



## Physiological Responses of Indigenous Vegetable of Sintrong (*Crassocephalum crepidioides*) due to Exposure to High Temperature

Ulfiana Mahira Rahma Utama<sup>1</sup>, Ani Kurniawati<sup>1\*</sup> and Didah Nur Faridah<sup>2</sup>

<sup>1</sup>Department of Agronomy and Horticulture, Faculty of Agriculture, IPB University, Bogor, Indonesia;

<sup>2</sup>Department of Food and Technology, Faculty of Agricultural Technology, IPB University, Bogor, Indonesia

\*Corresponding author: [ani\\_kurniawati@apps.ipb.ac.id](mailto:ani_kurniawati@apps.ipb.ac.id)

### Abstract

Sintrong is an Indonesian indigenous vegetable with leaves used for vegetables, digestive disorders, and burns. Changes in the environment due to an increase in temperature affect the growth and quality of sitrong, and its existence in the nature is threatened. This study aims to obtain information about the effect of exposure to high temperatures on the physiological character and flavonoid content of indigenous sintrong vegetables and obtain accession of sintrong, which can be developed as a functional vegetable. The Nested randomized group design was applied with two factors, temperature differences as the main plot and accession as a second plot. Four replications were conducted for each accession in the Cikabayan experimental garden of IPB. The results showed that exposure to high temperatures up to 32 °C increased the speed of flowering age, which was 4.76% and 7.14% faster and showed a high wilting rate of 36.66%, but decreased leaf area index up to 30.30% and 42.42% at the conditions above ambient temperature exposure (control). Flavonoid content did not show any effect due to exposure to high temperatures. The flavonoid content reached 1695.38 and 1834.83 mg QE 100 g<sup>-1</sup>. Bogor 1 accession showed the best performance so that the plants can be developed for functional vegetables. Based on the research findings, sintrong should be harvested earlier before flowering to obtain high leaf production and good-quality vegetables.

**Keywords:** anthocyanins; flavonoid; heat stress; stomata conductance; vegetables

**Cite this as:** Utama, U. M. R., Kurniawati, A., & Faridah, D. N. (2023). Physiological Responses of Indigenous Vegetable of Sintrong (*Crassocephalum crepidioides*) due to Exposure to High Temperature. *Caraka Tani: Journal of Sustainable Agriculture*, 38(1), 163-175. doi: <http://dx.doi.org/10.20961/carakatani.v38i1.67163>

### INTRODUCTION

Sintrong (*Crassocephalum crepidioides*) is one of the indigenous vegetables. The part of sintrong that is commonly used is the leaf. Adjatin et al. (2013) conducted the phytochemical screening of sintrong and reported that the leaves contain cathectic tannins, gallic tannins, coumarins, flavonoids and steroids. Sintrong leaves are used traditionally to treat digestive disorders and burns (Can et al., 2020). Sintrong has anti-bacterial (Owokotomo, 2018), antioxidant (Aniya et al., 2005), anti-inflammatory (Akinpelu et al., 2019), antitumor

(Tomimori et al., 2012) and antidiabetic activity (Bahar et al., 2017).

Climate change is a threat to nutrition and food security (Tanimonure, 2021), especially food that is still harvested from the wild. Global climate change is indicated by an increase in the average temperature of the earth's surface that reached 0.87 °C in 2006 to 2015 (IPCC, 2021). The increase in average temperature at the end of the 21<sup>st</sup> century is relatively larger, between 2.4 and 3.5 °C (IPCC, 2022). The development of sintrong, which is an indigenous vegetable, is a challenge because some of them are not cultivated. Environmental changes due to

\* Received for publication November 10, 2022  
Accepted after corrections January 21, 2023

increasing temperatures will affect the growth and quality of vegetables and will even threaten their existence in nature. Vegetables have different responses to changes in temperature, depending on each plant species (Bashandy and El-Shaieny, 2021). High temperatures affect all phases of plant growth and metabolism (Sailaja et al., 2014).

