertilizer use and soil disturbance, emerge as critical factors affecting N<sub>2</sub>O emissions. Improper fertilizer application and excessive soil disturbance can lead to increased N<sub>2</sub>O emissions, underscoring the necessity for careful N fertilizer management and conservation tillage techniques.

**Keywords:** agricultural land; closed chamber method; N<sub>2</sub>O gas emission; peatland soil; tropical mineral soil

**Cite this as:** Suardi, Darmawan, Djajakirana, G., Sumawinata, B., & Al Viandari, N. (2023). Assessing N<sub>2</sub>O Emissions from Tropical Crop Cultivation in Mineral and Peatland Soils: A Review. *Caraka Tani: Journal of Sustainable Agriculture*, 38(2), 308-326. doi: <http://dx.doi.org/10.20961/carakatani.v38i2.75235>

### INTRODUCTION

Agricultural activities are widely known to heavily influence the emission of greenhouse gases (GHGs) namely CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O into the atmosphere (Kweku et al., 2018). Plant activity can increase CO<sub>2</sub> emission from soil (Yerli et al., 2022). Although CO<sub>2</sub> is by far the most abundant GHG, N<sub>2</sub>O is also important because of its unique radiative properties and long residence time in the atmosphere (Hodnebrog et al., 2020). N<sub>2</sub>O molecules remain in the atmosphere for

around 114 years before being extracted through a chemical reaction. It is a different N<sub>2</sub>O lifespan than CO<sub>2</sub>; molecules of CO<sub>2</sub> hang in the atmosphere for around 5 to 200 years. Nevertheless, no single lifespan can be expounded for CO<sub>2</sub> because of the different rates of removal processes (IPCC, 2022a). N<sub>2</sub>O is also resulting in a global warming potential (GWP) of around 296 times that of CO<sub>2</sub> (Hodnebrog et al., 2020). Specifically, one pound of N<sub>2</sub>O in the atmosphere is 296 times that of one pound of CO<sub>2</sub>. In line with this, it was also stated that N<sub>2</sub>O has a shorter

\* Received for publication June 19, 2023  
Accepted after corrections July 17, 2023

atmospheric lifetime of approximately 110 years compared to CO<sub>2</sub>, which has a lifetime of around 120 years (Ritchie et al., 2020). The impact of this GHG is defined as “the ratio of the time-integrated radiative forcing from the instantaneous release of 1 kg of a trace substance relative to that of 1 kg of the reference gas, CO<sub>2</sub>” (Hsiao, 2022).

According to the latest information from the Intergovernmental Panel on Climate Change (IPCC), N<sub>2</sub>O is a GHG with a global warming potential nearly 310 times that of CO<sub>2</sub> within 100 years (IPCC, 2022b). N<sub>2</sub>O has a GWP of 273 times that of CO<sub>2</sub> for a 100-year timescale and a lifetime of 110 to 132 years (Prather et al., 2015). The global concentration of N<sub>2</sub>O has increased from 270 ppb to 319 ppb in the last decade, with approximately 80% of the world's total N<sub>2</sub>O emissions related to agricultural activities (IPCC, 2023).

In recent years, GWP has been used to evaluate the effect of agricultural activities on global warming (Costa et al., 2021). Nugroho et al. (2018) stated that the difference in soil properties such as soil texture, drainage, water-filled pore space (WFPS), and C/N ratio probably cause the magnitude of CO<sub>2</sub> emission. CO<sub>2</sub> flux is affected by environmental factors such as temperature, moisture content and humidity of the soil, as well as the type and extent of vegetation cover (Hendri et al., 2015; Busman et al., 2023). It is essential to understand the factors that contribute to N<sub>2</sub>O emissions from different crops and soil types to develop effective strategies for mitigating N<sub>2</sub>O emissions. The identification of crops and soil types that produce lower levels of N<sub>2</sub>O can help to reduce the impact of agricultural activities on the environment. Moreover, it is crucial to identify management practices that can help to reduce N<sub>2</sub>O emissions from crops grown in tropical mineral and peatland soil. The factors that may influence N<sub>2</sub>O emission include soil type and management characteristics (Soares et al., 2023).

Mineral and peatland soil in Indonesia mostly is utilized for agricultural cultivation (Gunawan, 2018; Uda et al., 2020; Ahmad, 2021). Therefore, the various agricultural activities in both mineral and peatland soil can contribute to N<sub>2</sub>O emissions. In mineral soil, common agricultural activities include the use of synthetic fertilizers, tillage and irrigation, which can all increase N<sub>2</sub>O emissions (C. Wang et al., 2021; Hassan et al., 2022). In contrast, in peatland soil, agricultural activities such as drainage and land conversion can significantly increase N<sub>2</sub>O emissions (Buschmann

et al., 2020). Excessive drainage of peatlands can create large negative impacts on the climate in the tropical regions. In addition, drainage (groundwater level  $\geq 0.5$  m) caused a larger than 60 times increase in N<sub>2</sub>O than under shallow groundwater conditions (Prananto et al., 2020). Drainage reduces soil water content and leads to an increase in O<sub>2</sub> availability, which can stimulate N<sub>2</sub>O producing microbial activity (C. Wang et al., 2021). Land conversion from natural peatland forests to agricultural land can also increase N<sub>2</sub>O emissions due to changes in soil physical and chemical properties, as well as changes in microbial communities (Arai et al., 2014; Dhandapani et al., 2023).

Given the above premise, the key objective of this study was to estimate the total flux of N<sub>2</sub>O from tropical land with mineral soil that is planted to corn, peanut and cassava crops. In addition, it also determines the total N<sub>2</sub>O flux from tropical soil with peatlands planted with *Acacia crassicarpa* and primary forests of various ages. N<sub>2</sub>O emission can be mitigated appropriately if the causal factors can be identified accurately, especially in the agricultural sector. Thus, this review article discusses the mechanism of N<sub>2</sub>O formation and factors that cause N<sub>2</sub>O emissions from various agricultural cultivation and soil types. At the same time, recognizing the factors that affect the flux of N<sub>2</sub>O from agricultural land, this study can be used as a preliminary study for developing practical methods to reduce N<sub>2</sub>O emissions, while optimizing fertilizer efficiency at field study.

Sustainable agricultural practices are crucial for reducing the environmental impact of agricultural activities, especially in developing countries (Hassan et al., 2022). In support of sustainable agricultural systems, this review highlights the significance of understanding the mechanisms of N<sub>2</sub>O formation and the factors contributing to N<sub>2</sub>O emissions in different agricultural and soil types. By identifying crops and soil management practices that lead to lower N<sub>2</sub>O emissions, it becomes possible to reduce the environmental impact of agricultural activities. This knowledge is particularly valuable for developing countries, where agriculture plays a crucial role in their economies and environmental sustainability (Lawrence et al., 2021). By gaining a deeper understanding of the factors contributing to N<sub>2</sub>O emissions, particularly in the context of tropical crop cultivation on mineral and peatland soils, stakeholders such as governments and farmers

can develop informed strategies and policies for the application of chemical fertilizers and agricultural management. Hence, it will extremely support sustainable agriculture.

## MATERIALS AND METHOD

Through desk study, various in-depth sources such as government reports from 2015 to 2023, peer-reviewed papers and other supporting data were particularly collected to complete this review article. Researchers methodically searched peer-reviewed papers that were published from 2020 until 2023 with the following keywords: “nitrous oxide” or  $N_2O$ ”, “legume”, “peanut”, “cassava”, “maize”, “tropical soil” and “peatland soil”. To obtain credible data on  $N_2O$  flux patterns, the following further criteria were adopted: only annually measured in situ fluxes were included and the samples were collected using the closed Chamber method.

This paper also integrated the review with primary data to get accurate data. For mineral soil, the field experiment was conducted in an agricultural field where corn (*Zea mays*), peanut (*Arachis hypogaea*), and cassava (*Manihot esculenta*) have been cultivated on a rotation basis for several decades. The field was located in Bantar Kambing Village, Bogor Regency, West Java Province, Indonesia (106°43'38.42" E, 6°32'12.36" S) (Figure 1). In this study, all crops were planted in November 2010. The growing period (from planting to harvesting) for corn and peanuts is about 2 to 3 months, but cassava requires a much longer growing period, which is around 7 to 9 months. Hence, the site observations were done at three different

ages for the cassava field: 0 to 3 months, 3 to 6 months, and 6 to 9 months, whereas for corn and peanut, measurements were undertaken for each growing period. Moreover, to determine the flux of  $N_2O$  it was necessary to secure similar environmental conditions, particularly microclimatic conditions (e.g., weather, rainfall). This study fell under the November 2010 to February 2011 growing season. The microclimatic description of the site can be summarized as follows: mean annual precipitation 2,500 to 5,000 mm year<sup>-1</sup>, mean annual temperature 20 to 30 °C. The soil is classified as *Aquic dystrudept* with clay textural class.

The peatland soil field experiment was conducted in Bukit Batu Hutani Alam Company (BBHA), Bengkalis Regency, Riau Province, Indonesia (101°47'42.30" E, 1°29'10.52" N) (Figure 1). The land use type is natural primary forest, which since 2009 has been designated as a UNESCO's Man and the Biosphere Programme (MAB) biosphere reserve and *A. crassiparva* with different ages (1 and 3 years) and natural primary forest and *A. crassiparva* 3 years R-L that means (without fine root and litter). The micro-climatic description of the site peatland can be summarized as: mean annual precipitation 809 to 4,078 mm year<sup>-1</sup>, mean annual temperature 26 to 32 °C. The soil is classified as a deep peatland of more than 3 m and could be classified as fibric (Sumawinata et al., 2014).

## Gas sampling and analysis

$N_2O$  fluxes were measured every week from planting until harvesting in crop fields on mineral soils. Measurements of gas at each site were always conducted between 07:00 a.m. and

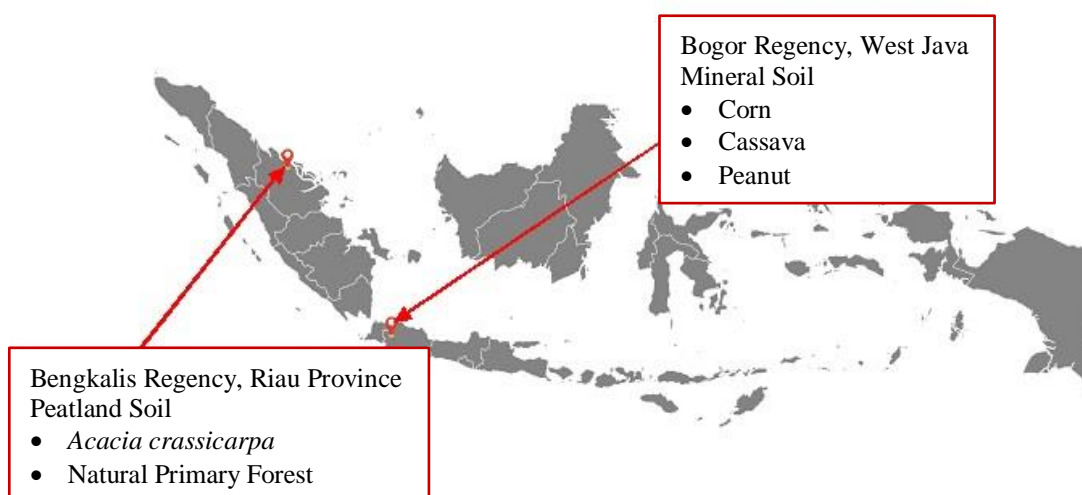


Figure 1. Location of the study sites in Riau and West Java Province

1:00 p.m. to minimize the diurnal variation in the flux pattern. Earlier studies had shown that diurnal  $N_2O$  production varies with diurnal soil temperature, which is at its lowest at sunrise (Reeves and Wang, 2015) and peaks in the late afternoon. The collection of gas  $N_2O$  samples was conducted monthly in peatland soil. Gas samples were collected during the daytime (from about 09:00 a.m. to almost 06:00 p.m.) when the site vegetation was likely to be most physiologically active.