High temperature is a significant limiting factor in supporting the productivity and quality of rice harvests (Liu et al., 2013). Jaisyurahman et al. (2020) reported that high temperatures 32.72 °C after the generative phase in rice affected the character of the total number of tillers, the number of filled grains per panicle, the number of empty grains per panicle, the total number of grains per panicle, the rate of seed filling, the percentage of filled grain and the total grain weight per plant. Heat stress over a long period of time in several tomato cultivars resulted in a significant decrease in all reproductive traits, such as pollen viability, pollen count, mud fertility, fruit set and number of flowers per flowering (Xu et al., 2016). Alhaithloul et al. (2021) reported that the identification of total phenolics, flavonoids and the abundance of secondary metabolites in tomato seedlings was associated with thermal stress. This accumulation is a defense mechanism due to thermal stress.

The impact of exposure to high temperatures can affect the availability of nutritious food for the community, especially food available in nature, because its growth depends entirely on the environment (Giulia et al., 2020). Conditions of increasing temperature above the tolerance limit can threaten the food sustainability. This study examined the effect of temperatures above the ambient temperature on the physiological character of the sintrong. This study will provide information about the effect of temperature exposure on physiological and flavonoid characteristics, as well as the best accessions that are able to adapt to exposure to high temperature.

## MATERIALS AND METHOD

The field research was conducted at 6°33'01" S and 106°42'51" E Cikabayan experimental field, IPB University, using UV house. Analysis of flavonoid content was conducted at Post Harvest Laboratory of Department of Agronomy and Horticulture, Faculty of Agriculture, IPB University, from May to September 2022. High-temperature condition was obtained in

a modification of the UV house building on the percentage of wall covering so as to get the temperature range based on the treatment. Temperature measurement was carried out using a thermo recorder. Thermo recorder can record temperature every 30 minutes. Average daily temperature was obtained from the average air temperature at 00:00 to 23:30 during treatment period. The maximum temperature was obtained from the highest daily average temperature and the minimum temperature was obtained from the lowest daily average temperature during the period. This study used a nested randomized block design with two factors of treatment, namely temperature difference as the main plot and accession as the second plot. The temperature difference consists of three levels, including the ambient temperature (T1), the daily average air temperature in the UV house with an average of 30±2 °C (T2), and the average daily air temperature in the UV house with an average of 32±2 °C (T3). Plant accessions consist of Bogor 1 (Bogor, lowland), Bogor 2 (Bogor, highland) and Cianjur 1 (Cianjur, lowland), based on the seeds. At each accession level, four replications were conducted to obtain 12 experimental units. One experimental unit was a plot of 1.5 x 1.5 m<sup>2</sup>, with a population per plot of 25 plants with a spacing of 20 x 20 cm<sup>2</sup>, so that the total plants were 900. High-temperature treatment was provided from transplanting stage until 45 days after planting (DAP). Plant maintenance includes watering, weeding, and fertilizing. Sintrong was harvested at 45 DAP.

Observation was conducted at flowering stage, at the age of full flowering of 50%. The wilting rate was observed by assessing the percentage of leaf fall. Leaf area index (LAI) was measured by the total leaf area on the area of land covered by the plant. The leaf area was measured with ImageJ. Relative growth rate (RGR) was measured at 30 and 40 DAP using the Hoffman and Poorter (2002) and pigment content was analyzed using Sims and Gamon (2004). The rate of photosynthesis, stomatal conductance, and transpiration was measured using the Licor-6400 XT portable photosynthesis system. Anatomical observations include the density of stomata and trichomes using an Olympus CX 23 led binocular microscope.

The preparation of flavonoid sample was conducted with a sample of dried leaves at a temperature of 60 °C for 24 hours. The sample

was mashed and sifted, and then stored in a container with silica gel. A sample of 0.01 g was weighed for extraction analysis and powder moisture content was established by weighing. The samples of 1 to 2 g were then ventilated at 105 °C for 5 hours. Extraction was conducted by taking 2 g of simplicial powder and 70% ethanol solvent, homogenized using a vortex for 15 minutes, then sonicated with sonicator for 15 minutes and macerated for 24 hours in dark space, centrifuged for 15 minutes to produce filtrate. The filtrate was then mixed and evaporated until the extract volume was 10 ml (Istiqomah, 2020). The flavonoid content was analyzed by the  $AlCl_3$  method (Pothitirat et al., 2009). Briefly, 5 ml of 2% aluminum trichloride in methanol was mixed with the same volume of sample ( $500 \mu\text{g ml}^{-1}$ ). Absorption reading at 415 nm was done after 10 minutes against a blank sample without aluminium trichloride using UV-visible spectrophotometer Shimadzu Type UV-1280. The standard used for the analysis of flavonoids was quercetin with the standard curve at concentrations of 2, 5, 25, 50 and  $75 \text{ mg l}^{-1}$ . The total flavonoid content was counted as grams of quercetin equivalents (QE)  $100 \text{ g}^{-1}$  of the extract (Vongsak et al., 2013). Collected data were analyzed statistically according to one-way analysis of variance (ANOVA) technique. If the treatment gave statistically significant effect ( $p < 0.05$ ), the different sources of variation were compared using Duncan's multiple range test (DMRT) using SAS 9.0.

## RESULTS AND DISCUSSION

Temperatures above ambient have a significant effect on flowering age, wilting rate, LAI and RGR of sintrong plants (Table 1).

The flowering ages of the sintrong at  $30.49 \pm 0.25$  and  $32.23 \pm 0.28$  °C were faster, 4.76% and 7.14% at above ambient temperature exposure conditions. The flowering ages at  $28.89 \pm 0.40$ ,  $30.49 \pm 0.25$ , and  $32.23 \pm 0.28$  °C were respectively 42, 40 and 39 days. Flowering age was calculated when the flowers first appeared on the sintrong based on nine samples of plant each plot. The transition from the vegetative to the generative phase is an important event; the flowering age has an important role in plant reproduction and the ability to adapt to the environment (Guirao et al., 2019). Factors that affect flowering time include vernalization, photoperiodicity, autonomic and gibberellin pathway (Mouradov et al., 2002). The flowering signals appear on the leaves and then transmitted to the apical meristems in the shoots where they flower (Tsoy and Mushegian, 2022). High temperatures will trigger a flowering signal so that flowering will occur more quickly. The regulatory gene in flowering is *Flowering Locus C* (FLC). FLC will encode protein for flowering (Xu et al., 2016). In addition, high temperatures reduce the flowering period, which in turn reduce the accumulation of dry matter in *Tanacetum cinerariifolium* (Suraweera et al., 2020).

The exposure of sintrong to high temperature shows a wilting response with the characteristics of the leaves falling or drooping. The temperature treatment of  $32.23 \pm 0.28$  °C showed a high wilting rate of 36.66% (Table 2). This is presumably because exposure to high temperatures causes the plant to lose a lot of water due to a faster transpiration rate. Guo et al. (2022) reported that heat stress in tomato plants induces an increase in the rate of transpiration, which results in dehydration of plant organs so that the leaves fall down and wither.

Table 1. Effect of high temperature and accession on the flowering age of sintrong, LAI and RGR

Treatment	Flowering age (days)	Wilting rate (%)	LAI	RGR ( $\text{g day}^{-1}$ )
Temperature (°C)				
T1 ( $28.89 \pm 0.40$ )	$42 \pm 0.50\text{a}$	$9.6 \pm 6.31\text{c}$	$3.3 \pm 0.34\text{a}$	$0.28 \pm 0.03\text{a}$
T2 ( $30.49 \pm 0.25$ )	$40 \pm 0.27\text{b}$	$22.05 \pm 7.66\text{b}$	$2.3 \pm 0.30\text{b}$	$0.19 \pm 0.04\text{b}$
T3 ( $32.23 \pm 0.28$ )	$39 \pm 0.62\text{c}$	$36.66 \pm 8.17\text{a}$	$1.9 \pm 0.45\text{c}$	$0.15 \pm 0.02\text{c}$
Accession				
Bogor 1	$41 \pm 1.64$	$24.40 \pm 11.31$	$2.78 \pm 0.59\text{a}$	$0.24 \pm 0.08$
Bogor 2	$41 \pm 1.25$	$21.89 \pm 13.65$	$2.35 \pm 0.71\text{b}$	$0.20 \pm 0.07$
Cianjur 1	$41 \pm 1.37$	$22.10 \pm 13.54$	$2.48 \pm 0.62\text{b}$	$0.18 \pm 0.07$

Note: Numbers followed by the same letter in the same column show no significant difference based on DMRT at level of 5%