Gas samples were collected using the Closed Chamber Method. The 30 cm x 30 cm x 30 cm chambers were each made of acrylic material, lined with UV paper, and equipped with a fan and a thermometer. During measurement, the chamber was placed on a metal chamber base. The chamber base was planted in the sample plot and was left undisturbed during the sample collection period. In each measurement plot, three chambers were placed to serve as replication. Further, the chamber base was installed in such a way as to eliminate or minimize the effects of fine roots and litter (treatment: -R-L).

The scheme of gas sample collection and the apparatus used are illustrated in Figure 2. The collection of gas samples was conducted in a time-series pattern, that is, at 10-, 20-, and 30-minute intervals for determining  $N_2O$ . For  $N_2O$  determination, as much as 10 ml gas samples were infused into vial bottles, and subsequently analyzed using Gas Chromatography Shimadzu 14B with detector Electron Capture Detector to quantify  $N_2O$  in the laboratory at Hokkaido

University, Japan. The environmental factors that affect the production and release of the gases in the field, such as air temperature, soil temperature, and air humidity, were also recorded (Sumawinata et al., 2014).

The  $N_2O$  flux was calculated as the difference between ambient concentration and the concentration in the closed chamber, assuming a linear relationship between concentration and time (Rapson and Dacres, 2014) which was occasionally checked during the experiment. The accumulated fluxes were calculated by linear interpolation between measurement days. Seasonal total amounts of  $N_2O$  emission were sequentially accumulated from the emissions between pairs of consecutive measurement intervals.

In addition to gas sampling, the environmental factor that affect  $N_2O$  flux such as air temperature, soil temperature, relative humidity, and soil samples for chemical and physical analysis were also recorded both mineral and peatland soil. Air temperature and relative humidity were measured using a thermo-hygrometer, while portable temperature probes were used to determine soil temperature at a depth of 5 cm. Soil samples were collected using a soil sampling ring.

## RESULTS AND DISCUSSION

### $N_2O$ gasses formed

The emission of  $N_2O$  occurs naturally through many sources associated with the N cycle, which is the natural circulation of N between the atmosphere, plants, animals, and microorganisms

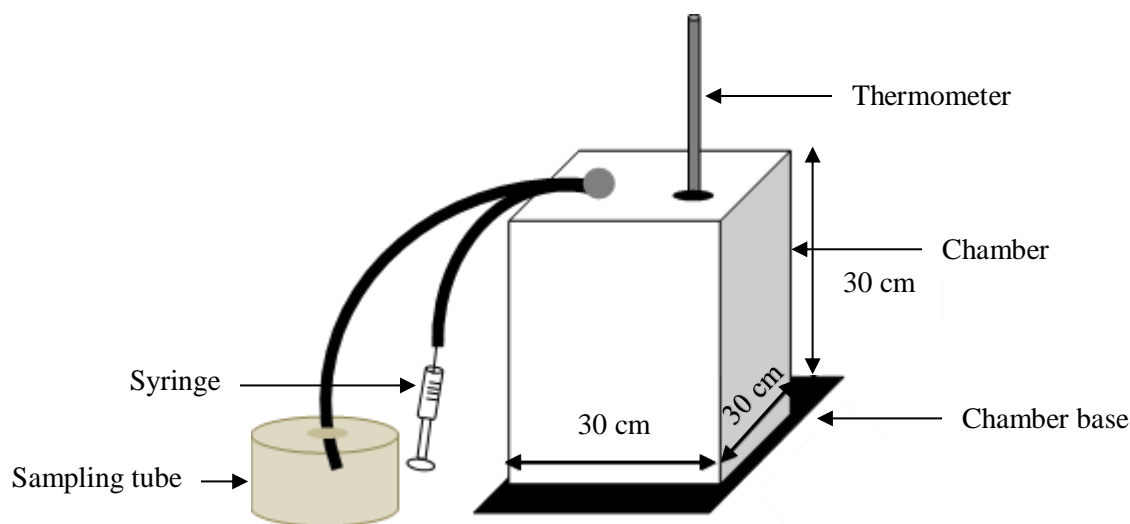


Figure 2. Measurement of  $N_2O$  flux using Closed Chamber Method

that live in soil and water (Delgado and Follett, 2002; Harris et al., 2022). Nitrogen takes various chemical forms throughout the N cycle, including  $N_2O$  (Martínez-Espinosa et al., 2023). Natural emissions of  $N_2O$  comes primarily from bacteria that analyze N in the soil and oceans (Richards et al., 2016; Ito et al., 2018). Bacteria play an important role in the  $N_2$  cycle in the atmosphere that also influence the formation of  $N_2O$ . Ammonia-oxidizing bacteria (AOB) and nitrite-oxidizing bacteria (NOB) lead autotrophic nitrification process. Nitrification initiates with a two-step process. At the first step,  $NH_3$  is oxidized to  $NO_2^-$  by AOB, followed by the oxidation of  $NO_2^-$  to  $NO_3^-$  stimulated by NOB. The process was then followed by anaerobic denitrification stimulated by denitrifying bacteria (Rajta et al., 2020). Almost all  $N_2O$  is formed because of nitrification and denitrification processes in the soil (Figure 3).

$N_2O$  production in the soil primarily occurs through microbial nitrification and denitrification processes. Nitrification involves the oxidation  $NH_4^+$  to  $NO_2^-$  and further to  $NO_3^-$  by nitrifying bacteria. Denitrification, on the other hand, converts  $NO_3^-$  to  $N_2$  under anaerobic conditions through a series of reduction steps, including the reduction of  $NO_3^-$  to  $NO_2^-$ ,  $NO$ , and ultimately  $N_2O$ .  $N_2O$  can further be reduced to  $N_2$  by denitrifying bacteria. Nitrification helps convert  $NH_3$  into plant-available  $NO_3^-$ , while

denitrification removes excess  $NO_3^-$  and releases N back into the atmosphere. Understanding these microbial processes is essential for managing and mitigating  $N_2O$  emissions in agricultural and natural ecosystems (Ghaly and Ramakrisnan, 2015).

Another microorganism involved in the nitrification and denitrification process is fungi, which can emit  $N_2$  and  $N_2O$  by denitrification and codenitrification. On the other hand, Archaea stimulating nitrification in marine ecosystems can promote denitrification in soils (Lu et al., 2020). According to Zajac and Zubrowska-Sudol (2022), nitrification releases  $N_2O$  when  $O_2$  is limited. Macdonald et al. (2016); Shaaban et al. (2018); and C. Wang et al. (2021) reported that  $N_2O$  emission will proportionately rise when pH and organic matter increase. Certainly, it happens when soil moisture also rises (from air dry to field capacity) around 5 to 40 °C. Nevertheless,  $N_2O$  emission can be lessened by applying nitrification inhibitors (Lam et al., 2017). Hence, the synthesis of  $N_2O$  is a complex process caused by numerous factors, such as pH, texture, soil moisture, soil temperature, organic matter, and abundance of  $O_2$  (Macdonald et al., 2016; Wrage-Mönnig et al., 2018; C. Wang et al., 2021; Romero et al., 2021).

Around 40% of  $N_2O$  is formed from human activities such as agriculture, fuel combustion and industrial processes (Addington et al., 2021).

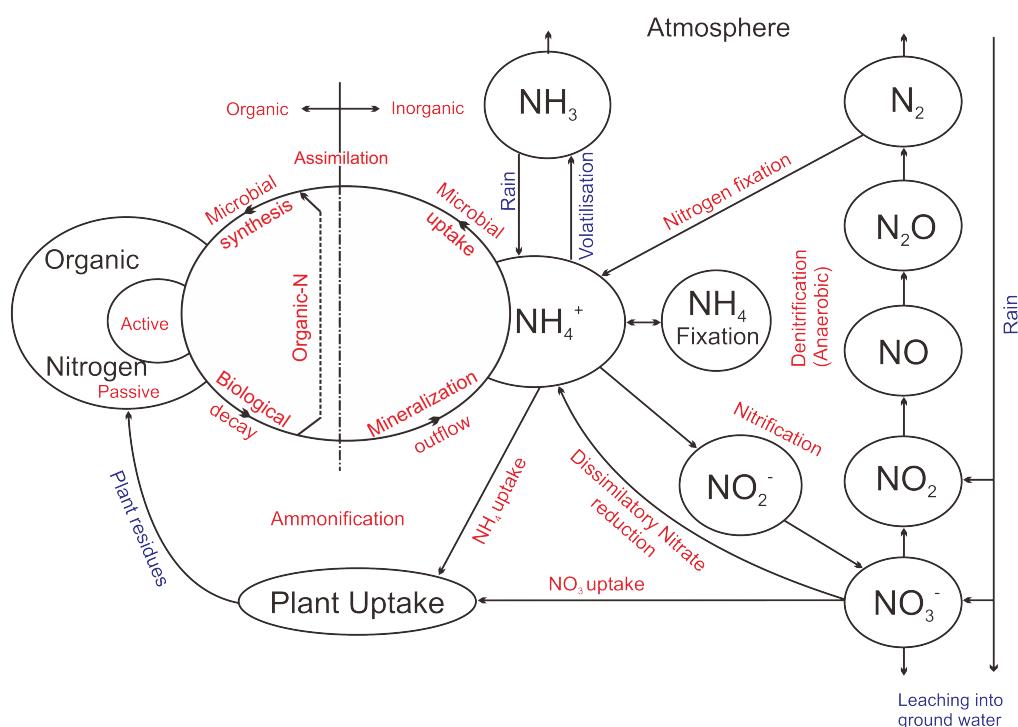


Figure 3. Nitrogen cycle (Ghaly and Ramakrisnan, 2015)

The amount of N<sub>2</sub>O emissions in Indonesia is presented in Table 1.

Table 1 provides an overview of the main sources of N<sub>2</sub>O emissions in Indonesia for the years 2010, 2015 and 2020. Agriculture is identified as the largest contributor, primarily due to the use of N-based fertilizers and other agricultural practices. Fossil fuel combustion and industrial processes also contribute to N<sub>2</sub>O emissions, albeit to a lesser extent. Land use change and forest activities emerge as significant sources, particularly in 2015, indicating the impact of deforestation and forest degradation. Waste management plays a relatively stable role in N<sub>2</sub>O emissions.

### Factors determining N<sub>2</sub>O production in agriculture cultivation

As aforementioned that N<sub>2</sub>O is formed during enzymatic nitrification and denitrification processes and chemo-denitrification (Stanton et al., 2018). N<sub>2</sub>O emission is affected by the chemical and physical conditions of the soil, such as soil aeration, temperature, moisture, compaction, texture, pH, available N, C/N ratio, organic matter, and by soil management and crop rotation (Ashiq et al., 2021). Moreover, crop rotation, N sources, and fertilizing method (time and depth) also influence N<sub>2</sub>O emission (Rychel et al., 2020).

#### Soil properties

The dynamic of N<sub>2</sub>O emission not only influence by soil N availability but also soil properties such as pH, soil moisture, organic matter addition (Baruah et al., 2010). Zhu et al. (2021) reported that high soil moisture stimulates N<sub>2</sub>O emission, but amount of N<sub>2</sub>O flux is differences among different of soil type. Soil moisture has a great role in N<sub>2</sub>O emission since it controls the O<sub>2</sub> supply to soil microbes (Butterbach-Bahl et al., 2013). Around 70% to 80% WFPS (depending on the soil type) is the optimum range for N<sub>2</sub>O emission from soil,

with the major final product is N<sub>2</sub> (denitrification process) (Davidson et al., 2000). In line with this statement, Butterbach-Bahl et al. (2013) reported that the optimum N<sub>2</sub>O emission from soil was under wetter conditions than 80% WFPS. At higher soil moisture, the major end product of denitrification is N<sub>2</sub>.

Beside soil moisture, it was found that temperature has a significant influence on denitrification process (Qu et al., 2022). Increasing temperature due to soil respiration leads to a depletion of soil oxygen concentrations and increases soil anaerobiosis. These activities also stimulate several temperature-sensitive microbial processes within the N cycle, and certain effect on N<sub>2</sub>O emission (Butterbach-Bahl et al., 2013).

Zhu et al. (2021) reported that N<sub>2</sub>O fluxes also increased following either manure application. Soil N<sub>2</sub>O emissions were larger for urine than dung addition. Several macro-faunas such as termites are predicted to influence N<sub>2</sub>O emission. It is possible that termites may transfer some organic matters from dung or manure away and effect on soil texture and pH. Clay particles hold water in soil aggregates. Furthermore, clay soils mostly have a low gas diffusivity due to small average pore sizes (Hu et al., 2018). In addition, high clay content normally ties in a high cation exchange capacity (CEC). As a consequence, the mineralized NH<sub>4</sub> ion can be adsorbed since the CEC is high or even fixed to the clay minerals (Chantigny et al., 2004), it takes an impact on decreasing soil NH<sub>4</sub> ion availability for nitrification and, thus, NO<sub>3</sub> ion production. Hence, the soil NH<sub>4</sub><sup>+</sup> sorption capacity has been found to affect N<sub>2</sub>O production (Venterea et al., 2015).

#### N fertilizing

Even though NH<sub>4</sub><sup>+</sup> is used in the nitrification process and NO<sub>3</sub><sup>-</sup> is used in the denitrification process, N<sub>2</sub>O emission from agricultural fields

Table 1. Main sources of N<sub>2</sub>O in the atmosphere in Indonesia

Source	Gg CO <sub>2</sub> e		
	2010	2015	2020
Anthropic sources:			
Fossil fuel combustion	15.12	17.39	0.03
Industrial process and product use	0.30	5.00	3.72
Agriculture	152.26	157.29	171.09
Land use change and forestry	0.00	30.89	2.53
Other (waste)	9.27	9.96	11.91
Total anthropic sources	167.68	210.57	189.28

Source: Ministry of Environment and Forestry of Indonesia (2022)

usually comes in proportion to the N application rate (Wrage-Mönnig et al., 2018). In addition, Roy et al. (2014) stated that the increase in N<sub>2</sub>O emissions is directly proportional to the application rate of N fertilizer.

In conventional Indonesian agriculture, urea is an N fertilizer that is commonly used. Urea is subject to N losses from cultivated fields that include “denitrification”, or the conversion of NO<sub>3</sub><sup>-</sup> into N<sub>2</sub> (gas); leaching (the downward movement of NO<sub>3</sub><sup>-</sup> out of the root zone); plant uptake and removal in harvested portions of the crop; and NH<sub>3</sub> volatilization (from soils and some plants). Two other reactions, “immobilization” (uptake by microorganisms) and “exchange” (binding of soil particles), are considered temporary “losses” because the N remains in the soil, and most of it becomes available (Menegat et al., 2022).

The application of N fertilizer increases the potential for N<sub>2</sub>O production by providing a substrate. There is strong evidence that the form of N fertilizer, especially inorganic N, influences the production of N<sub>2</sub>O (Xie et al., 2022). For instance, Chen et al. (2019) pointed out that N fertilizer above plant needs is more likely to result in N<sub>2</sub>O flux. Similarly, Hassan et al. (2022) stated the importance of matching N application. N<sub>2</sub>O flux is significantly increased using N fertilizers, especially when followed by rain (Suwardi et al., 2015). Due to rain, increasing groundwater levels can affect the denitrification process (Weymann et al., 2009; Jurado et al., 2017).

#### Soil tillage

Tillage practices have been widely known for affecting cropping systems, both conservation tillage practices such as no-tillage, reduced or minimum tillage, and conventional tillage (Lv et al., 2023). Generally, conservation tillage is related to water infiltration improvement, ameliorating soil structure, and reduced soil erosion (Rehman et al., 2023). Soil tillage practices also affect N<sub>2</sub>O emissions due to the N<sub>2</sub> cycle in soil. The activity of *Nitrosomonas* sp. and *Nitrobacter* sp. that handle nitrification are determined by optimal soil temperature and moisture condition.

Abundant soil moisture and available organic carbon could support the complete reduction of low to moderate levels of NO<sub>3</sub> to N<sub>2</sub> gas, consequently reducing the net amount of N<sub>2</sub>O produced (Senbayram et al., 2019). Furthermore, higher levels of NO<sub>3</sub> led to higher net N<sub>2</sub>O:N<sub>2</sub> emission ratios, because the reduction of NO<sub>3</sub> in

comparison with N<sub>2</sub>O was more energy-efficient (Friedl et al., 2020). According to Huang et al. (2022), it seems that during the period of transition from conventional tillage to no-tillage, the N immobilization is accompanied by slow soil organic matter turnover. Alam et al. (2020) suggested that N availability for the crops was lower under no-tillage than under conventional tillage. Moreover, others found that the higher application rates of inorganic fertilizer can be beneficial to the N uptake of crop during the transition period from standard to minimum tillage to account for the lower N mineralization rates (Mondal and Chakraborty, 2022). Some studies showed that N<sub>2</sub>O emission was greater in no-tilled soils than in conventionally tilled soils (Gregorich et al., 2008; Holder et al., 2019), but other studies reported that N<sub>2</sub>O emission was greater in conventionally tilled soils than in no-tilled soils (Yoo et al., 2016; Sosulski et al., 2022).

#### Other factors

The results of researchers exploration on the factors that affect N<sub>2</sub>O flux for both mineral soil and peatland showed that in corn fields, the average soil moisture was 46.4%, peanuts 36.9%, and cassava 43.4%. In peatland areas, the average soil moisture for *A. crassiparva* at 1 year was 190.6% while *A. crassiparva* at 3 years was 202.5%, and the average soil moisture in natural primary forest was 426.3%. Soil moisture is a critical factor in N<sub>2</sub>O production as it affects the availability of O<sub>2</sub> and the activity of soil microorganisms. Higher soil moisture levels can lead to higher N<sub>2</sub>O emissions (Ramzan et al., 2020).

#### N<sub>2</sub>O fluxes in the crop fields

Global N<sub>2</sub>O emissions from row-crop agriculture are considered to be the greatest contributor to global N<sub>2</sub>O flux (Lawrence et al., 2021), with cultivated soils comprising 27% of the total N<sub>2</sub>O-N added from all known sources. It might be caused by normal fertilization application only for on-row crops (Shcherbak and Robertson, 2019). Hence, the concentration of N element on-row is typically higher than inter-row crops. Furthermore, fertilization also causes increased production of N<sub>2</sub>O. Corollary to this, Winkhart et al. (2022) found that the highest N<sub>2</sub>O flux in corn occurred during the early days after planting, resulting from the intensive use of organic manure and N fertilizer by most traditional farmers, which creates an environment that is suitable to the formation of N<sub>2</sub>O. Similarly,

Wu et al. (2022) stated that production increases, either through the process of nitrification or through denitrification, when N fertilizer is applied.

Researchers study showed that total N<sub>2</sub>O fluxes during the study period on corn (77-d), peanut (75-d) and cassava (246-d) fields were 2.10 kg N-N<sub>2</sub>O ha<sup>-1</sup>, 0.19 kg N-N<sub>2</sub>O ha<sup>-1</sup> and 0.62 kg N-N<sub>2</sub>O ha<sup>-1</sup> respectively (Table 2). The N<sub>2</sub>O flux of corn is higher than peanut and cassava, and this is likely due to the differences in their respective growth and management practices. Corn is a crop that requires a lot of N fertilizer to support its growth and yield. The highest fluxes occurred following basal and top dressing N fertilization in maize regardless of the growing season and forage is previously grown (Grassmann et al., 2020).

Nitrogen fertilizer is a primary source of N<sub>2</sub>O emissions from agricultural soils. The higher N<sub>2</sub>O flux in corn may be attributed to the higher nitrogen inputs in the form of fertilizer that are typically used to grow this crop. Corn cultivation may be more intensive than peanut and cassava, so corn needs higher N fertilizer inputs than other crops (Deng et al., 2015). In contrast, peanuts and cassava are crops that require relatively less N fertilizer, and therefore may release less N<sub>2</sub>O. In the cassava field, the data likewise showed that N<sub>2</sub>O flux increased significantly with the application of fertilizer, particularly when followed by rain. Rainwater can contain additional N that can contribute to N<sub>2</sub>O production in agricultural sites. Nitrogen deposition from rainwater can increase the availability of N in the soil, which can lead to increased N<sub>2</sub>O emissions (Ma et al., 2018).

Researchers study site is located in an area extensively used for agricultural cultivation, which is a tropical region with relatively high rainfall. Areas with high rainfall can cause submergence during intense rainfall, which occurs when the soil's ability to drain water is compromised, often due to factors such as heavy rainfall, poor soil structure, or inadequate drainage (Fukao et al., 2019). Rainfall can

Table 2. Total N<sub>2</sub>O fluxes in crop fields on mineral soils

Land use type/Age	Total N-N <sub>2</sub> O flux (kg N-N <sub>2</sub> O ha <sup>-1</sup> )
Corn (77 days)	2.10
Peanut (75 days)	0.19
Cassava (246 days)	0.62

increase the water content of the soil, which can affect N<sub>2</sub>O production through denitrification processes.

Groundwater can affect denitrification process, either directly or indirectly by way of: (1) forming a suitable environment for microorganisms to grow and move, (2) restricting the availability of O<sub>2</sub> in the soil micropores, and (3) facilitating the release of C and N substrates through wetting and drying cycles. Yet, soil water limits the presence of O<sub>2</sub> in the soil pores, so the N<sub>2</sub>O easily formed in a bit anaerobic condition (Geng et al., 2017). Rainfall caused excessive dampness of the soil, which could bring about denitrification processes, thereby producing N<sub>2</sub>O (Geng et al., 2017; Friedl et al., 2020).

N<sub>2</sub>O is produced when plant-based N is subjected to the bacterial processes of denitrification and nitrification. Denitrification is the reduction of NO<sub>3</sub><sup>-</sup> back into the largely inert N<sub>2</sub> for completing the cycle (Abatenh et al., 2018). The most important factors controlling these processes are N (NH<sub>4</sub><sup>+</sup> and NO<sub>3</sub><sup>-</sup>) concentrations in mineral soils, O<sub>2</sub> under partial pressure and, in the case of denitrification, available carbon to fuel the heterotrophic processes (Giles et al., 2012).

In this study, the N<sub>2</sub>O fluxes ranged from -0.132 mg N-N<sub>2</sub>O m<sup>-2</sup> d<sup>-1</sup> in the cassava field to 41.283 mg N-N<sub>2</sub>O m<sup>-2</sup> d<sup>-1</sup> in the corn field, which received the highest degree of fertilizer application. The N<sub>2</sub>O flux from the corn field registered the highest, compared to those coming from the peanut and cassava fields. Generally, the highest N<sub>2</sub>O fluxes occurred in the first week after fertilization, and more so when followed by rain. The mean peak N<sub>2</sub>O flux in the corn field was 23.499 mg N-N<sub>2</sub>O m<sup>-2</sup> d<sup>-1</sup> at the beginning of planting, while in the peanut and cassava fields, the corresponding values were 0.550 mg N-N<sub>2</sub>O m<sup>-2</sup> d<sup>-1</sup> and 1.278 mg N-N<sub>2</sub>O m<sup>-2</sup> d<sup>-1</sup> respectively. The summary data on N<sub>2</sub>O fluxes and environmental variables are shown in Figure 4, 5 and 6. N<sub>2</sub>O flux on-row in the corn field was higher than in the inter-row, especially at the beginning of planting. In comparison, the N<sub>2</sub>O flux in the cassava field did not show any significant difference between on-row and inter-row.

### Comparison of mineral and peatland N<sub>2</sub>O fluxes in the peatland fields

The N<sub>2</sub>O flux values presented in the Table 3 suggest that there are significant differences in N<sub>2</sub>O emissions among different land use types



and ages. In mineral soils, corn has the highest  $N_2O$  flux at  $9.95 \text{ kg N-N}_2\text{O ha}^{-1} \text{ y}^{-1}$ . In peatland soils, *A. crassicaarpa* (1 year) has the highest  $N_2O$  flux at  $13.85 \text{ kg N-N}_2\text{O ha}^{-1} \text{ y}^{-1}$ . The application of chemical fertilizer is the prevailing farm practice in cropland in the mineral soil around  $5 \text{ ton ha}^{-1}$  of manure is applied on cassava fields and  $500 \text{ kg ha}^{-1}$  of phonska is applied on peanut fields. The fertilizer dose of corn fields is  $373 \text{ kg ha}^{-1}$  of urea,  $92 \text{ kg ha}^{-1}$  of SP36, and  $186 \text{ kg ha}^{-1}$  of phonska. This is because of fertilizer that makes  $N_2O$  flux on corn fields the highest than others.

Based on the  $N_2O$  flux values presented in the Table 3 and the fertilizer doses provided, the amount of N lost from the fertilizer in the form of  $N_2O$  can be calculated for each crop. For corn, with an  $N_2O$  flux of  $9.95 \text{ kg N-N}_2\text{O ha}^{-1} \text{ y}^{-1}$  and a fertilizer dose of  $373 \text{ kg ha}^{-1}$  of urea,  $92 \text{ kg ha}^{-1}$  of SP36, and  $186 \text{ kg ha}^{-1}$  of phonska, the amount of N lost as  $N_2O$  is estimated to be  $1.89 \text{ kg N ha}^{-1}$

$\text{y}^{-1}$ . For peanut, with an  $N_2O$  flux of  $0.90 \text{ kg N-N}_2\text{O ha}^{-1} \text{ y}^{-1}$  and a fertilizer dose of  $500 \text{ kg ha}^{-1}$  of phonska, the amount of N lost as  $N_2O$  is estimated to be  $0.07 \text{ kg N ha}^{-1} \text{ y}^{-1}$ . Lastly, for cassava, with an  $N_2O$  flux of  $0.92 \text{ kg N-N}_2\text{O ha}^{-1} \text{ y}^{-1}$  and a fertilizer dose of  $5 \text{ tons ha}^{-1}$  of manure (assuming 0.5% N content), the amount of N lost as  $N_2O$  is estimated to be  $0.00023 \text{ kg N ha}^{-1} \text{ y}^{-1}$ . A study in Chile by Salazar et al. (2021) found that the N fertilizer efficiency (NFE) in maize was 45.5% in the maize-fallow rotation and 41.7% in the maize-cover crop rotation. The study also found that the  $\text{NO}_3^-$  leaching losses were 5.4 and  $4.5 \text{ kg N ha}^{-1}$  in the maize-fallow and maize-cover crop rotations, respectively. Bijay-Singh and Craswell (2021) found that only a portion of the fertilizer N is directly lost from the cropping system via leaching and/or in gaseous forms ( $N_2O$  and  $\text{NH}_3$ ).

The P addition has stimulated the activities of nitrifying or denitrifying bacteria due to the relief

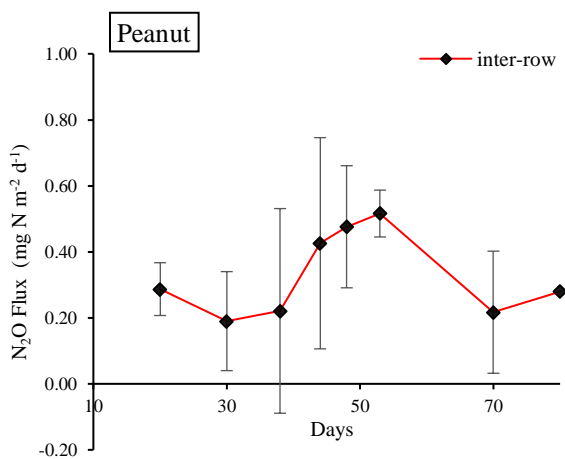


Figure 4.  $N_2O$  flux from peanut field

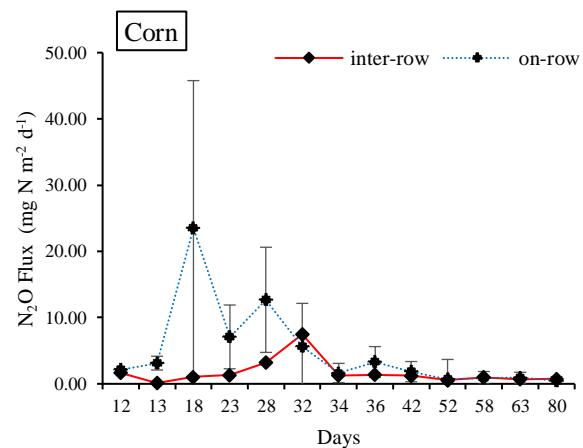


Figure 5.  $N_2O$  flux from corn field

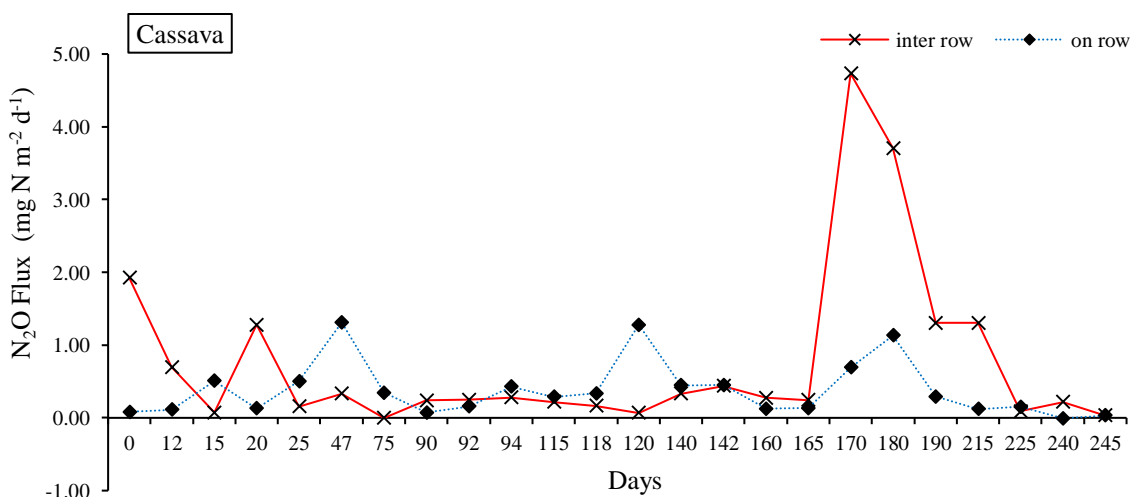


Figure 6.  $N_2O$  flux from cassava field

from their P shortage. Phospat addition may lead to increases in nitrification or denitrification activity, which results in higher N<sub>2</sub>O and NO emissions (Mori et al., 2010). The addition of fertilizer P significantly increases soil N<sub>2</sub>O emissions. Increased soil moisture and the application of N fertilizer also increase N<sub>2</sub>O emissions. Combined applications of fertilizers P and N indicate higher N<sub>2</sub>O emissions compared to their applications on a single basis (Sarkar et al., 2017). This is attributed to the alteration of soil properties and microbial activity, leading to increased rates of nitrification and denitrification. The presence of both P and N fertilizers can enhance N availability and microbial activity, creating favorable conditions for N<sub>2</sub>O production (Zhou et al., 2020). Additionally, the interaction between P and N fertilizers can disrupt the N cycle and reduce N uptake by plants, resulting in more N available for N<sub>2</sub>O emissions (Grohskopf et al., 2019). By significant addition of N, it will directly increase N substrates to nitrification and denitrification process and it would probably be rising N<sub>2</sub>O (F. Wang et al., 2014). This also happens when adding P fertilizer on soil. Increasing soil available P after P fertilizer application significantly will increase soil microbial biomass carbon.

In the corn field, N<sub>2</sub>O flux occurred within around three weeks of fertilizer application, suggesting increased N availability. Furthermore, peak N<sub>2</sub>O emissions occurred within heavy daily rainfall events  $\pm 2400$  mm. Soil aeration following saturation is important for the diffusion and release of GHGs. Continuous saturation leads to strong anaerobic conditions in the soil, which can lead to the full reduction of NO<sub>3</sub><sup>-</sup> to N<sub>2</sub> (Crézé and Madramootoo, 2019). Likewise, in the *Acacia* plantation, N addition significantly increased the rates of nitrification (Mori et al., 2017).

The N<sub>2</sub>O flux values for *A. crassicaarpa* plantations decrease as the plantation ages. This could be due to the establishment of a stable soil microbial community, which can reduce N<sub>2</sub>O emissions. Additionally, the reduction in fertilizer use as the plantation ages can also contribute to the decrease in N<sub>2</sub>O emissions. Research results from Ishizuka et al. (2021) also show that N<sub>2</sub>O emissions within 1 year of *Acacia* plantation can potentially emit large amounts of N<sub>2</sub>O emissions. However, the large amount of N<sub>2</sub>O emission decreases rapidly and reaches the same level of equilibrium as the flux observed at the age of 3 years of *Acacia* plantation. The fertilizer addition significantly increased N<sub>2</sub>O emission *Acacia* plantation in the first year. However, in the third year, soil N<sub>2</sub>O emissions significantly decreased (Zhang et al., 2014). The difference in N<sub>2</sub>O flux between that from the *A. crassicaarpa* 3 years and that from a similar plantation but with no fine root and litter (-R-L) reflects differences in rate of the respiration and root exudation (Sumawinata et al., 2014).

The values for natural primary forests also show variation, with slightly lower N<sub>2</sub>O flux values for the “natural primary forest- R-L” category. This could be due to differences in vegetation cover and soil moisture content in the different areas. Moreover, in tropical regions, plants and microbes compete fiercely for resources. As a result of this competition, plants tend to absorb more nutrients while losing less nitrogen gas, but the extent of their absorption depends heavily on their density. In a particular, the higher root density significantly increases the competition for nitrogen uptake through roots. It can be inferred that biotic factors play a vital role in this competition, and the formation of mycorrhiza and root density is critical in shifting

Table 3. Total N<sub>2</sub>O fluxes in agricultural land on peatland soils

Land use type/Age	Total N <sub>2</sub> O flux (kg N-N <sub>2</sub> O ha <sup>-1</sup> y <sup>-1</sup> )
Mineral soils:	
Corn (77 days)	9.95
Peanut (75 days)	0.90
Cassava (246 days)	0.92
Peatland soils:	
<i>A. crassicaarpa</i> (1 year)	13.85
<i>A. crassicaarpa</i> (3 years)	6.65
<i>A. crassicaarpa</i> (3 years-R-L)	2.87
Natural primary forest	5.66
Natural primary forest-R-L	4.31

Note: -R-L = without fine root and litter

the acquisition of nitrogen to the roots (Netherway et al., 2021).

In a natural primary forest, the nutrient cycling process is often controlled by a complex network of interactions between plants, microbes, and animals, which can help to reduce nitrogen losses and maintain a relatively stable nitrogen balance. Additionally, natural forests usually have a lower soil disturbance compared to agricultural or plantation areas, which can limit the exposure of soil to air and reduce the conditions for  $N_2O$  production. Furthermore, the presence of fine roots and litter can contribute to  $N_2O$  emissions. Because it contains large amounts of carbon and other nutrients that can support the growth of microorganisms and promote the conversion of  $N_2O$  to harmless gases like  $N_2$  or  $NO$ . Microorganisms obtain N from root and soil residues. The soil continuously received root exudates from dead fine roots. Therefore, the natural primary forest without fine roots and litter may lead to a slight increase in  $N_2O$  emissions (Kim et al., 2022).

Overall, the results highlight the importance of considering land use type and age when assessing  $N_2O$  emissions. Management practices such as fertilizer use, soil disturbance, and vegetation cover can all have significant impacts on  $N_2O$  emissions (Roy et al., 2014; Krauss et al., 2017; Menegat et al., 2022). The management practices in the agricultural field are crucial in impacting  $N_2O$  emissions since it dictates the quantity of soil N input, which can alter the soil environment and microbial conditions. X. Wang et al. (2021) stated that management variables comprise the quantity and sort of fertilizer applied, the types of crops grown, and the tilling procedures used, which can also impact the amount of crop residue left on the soil surface. This section is crucial that the effects of specific agricultural management practices can affect  $N_2O$  emissions.

### **The impact of N fertilizer on $N_2O$ flux emissions in agricultural land**

In agricultural land, increases in  $N_2O$  emissions typically follow N fertilization but only for a short time. After this time, the emission rates are reduced, fluctuating around a low baseline level which is independent of the amount of fertilizer applied (Ruiz et al., 2021). Mineral N applications, along with organic matter amendments, generally increase total denitrification and  $N_2O$  production.  $N_2O$  emissions from the soil can vary by order of magnitude in a location, both spatially and

temporally.  $N_2O$  emissions are predicted not only from fertilization but also from biological/bacterial activity, which plays an important role in  $N_2O$  production (Harris et al., 2022). The results of this study suggest that as farmers apply excessive N-fertilizer, the formation of  $N_2O$  is thus increased. Beside the observable high levels of  $NH_4^+$  and  $NO_3^-$ , the excess N concentration in the corn field can also be seen from the visible presence of succulent plant stems in the study area.

Nevertheless, emission peaks did not exclusively occur immediately after fertilizer application. Likewise, some studies have shown that  $N_2O$  emission from agricultural soil is significantly increased by the application of synthetic N fertilizers (Linn and Doran, 1984; Bronson et al., 2021). Moreover, Hassan et al. (2022) highlighted that the potential for  $N_2O$  emissions from agricultural soils may be lowered by changes in fertility and irrigation management. Fertilization and drainage can be coordinated so that drainage does not occur soon after fertilization and so that drainage does not greatly increase  $N_2O$  emissions during the growing season.

The increasing levels of atmospheric  $N_2O$  are of particular concern due to its considerably higher GWP relative to  $CO_2$ . For example, over a 20-yr period, 1 kg of  $N_2O$  will have 275 times the influence on global warming as 1 kg of  $CO_2$  (Solecki et al., 2022). Aside from emitting  $N_2O$ , the impacts of excessive fertilizer application can also damage plants, and would deserve the farmers, ultimately.  $NO_3^-$  leaching is also important from a water-quality perspective because it contributes to aquatic eutrophication and can pose a health risk to humans.

Nitrogen cycling reactions can affect volatilization by influencing the amount of  $NH_3$  available to volatilize. The process of volatilization of  $NH_3$  occurs not only in lowland rice but also in other crops. A study by Santos et al. (2020) found that  $NH_3$  loss differed among N sources in corn, suggesting that  $NH_3$  volatilization may occur in corn fields. Janke et al. (2022) pointed out that volatilization losses from urea can be reduced by in-soil banding or by incorporation of the fertilizer immediately after application. However, under a reduced tillage system, banding or incorporation leads to undesirable soil disturbance. At any rate, land management practices or systems that reduce GHG emissions would aid in slowing climate change. Guenet et al. (2021) stated that impacts

on farmers and land managers give part of which contributes to indirect N<sub>2</sub>O emissions, through excess N, changes in loss NO<sub>3</sub><sup>-</sup> leaching and N<sub>2</sub>O emissions that must be considered.

## CONCLUSIONS

This review examines the significant contributions of different sectors to N<sub>2</sub>O emissions in Indonesia and emphasizes the dominant role of agriculture as the primary source. Several factors affecting on N<sub>2</sub>O both on mineral and peat soil are soil properties (soil temperature, moisture), soil input such as organic matter application and N fertilizing, and agriculture practice management such as soil tillage system. Nitrogen fertilizing takes a significant effect on N<sub>2</sub>O emission. Sustainable agricultural practices, including precise N fertilizer management and minimizing soil disturbance, are crucial for reducing N<sub>2</sub>O emissions from agricultural activities. Additionally, addressing N<sub>2</sub>O emissions in Indonesia requires a comprehensive approach that encompasses clean energy transitions, land use policies, and improved waste management. The findings of this review provide valuable insights for policymakers, researchers, and practitioners, offering a foundation for developing effective strategies to mitigate N<sub>2</sub>O emissions in Indonesia and similar tropical farming systems. By implementing these strategies, Indonesia can make significant progress in reducing its overall N<sub>2</sub>O emissions and contribute to global efforts in combating climate change.

## ACKNOWLEDGEMENT

The authors would like to thank Sinar Mas Forestry and Hokkaido University for funding assistance in this research.

## REFERENCES

- Abatenh, E., Gizaw, B., Tsegaye, Z., & Tefera, G. (2018). Microbial function on climate change—A review. *Environment Pollution and Climate Change*, 2(1), 147. <https://doi.org/10.4172/2573-458x.1000147>
- Addington, O., Zeng, Z. C., Pongetti, T., Shia, R. L., Gurney, K. R., Liang, J., Roest, G., He, L., Yung, Y. L., & Sander, S. P. (2021). Estimating nitrous oxide (N<sub>2</sub>O) emissions for the Los Angeles Megacity using mountaintop remote sensing observations. *Remote Sensing of Environment*, 259, 112351. <https://doi.org/10.1016/j.rse.2021.112351>
- Ahmad, A. (2021). Soil fertility mapping of corn plant based on minerals in Jeneponto Regency. *Jurnal Ecosolum*, 10(2), 1–14. Retrieved from <http://journal.unhas.ac.id/index.php/ecosolum/article/view/18682>
- Alam, M. K., Bell, R. W., Haque, M. E., Islam, M. A., & Kader, M. A. (2020). Soil nitrogen storage and availability to crops are increased by conservation agriculture practices in rice-based cropping systems in the Eastern Gangetic Plains. *Field Crops Research*, 250, 107764. <https://doi.org/10.1016/j.fcr.2020.107764>
- Arai, H., Hadi, A., Darung, U., Limin, S. H., Takahashi, H., Hatano, R., & Inubushi, K. (2014). Land use change affects microbial biomass and fluxes of carbon dioxide and nitrous oxide in tropical peatlands. *Soil Science and Plant Nutrition*, 60(3), 423–434. <https://doi.org/10.1080/00380768.2014.903576>
- Ashiq, W., Vasava, H., Cheema, M., Dunfield, K., Daggupati, P., & Biswas, A. (2021). Interactive role of topography and best management practices on N<sub>2</sub>O emissions from agricultural landscape. *Soil and Tillage Research*, 212, 105063. <https://doi.org/10.1016/j.still.2021.105063>
- Baruah, K. K., Gogoi, B., Gogoi, P., & Gupta, P. K. (2010). N<sub>2</sub>O emission in relation to plant and soil properties and yield of rice varieties. *Agronomy for Sustainable Development*, 30(4), 733–742. <https://doi.org/10.1051/agro/2010021>
- Bijay-Singh, & Craswell, E. (2021). Fertilizers and nitrate pollution of surface and ground water: An increasingly pervasive global problem. *SN Applied Sciences*, 3(4), 518. <https://doi.org/10.1007/s42452-021-04521-8>
- Bronson, K. F., Hunsaker, D. J., El-Shikha, D. M., Rockholt, S. M., Williams, C. F., Rasutis, D., Soratana, K., & Venterea, R. T. (2021). Nitrous oxide emissions, N uptake, biomass, and rubber yield in N-fertilized, surface-irrigated guayule. *Industrial Crops and Products*, 167, 113561. <https://doi.org/10.1016/j.indcrop.2021.113561>

- Buschmann, C., Röder, N., Berglund, K., Berglund, Ö., Lærke, P. E., Maddison, M., Mander, Ü., Myllys, M., Osterburg, B., & van den Akker, J. J. H. (2020). Perspectives on agriculturally used drained peat soils: Comparison of the socioeconomic and ecological business environments of six European regions. *Land Use Policy*, *90*, 104181. <https://doi.org/10.1016/j.landusepol.2019.104181>
- Busman, N. A., Melling, L., Goh, K. J., Imran, Y., Sangok, F. E., & Watanabe, A. (2023). Soil CO<sub>2</sub> and CH<sub>4</sub> fluxes from different forest types in tropical peat swamp forest. *Science of the Total Environment*, *858*, 159973. <https://doi.org/10.1016/j.scitotenv.2022.159973>
- Butterbach-Bahl, K., Baggs, E. M., Dannenmann, M., Kiese, R., & Zechmeister-Boltenstern, S. (2013). Nitrous oxide emissions from soils: How well do we understand the processes and their controls? *Philosophical Transactions of the Royal Society B: Biological Sciences*, *368*, 20130122. <https://doi.org/10.1098/rstb.2013.0122>
- Chantigny, M., Rochette, P., Angers, D., Massé, D., & Côté, D. (2004). Ammonia volatilization and selected soil characteristics following application of anaerobically digested pig slurry. *Soil Science Society of America Journal*, *68*(1), 306–312. <https://doi.org/10.2136/sssaj2004.3060>
- Chen, H., Zhou, J., Li, B., & Xiong, Z. (2019). Yield-scaled N<sub>2</sub>O emissions as affected by nitrification inhibitor and overdose fertilization under an intensively managed vegetable field: A three-year field study. *Atmospheric Environment*, *206*, 247–257. <https://doi.org/10.1016/j.atmosenv.2019.02.036>
- Costa, C., Wironen, M., Racette, K., & Wollenberg, E. (2021). *Global Warming Potential\* (GWP\*): Understanding the implications for mitigating methane emissions in agriculture*. Wageningen, The Netherlands: CGIAR Research Program on Climate Change, Agriculture and Food Security (CCAFS). Retrieved from <https://hdl.handle.net/10568/114632>
- Crézé, C. M., & Madramootoo, C. A. (2019). Water table management and fertilizer application impacts on CO<sub>2</sub>, N<sub>2</sub>O and CH<sub>4</sub> fluxes in a corn agro-ecosystem. *Scientific Reports*, *9*, 2692. <https://doi.org/10.1038/s41598-019-39046-z>
- Davidson, E. A., Keller, M., Erickson, H. E., Verchot, L. V., & Veldkamp, E. (2000). Testing a conceptual model of soil emissions of nitrous and nitric oxides: Using two functions based on soil nitrogen availability and soil water content, the hole-in-the-pipe model characterizes a large fraction of the observed variation of nitric oxide and nitrous oxide emissions from soils. *BioScience*, *50*(8), 667–680. [https://doi.org/10.1641/0006-3568\(2000\)050\[0667:TACMOS\]2.0.CO;2](https://doi.org/10.1641/0006-3568(2000)050[0667:TACMOS]2.0.CO;2)
- Delgado, J. A., & Follett, R. F. (2002). Carbon and nutrient cycles. *Journal of Soil and Water Conservation*, *57*(6), 455–464. Retrieved from <https://www.jswnonline.org/content/57/6/455.short>
- Deng, Q., Hui, D., Wang, J., Iwuzo, S., Yu, C.-L., Jima, T., Smart, D., Reddy, C., & Dennis, S. (2015). Corn yield and soil nitrous oxide emission under different fertilizer and soil management: A three-year field experiment in middle Tennessee. *PLoS ONE*, *10*(4), e0125406. <https://doi.org/10.1371/journal.pone.0125406>
- Dhandapani, S., Evers, S., Boyd, D., Evans, C. D., Page, S., Parish, F., & Sjogersten, S. (2023). Assessment of differences in peat physico-chemical properties, surface subsidence and GHG emissions between the major land-uses of Selangor peatlands. *CATENA*, *230*, 107255. <https://doi.org/10.1016/j.catena.2023.107255>
- Friedl, J., Cardenas, L. M., Clough, T. J., Dannenmann, M., Hu, C., & Scheer, C. (2020). Measuring denitrification and the N<sub>2</sub>O:(N<sub>2</sub>O + N<sub>2</sub>) emission ratio from terrestrial soils. *Current Opinion in Environmental Sustainability*, *47*, 61–71. <https://doi.org/10.1016/j.cosust.2020.08.006>
- Fukao, T., Barrera-Figueroa, B. E., Juntawong, P., & Peña-Castro, J. M. (2019). Submergence and waterlogging stress in plants: A review highlighting research opportunities and understudied aspects. *Frontiers in Plant Science*, *10*, 340. <https://doi.org/10.3389/fpls.2019.00340>
- Geng, S., Chen, Z., Han, S., Wang, F., & Zhang, J. (2017). Rainfall reduction amplifies the stimulatory effect of nitrogen addition on

- N<sub>2</sub>O emissions from a temperate forest soil. *Scientific Reports*, 7, 43329. <https://doi.org/10.1038/srep433290>
- Ghaly, A., & Ramakrisnan, V. (2015). Nitrogen sources and cycling in the ecosystem and its role in air, water and soil pollution: A critical review. *Journal of Pollution Effects & Control*, 3(2), 136. <https://doi.org/10.4172/2375-4397.1000136>
- Giles, M., Morley, N., Baggs, E. M., & Daniell, T. J. (2012). Soil nitrate reducing processes—Drivers, mechanisms for spatial variation, and significance for nitrous oxide production. *Frontiers in Microbiology*, 3, 30024. <https://doi.org/10.3389/fmicb.2012.00407>
- Grassmann, C. S., Mariano, E., Rocha, K. F., Gilli, B. R., & Rosolem, C. A. (2020). Effect of tropical grass and nitrogen fertilization on nitrous oxide, methane, and ammonia emissions of maize-based rotation systems. *Atmospheric Environment*, 234, 117571. <https://doi.org/10.1016/j.atmosenv.2020.117571>
- Gregorich, E. G., Rochette, P., St-Georges, P., McKim, U. F., & Chan, C. (2008). Tillage effects on N<sub>2</sub>O emission from soils under corn and soybeans in Eastern Canada. *Canadian Journal of Soil Science*, 88(2), 153–161. <https://doi.org/10.4141/CJSS06041>
- Grohskopf, M. A., Corrêa, J. C., Fernandes, D. M., Teixeira, P. C., Cruz, C. V., & Mota, S. C. A. (2019). Interaction between phosphorus and nitrogen in organomineral fertilizer. *Communications in Soil Science and Plant Analysis*, 50(21), 2742–2755. <https://doi.org/10.1080/00103624.2019.1678632>
- Guenet, B., Gabrielle, B., Chenu, C., Arrouays, D., Balesdent, J., Bernoux, M., Bruni, E., Caliman, J. P., Cardinael, R., Chen, S., Ciais, P., Desbois, D., Fouche, J., Frank, S., Henault, C., Lugato, E., Naipal, V., Nesme, T., Obersteiner, M., ... Zhou, F. (2021). Can N<sub>2</sub>O emissions offset the benefits from soil organic carbon storage? *Global Change Biology*, 27(2), 237–256. <https://doi.org/10.1111/gcb.15342>
- Gunawan, H. (2018). Indonesian peatland functions: Initiated peatland restoration and responsible management of peatland for the benefit of local community, case study in Riau and West Kalimantan Provinces. *Asia in Transition*, 7, 117–138. [https://doi.org/10.1007/978-981-10-8881-0\\_6](https://doi.org/10.1007/978-981-10-8881-0_6)
- Harris, E., Yu, L., Wang, Y. P., Mohn, J., Henne, S., Bai, E., Barthel, M., Bauters, M., Boeckx, P., Dorich, C., Farrell, M., Krummel, P. B., Loh, Z. M., Reichstein, M., Six, J., Steinbacher, M., Wells, N. S., Bahn, M., & Rayner, P. (2022). Warming and redistribution of nitrogen inputs drive an increase in terrestrial nitrous oxide emission factor. *Nature Communications*, 13(1), 4310. <https://doi.org/10.1038/s41467-022-32001-z>
- Hassan, M. U., Aamer, M., Mahmood, A., Awan, M. I., Barbanti, L., Seleiman, M. F., Bakhsh, G., Alkharabsheh, H. M., Babur, E., Shao, J., Rasheed, A., & Huang, G. (2022). Management strategies to mitigate N<sub>2</sub>O emissions in agriculture. *Life*, 12(3), 439. <https://doi.org/10.3390/life12030439>
- Hendri, J., Sumawinata, B., & Baskoro, D. P. T. (2015). CO<sub>2</sub> flux from tropical land uses on andisol in West Java, Indonesia. *Journal of Tropical Soils*, 19(3), 121–130. <https://doi.org/10.5400/jts.2014.v19i3.121-130>
- Hodnebrog, Ø., Myhre, G., Kramer, R. J., Shine, K. P., Andrews, T., Faluvegi, G., Kasoar, M., Kirkevåg, A., Lamarque, J. F., Mülmenstädt, J., Olivie, D., Samset, B. H., Shindell, D., Smith, C. J., Takemura, T., & Voulgarakis, A. (2020). The effect of rapid adjustments to halocarbons and N<sub>2</sub>O on radiative forcing. *Npj Climate and Atmospheric Science*, 3, 43. <https://doi.org/10.1038/s41612-020-00150-x>
- Holder, A. J., McCalmont, J. P., Rowe, R., McNamara, N. P., Elias, D., & Donnison, I. S. (2019). Soil N<sub>2</sub>O emissions with different reduced tillage methods during the establishment of Miscanthus in temperate grassland. *GCB Bioenergy*, 11(3), 539–549. <https://doi.org/10.1111/gcbb.12570>
- Hsiao, C. M. (2022). Economic growth, CO<sub>2</sub> emissions quota and optimal allocation under uncertainty. *Sustainability*, 14(14), 8706. <https://doi.org/10.3390/su14148706>
- Hu, W., Jiang, Y., Chen, D., Lin, Y., Han, Q., & Cui, Y. (2018). Impact of pore geometry and water saturation on gas effective diffusion coefficient in soil. *Applied Sciences*, 8(11), 2097. <https://doi.org/10.3390/app8112097>
- Huang, D., Chen, X., Zhang, S., Zhang, Y., Gao, Y., Zhang, Y., & Liang, A. (2022). No-tillage

- improvement of nitrogen absorption and utilization in a Chinese mollisol using  $^{15}\text{N}$ -Tracing Method. *Atmosphere*, 13(4), 530. <https://doi.org/10.3390/atmos13040530>
- IPCC. (2022a). *Climate change 2022: Impacts, adaptation and vulnerability. Contribution of working group III to the sixth assessment report of the intergovernmental panel on climate change*. Cambridge: Cambridge University Press. <https://doi.org/10.1017/9781009157926>
- IPCC. (2022b). Summary for Policymakers. In P. R. Shukla, J. Skea, R. Slade, A. Al Khouradajie, R. van Diemen, D. McCollum, M. Pathak, S. Some, P. Vyas, R. Fradera, M. Belkacemi, A. Hasija, G. Lisboa, S. Luz, & J. Malley (Eds.), *Climate Change 2022 - Mitigation of Climate Change: Working Group III Contribution to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change* (pp. 3–48). Cambridge: Cambridge University Press. <https://doi.org/10.1017/9781009157926>
- IPCC. (2023). Summary for Policymakers. In *Climate Change 2021 – The Physical Science Basis: Working Group I Contribution to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change* (pp. 3–32). Cambridge: Cambridge University Press. <https://dx.doi.org/10.1017/9781009157896.001>
- Ishizuka, S., Ohta, S., Mori, T., Konda, R., Gobara, Y., Hamotani, Y., Kawabata, C., Wicaksono, A., Heriyanto, J., & Hardjono, A. (2021).  $\text{N}_2\text{O}$  emissions in Acacia mangium stands with different ages, in Sumatra, Indonesia. *Forest Ecology and Management*, 498, 119539. <https://doi.org/10.1016/j.foreco.2021.119539>
- Ito, A., Nishina, K., Ishijima, K., Hashimoto, S., & Inatomi, M. (2018). Emissions of nitrous oxide ( $\text{N}_2\text{O}$ ) from soil surfaces and their historical changes in East Asia: A model-based assessment. *Progress in Earth and Planetary Science*, 5, 55. <https://doi.org/10.1186/s40645-018-0215-4>
- Janke, C., Moody, P., Fujinuma, R., & Bell, M. (2022). The impact of banding polymer-coated urea on nitrogen availability and distribution in contrasting soils. *Journal of Soil Science and Plant Nutrition*, 22, 3081–3095. <https://doi.org/10.1007/s42729-022-00869-x>
- Jurado, A., Borges, A. V., & Brouyère, S. (2017). Dynamics and emissions of  $\text{N}_2\text{O}$  in groundwater: A review. *Science of the Total Environment*, 584–585, 207–218. <https://doi.org/10.1016/j.scitotenv.2017.01.127>
- Kim, K., Gil, J., Ostrom, N. E., Gandhi, H., Oerther, M. S., Kuzyakov, Y., Guber, A. K., & Kravchenko, A. N. (2022). Soil pore architecture and rhizosphere legacy define  $\text{N}_2\text{O}$  production in root detritusphere. *Soil Biology and Biochemistry*, 166, 108565. <https://doi.org/10.1016/j.soilbio.2022.108565>
- Krauss, M., Ruser, R., Müller, T., Hansen, S., Mäder, P., & Gattinger, A. (2017). Impact of reduced tillage on greenhouse gas emissions and soil carbon stocks in an organic grass-clover ley—Winter wheat cropping sequence. *Agriculture, Ecosystems and Environment*, 239, 324–333. <https://doi.org/10.1016/j.agee.2017.01.029>
- Kweku, D., Bismark, O., Maxwell, A., Desmond, K., Danso, K., Oti-Mensah, E., Quachie, A., & Adormaa, B. (2018). Greenhouse effect: Greenhouse gases and their impact on global warming. *Journal of Scientific Research and Reports*, 17(6), 1–9. <https://doi.org/10.9734/jsrr/2017/39630>
- Lam, S. K., Suter, H., Mosier, A. R., & Chen, D. (2017). Using nitrification inhibitors to mitigate agricultural  $\text{N}_2\text{O}$  emission: A double-edged sword? *Global Change Biology*, 23(2), 485–489. <https://doi.org/10.1111/gcb.13338>
- Lawrence, N. C., Tenesaca, C. G., VanLooche, A., & Hall, S. J. (2021). Nitrous oxide emissions from agricultural soils challenge climate sustainability in the US Corn Belt. *Proceedings of the National Academy of Sciences of the United States of America*, 118(46), e2112108118. <https://doi.org/10.1073/pnas.2112108118>
- Linn, D. M., & Doran, J. W. (1984). Effect of water-filled pore space on carbon dioxide and nitrous oxide production in tilled and nontilled soils. *Soil Science Society of America Journal*, 48(6), 1267–1272. <https://doi.org/10.2136/sssaj1984.03615995004800060013x>
- Lu, X., Taylor, A. E., Myrold, D. D., & Neufeld, J. D. (2020). Expanding perspectives of soil nitrification to include ammonia-oxidizing archaea and comammox bacteria. *Soil Science*

- Society of America Journal*, 84(2), 287–302. <https://doi.org/10.1002/saj2.20029>
- Lv, L., Gao, Z., Liao, K., Zhu, Q., & Zhu, J. (2023). Impact of conservation tillage on the distribution of soil nutrients with depth. *Soil and Tillage Research*, 225, 105527. <https://doi.org/10.1016/j.still.2022.105527>
- Ma, Y., Schwenke, G., Sun, L., Liu, D. L., Wang, B., & Yang, B. (2018). Modeling the impact of crop rotation with legume on nitrous oxide emissions from rain-fed agricultural systems in Australia under alternative future climate scenarios. *Science of The Total Environment*, 630, 1544–1552. <https://doi.org/10.1016/j.scitotenv.2018.02.322>
- Macdonald, L., Farrell, M., & Baldock, J. (2016). The influence of increasing organic matter content on N<sub>2</sub>O emissions. *Proceedings of the 2016 International Nitrogen Initiative Conference, "Solutions to Improve Nitrogen Use Efficiency for the World," pp. 1–5*. Retrieved from [https://www.researchgate.net/publication/312044643\\_The\\_influence\\_of\\_increasing\\_organic\\_matter\\_content\\_on\\_N2O\\_emissions](https://www.researchgate.net/publication/312044643_The_influence_of_increasing_organic_matter_content_on_N2O_emissions)
- Martínez-Espinosa, R. M., Hatano, R., Wu, Y., & Shaaban, M. (2023). Editorial: Nitrogen dynamics and load in soils. *Frontiers in Environmental Science*, 11, 1197902. <https://doi.org/10.3389/fenvs.2023.1197902>
- Menegat, S., Ledo, A., & Tirado, R. (2022). Greenhouse gas emissions from global production and use of nitrogen synthetic fertilisers in agriculture. *Scientific Reports*, 12, 14490. <https://doi.org/10.1038/s41598-022-18773-w>
- Ministry of Environment and Forestry of Indonesia's. (2022). *Laporan inventarisasi gas rumah kaca (GRK) dan monitoring, pelaporan, verifikasi (MPV) tahun 2021*. Retrieved from <https://signsmart.menlhk.go.id/v2.1/app/frontend/pedomam/detail/44>
- Mondal, S., & Chakraborty, D. (2022). Soil nitrogen status can be improved through no-tillage adoption particularly in the surface soil layer: A global meta-analysis. *Journal of Cleaner Production*, 366, 132874. <https://doi.org/10.1016/j.jclepro.2022.132874>
- Mori, T., Ohta, S., Ishizuka, S., Konda, R., Wicaksono, A., Heriyanto, J., & Hardjono, A. (2010). Effects of phosphorus addition on N<sub>2</sub>O and NO emissions from soils of an Acacia mangium plantation. *Soil Science and Plant Nutrition*, 56(5), 782–788. <https://doi.org/10.1111/j.1747-0765.2010.00501.x>
- Mori, T., Wachrinrat, C., Staporn, D., Meunpong, P., Suebsai, W., Matsubara, K., Boonsri, K., Lumban, W., Kuawong, M., Phukdee, T., Srifai, J., & Boonman, K. (2017). Effects of phosphorus addition on nitrogen cycle and fluxes of N<sub>2</sub>O and CH<sub>4</sub> in tropical tree plantation soils in Thailand. *Agriculture and Natural Resources*, 51(2), 91–95. <https://doi.org/10.1016/j.anres.2016.03.002>
- Netherway, T., Bengtsson, J., Krab, E. J., & Bahram, M. (2021). Biotic interactions with mycorrhizal systems as extended nutrient acquisition strategies shaping forest soil communities and functions. *Basic and Applied Ecology*, 50, 25–42. <https://doi.org/10.1016/j.baae.2020.10.002>
- Nugroho, P. A., Sudadi, U., & Suwardi, S. (2018). Effect of fertilizer management on soil carbon dioxide fluxes in grassland and cornfield during winter. *Journal of Agricultural Science and Technology*, 20(4), 841–853. Retrieved from <http://jast.modares.ac.ir/article-23-19906-en.html>
- Prananto, J. A., Minasny, B., Comeau, L. P., Rudiyanto, R., & Grace, P. (2020). Drainage increases CO<sub>2</sub> and N<sub>2</sub>O emissions from tropical peat soils. *Global Change Biology*, 26(8), 4583–4600. <https://doi.org/10.1111/gcb.15147>
- Prather, M. J., Hsu, J., DeLuca, N. M., Jackman, C. H., Oman, L. D., Douglass, A. R., Fleming, E. L., Strahan, S. E., Steenrod, S. D., Søvde, O. A., Isaksen, I. S. A., Froidevaux, L., & Funke, B. (2015). Measuring and modeling the lifetime of nitrous oxide including its variability. *Journal of Geophysical Research. Atmospheres*, 120(11), 5693–5705. <https://doi.org/10.1002/2015JD023267>
- Qu, W., Suo, L., Liu, R., Liu, M., Zhao, Y., Xia, L., Fan, Y., Zhang, Q., & Gao, Z. (2022). Influence of temperature on denitrification and microbial community structure and diversity: A laboratory study on nitrate removal from groundwater. *Water*, 14(3), 436. <https://doi.org/10.3390/w14030436>
- Rajta, A., Bhatia, R., Setia, H., & Pathania, P. (2020). Role of heterotrophic aerobic



- denitrifying bacteria in nitrate removal from wastewater. *Journal of Applied Microbiology*, 128(5), 1261–1278. <https://doi.org/10.1111/jam.14476>
- Ramzan, S., Rasool, T., Bhat, R., Ahmed, P., Ashraf, I., Rashid, N., Shafiq, M., & Ikhtlaq, M. (2020). Agricultural soils a trigger to nitrous oxide: A persuasive greenhouse gas and its management. *Environmental Monitoring and Assessment*, 192, 436. <https://doi.org/10.1007/s10661-020-08410-2>
- Rapson, T. D., & Dacres, H. (2014). Analytical techniques for measuring nitrous oxide. *TrAC Trends in Analytical Chemistry*, 54, 65–74. <https://doi.org/10.1016/j.trac.2013.11.004>
- Rehman, S. ur, Ijaz, S. S., Raza, M. A., Mohi Ud Din, A., Khan, K. S., Fatima, S., Raza, T., Mehmood, S., Saeed, A., & Ansar, M. (2023). Soil organic carbon sequestration and modeling under conservation tillage and cropping systems in a rainfed agriculture. *European Journal of Agronomy*, 147, 126840. <https://doi.org/10.1016/j.eja.2023.126840>
- Richards, M. B., Butterbach-bahl, K., Jat, M. L., Lipinski, B., Ortiz-Monasterio, I., & Sapkota, T. (2016). Site-specific nutrient management: Implementation guidance for policymakers and investors. *Climate-Smart Agriculture Practice Brief*. Copenhagen, Denmark: CGIAR Research Program on Climate Change, Agriculture and Food Security (CCAFS). Retrieved from <https://cgspace.cgiar.org/handle/10568/69016>
- Ritchie, H., Roser, M., & Rosado, P. (2020). CO<sub>2</sub> and Greenhouse Gas Emissions. *Our World in Data*. Retrieved from <https://ourworldindata.org/greenhouse-gas-emissions>
- Romero, C. M., Hao, X., Li, C., Owens, J., Schwinghamer, T., McAllister, T. A., & Okine, E. (2021). Nutrient retention, availability and greenhouse gas emissions from biochar-fertilized Chernozems. *Catena*, 198, 105046. <https://doi.org/10.1016/j.catena.2020.105046>
- Roy, A. K., Wagner-Riddle, C., Deen, B., Lauzon, J., & Bruulsema, T. (2014). Nitrogen application rate, timing and history effects on nitrous oxide emissions from corn (*Zea mays* L.). *Canadian Journal of Soil Science*, 94(4), 563–573. <https://doi.org/10.4141/CJSS2013-118>
- Ruiz, M. S. M., Reiser, M., & Kranert, M. (2021). Nitrous oxide emission fluxes in coffee plantations during fertilization: A case study in Costa Rica. *Atmosphere*, 12(12), 1656. <https://doi.org/10.3390/atmos12121656>
- Rychel, K., Meurer, K. H. E., Börjesson, G., Strömngren, M., Getahun, G. T., Kirchmann, H., & Kätterer, T. (2020). Deep N fertilizer placement mitigated N<sub>2</sub>O emissions in a Swedish field trial with cereals. *Nutrient Cycling in Agroecosystems*, 118(2), 133–148. <https://doi.org/10.1007/s10705-020-10089-3>
- Salazar, O., Diaz, R., Nario, A., Videla, X., Alonso-Ayuso, M., & Quemada, M. (2021). Nitrogen fertilizer efficiency determined by the 15N dilution technique in maize followed or not by a cover crop in Mediterranean Chile. *Agriculture*, 11(8), 721. <https://doi.org/10.3390/agriculture11080721>
- Santos, W. de M., Alves, B. J. R., Urquiaga, S., Pacheco, E. P., Barros, I. de, Fernandes, M. F., Batista, J. N., Bender, E. P., Souza, H. N. de, & Jantalia, C. P. (2020). Ammonia volatilization and yield of corn fertilized with different nitrogen sources in the Brazilian semiarid. *Pesquisa Agropecuária Brasileira*, 55, e01036. <https://doi.org/10.1590/S1678-3921.pab2020.v55.01036>
- Sarkar, M. I. U., Islam, M. N., Jahan, A., Islam, A., & Biswas, J. C. (2017). Rice straw as a source of potassium for wetland rice cultivation. *Geology, Ecology, and Landscapes*, 1(3), 184–189. <https://doi.org/10.1080/24749508.2017.1361145>
- Senbayram, M., Budai, A., Bol, R., Chadwick, D., Marton, L., Gündogan, R., & Wu, D. (2019). Soil NO<sub>3</sub><sup>-</sup> level and O<sub>2</sub> availability are key factors in controlling N<sub>2</sub>O reduction to N<sub>2</sub> following long-term liming of an acidic sandy soil. *Soil Biology and Biochemistry*, 132(3), 165–173. <https://doi.org/10.1016/j.soilbio.2019.02.009>
- Shaaban, M., Wu, Y., Khalid, M. S., Peng, Q. an, Xu, X., Wu, L., Younas, A., Bashir, S., Mo, Y., Lin, S., Zafar-ul-Hye, M., Abid, M., & Hu, R. (2018). Reduction in soil N<sub>2</sub>O emissions by pH manipulation and enhanced nosZ gene transcription under different water regimes. *Environmental Pollution*, 235, 625–631. <https://doi.org/10.1016/j.envpol.2017.12.066>
- Shcherbak, I., & Robertson, G. P. (2019). Nitrous oxide (N<sub>2</sub>O) emissions from subsurface soils

- of agricultural ecosystems. *Ecosystems*, 22(7), 1650–1663. <https://doi.org/10.1007/s10021-019-00363-z>
- Soares, J. R., Souza, B. R., Mazzetto, A. M., Galdos, M. V., Chadwick, D. R., Campbell, E. E., Jaiswal, D., Oliveira, J. C., Monteiro, L. A., Vianna, M. S., Lamparelli, R. A. C., Figueiredo, G. K. D. A., Sheehan, J. J., & Lynd, L. R. (2023). Mitigation of nitrous oxide emissions in grazing systems through nitrification inhibitors: A meta-analysis. *Nutrient Cycling in Agroecosystems*, 125(3), 359–377. <https://doi.org/10.1007/s10705-022-10256-8>
- Solecki, W., Singh, C., Ley, D., & Revi, A. (2022). *Climate Change 2022. Impacts, Vulnerability and Adaptation. Summary for Policymakers* (Issue IPCC WGII Sixth Assessment Report). Retrieved from <https://www.ipcc.ch/report/ar6/wg2/chapter/summary-for-policymakers/>
- Sosulski, T., Niedziński, T., Jadczyński, T., & Szymańska, M. (2022). Influence of reduced tillage, fertilizer placement, and soil afforestation on CO<sub>2</sub> Emission from arable sandy soils. *Agronomy*, 12(12), 3102. <https://doi.org/10.3390/agronomy12123102>
- Stanton, C. L., Reinhard, C. T., Kasting, J. F., Ostrom, N. E., Haslun, J. A., Lyons, T. W., & Glass, J. B. (2018). Nitrous oxide from chemodenitrification: A possible missing link in the proterozoic greenhouse and the evolution of aerobic respiration. *Geobiology*, 16(6), 597–609. <https://doi.org/10.1111/gbi.12311>
- Sumawinata, B., Djajakirana, G., Suwardi, & Darmawan. (2014). *Carbon dynamics in tropical peatland planted forests (One-year research findings in Sumatra, Indonesia)* (First edition). Bogor: IPB Press. Retrieved from [https://scholar.google.com/scholar?hl=id&as\\_sdt=0%2C5&q=Carbon+Dynamics+In+Tropical+Peatland+Planted+Forests&btnG=](https://scholar.google.com/scholar?hl=id&as_sdt=0%2C5&q=Carbon+Dynamics+In+Tropical+Peatland+Planted+Forests&btnG=)
- Uda, S. K., Hein, L., & Adventa, A. (2020). Towards better use of Indonesian peatlands with paludiculture and low-drainage food crops. *Wetlands Ecology and Management*, 28(3), 509–526. <https://doi.org/10.1007/s11273-020-09728-x>
- Venterea, R. T., Clough, T. J., Coulter, J. A., Breuillin-Sessoms, F., Wang, P., & Sadowsky, M. J. (2015). Ammonium sorption and ammonia inhibition of nitrite-oxidizing bacteria explain contrasting soil N<sub>2</sub>O production. *Scientific Reports*, 5(1), 12153. <https://doi.org/10.1038/srep12153>
- Wang, C., Amon, B., Schulz, K., & Mehdi, B. (2021). Factors that influence nitrous oxide emissions from agricultural soils as well as their representation in simulation models: A review. *Agronomy*, 11(4), 770. <https://doi.org/10.3390/agronomy11040770>
- Wang, F., Li, J., Wang, X., Zhang, W., Zou, B., Neher, D. A., & Li, Z. (2014). Nitrogen and phosphorus addition impact soil N<sub>2</sub>O emission in a secondary tropical forest of South China. *Scientific Reports*, 4, 5615. <https://doi.org/10.1038/srep05615>
- Wang, X., Bai, J., Xie, T., Wang, W., Zhang, G., Yin, S., & Wang, D. (2021). Effects of biological nitrification inhibitors on nitrogen use efficiency and greenhouse gas emissions in agricultural soils: A review. *Ecotoxicology and Environmental Safety*, 220, 112338. <https://doi.org/10.1016/j.ecoenv.2021.112338>
- Weymann, D., Well, R., von der Heide, C., Böttcher, J., Flessa, H., & Duijnisveld, W. H. M. (2009). Recovery of groundwater N<sub>2</sub>O at the soil surface and its contribution to total N<sub>2</sub>O emissions. *Nutrient Cycling in Agroecosystems*, 85(3), 299–312. <https://doi.org/10.1007/s10705-009-9269-4>
- Winkhart, F., Mösl, T., Schmid, H., & Hülsbergen, K. J. (2022). Effects of organic maize cropping systems on nitrogen balances and nitrous oxide emissions. *Agriculture*, 12(7), 907. <https://doi.org/10.3390/agriculture12070907>
- Wrage-Mönnig, N., Horn, M. A., Well, R., Müller, C., Velthof, G., & Oenema, O. (2018). The role of nitrifier denitrification in the production of nitrous oxide revisited. *Soil Biology and Biochemistry*, 123, A3–A16. <https://doi.org/10.1016/j.soilbio.2018.03.020>
- Wu, M., Li, J., Leu, A. O., Erler, D. V., Stark, T., Tyson, G. W., Yuan, Z., McIlroy, S. J., & Guo, J. (2022). Anaerobic oxidation of propane coupled to nitrate reduction by a lineage within the class Symbiobacteriia. *Nature Communications*, 13(1), 6115. <https://doi.org/10.1038/s41467-022-33872-y>
- Xie, L., Li, L., Xie, J., Wang, J., Anwar, S., Du, C., & Zhou, Y. (2022). Substituting inorganic

- fertilizers with organic amendment reduced nitrous oxide emissions by affecting nitrifiers' microbial community. *Land*, *11*(10), 1702. <https://doi.org/10.3390/land11101702>
- Yerli, C., Cakmakci, T., & Sahin, U. (2022). CO<sub>2</sub> emissions and their changes with H<sub>2</sub>O emissions, soil moisture, and temperature during the wetting–drying process of the soil mixed with different biochar materials. *Journal of Water and Climate Change*, *13*(12), 4273–4282. <https://doi.org/10.2166/wcc.2022.293>
- Yoo, J., Woo, S. H., Park, K. Do, & Chung, K. Y. (2016). Effect of no-tillage and conventional tillage practices on the nitrous oxide (N<sub>2</sub>O) emissions in an upland soil: Soil N<sub>2</sub>O emission as affected by the fertilizer applications. *Applied Biological Chemistry*, *59*(6), 787–797. <https://doi.org/10.1007/s13765-016-0226-z>
- Zajac, O., & Zubrowska-Sudol, M. (2022). Nitrification kinetics, N<sub>2</sub>O emission, and energy use in intermittently aerated hybrid reactor under different organic loading rates. *International Journal of Environmental Science and Technology*, 1–14. <https://doi.org/10.1007/s13762-022-04715-6>
- Zhang, W., Zhu, X., Luo, Y., Rafique, R., Chen, H., Huang, J., & Mo, J. (2014). Responses of nitrous oxide emissions to nitrogen and phosphorus additions in two tropical plantations with N-fixing vs. Non-N-fixing tree species. *Biogeosciences*, *11*(18), 4941–4951. <https://doi.org/10.5194/bg-11-4941-2014>
- Zhou, H., Shi, H.-B., Guo, J.-W., Zhang, W.-C., & Wang, W.-G. (2020). Effects of the combined application of organic and inorganic fertilizers on N<sub>2</sub>O emissions from saline soil. *Environmental Science*, *41*(8), 3811–3821. <https://doi.org/10.13227/j.hjcx.202002046>
- Zhu, Y., Butterbach-Bahl, K., Merbold, L., Leitner, S., & Pelster, D. E. (2021). Nitrous oxide emission factors for cattle dung and urine deposited onto tropical pastures: A review of field-based studies. *Agriculture, Ecosystems & Environment*, *322*, 107637. <https://doi.org/10.1016/j.agee.2021.107637>