Table 2. Effect of high temperature and accession on leaf pigment content

Treatment	Pigment content (mg g <sup>-1</sup> )				
	Chlorophyll a	Chlorophyll b	Total chlorophyll	Carotenoids	Anthocyanins
Temperature (°C)					
T1 (28.89±0.40)	1.06±0.08b	0.37±0.03b	1.43±0.11b	0.37±0.02b	0.02±0.005b
T2 (30.49±0.25)	1.09±0.12b	0.39±0.05b	1.48±0.17b	0.38±0.04b	0.03±0.004b
T3 (32.23±0.28)	1.27±0.13a	0.47±0.05a	1.75±0.18a	0.43±0.03a	0.05±0.01a
Accession					
Bogor 1	1.17±0.11a	0.42±0.04a	1.60±0.15a	0.40±0.03a	0.03±0.01
Bogor 2	1.19±0.13a	0.38±0.06b	1.45±0.19b	0.37±0.04b	0.03±0.02
Cianjur 1	1.06±0.17b	0.42±0.06a	1.61±0.23a	0.41±0.05a	0.03±0.01

Note: Numbers followed by the same letter in the same column show no significant difference based on DMRT at level of 5%

LAI is the ratio of the total leaf area of the plant to the projected area covered. The ambient temperature treatments with 30.49±0.25 and 32.23±0.28 °C showed LAI of 3.3, 2.3, and 1.9, respectively. The percentage of decrease in LAI above the ambient temperature of 30.49±0.25 and 32.23±0.28 °C was 30.30% and 42.42%, respectively (Table 2). The LAI decreases and the temperature increases. High LAI values at ambient temperature indicate that young leaves on the shoots are able to absorb large amounts of radiation and high CO<sub>2</sub> assimilation rate and translocate to other plant parts in large quantities. High temperatures with an average of 35.3 °C reduced LAI in peas, and LAI value in Arka Chaitra variety peas aged 60 days after sowing was 0.88 (Verma, 2019). Exposure to high temperatures also decreased LAI faster due to the presence of wheat leaf senescence. Senescent induces a signal of seed maturity so that the plant will complete its life cycle faster (Chen et al., 2017). LAI is influenced by leaf distribution and density, which are related to plant spacing and population.

The closer the spacing, the higher the leaf density and the lower the exposure to sunlight reaching the lower leaf layers so LAI value increases (Du et al., 2022). The inhibition of leaf expansion will have an impact on decreasing the capacity of the leaves to absorb light, so that it will minimize the performance of the rate of photosynthesis in plants.

RGR observations were at the age of 30 (vegetative phase) and 40 DAP (maximum vegetative phase) with destructively weighing the dry weight of roots, stems and leaves at 80 °C for 2 x 24 hours. Exposure to high temperature decreased RGR compared to ambient temperature. The RGR value at 30.49±0.25 °C decreased by 32.14% while at 32.23±0.28 °C

decreased by 46.42% (Table 2). This decrease is thought to be due to changes in photosynthate allocation (Hoffmann and Poorter, 2002). RGR observations were carried out at the age of 40 DAP, when the sintrong entered its maximum vegetative state, so that the accumulation of photosynthate was no longer focused on vegetative growth, which would reduce the RGR. RGR was also lower during the stress phase in heat-sensitive wheat genotypes (Abdelhakim et al., 2021).

Differences in accessions did not affect flowering age, wilting rate and RGR, but differences in accessions affected LAI. Accessions of Bogor 1, Bogor 2 and Cianjur 1 have the same flowering age of 41 days. Bogor 1 has the largest LAI value of 2.78, followed by Cianjur 1 and Bogor 2, which are not significantly different. This is presumably because Bogor 1 comes from the lowlands and is shaded, so it has a wider leaf area than other accession. The higher leaf area will increase the LAI value.

Exposure to high temperature had a significant effect on chlorophyll a, chlorophyll b, total chlorophyll, carotenoids and anthocyanins (Table 2). Chlorophyll a and b increased with increasing temperature. The increases in chlorophyll a and b at a temperature of 32.23±0.28 °C were 16.53% and 21.27%, respectively. Chlorophyll a is in the reaction center of photosystems I and II, and in the antenna pigment, while chlorophyll b is found in the antenna pigment, only which is useful for capturing light (Handayani et al., 2013). Exposure to high temperatures caused the loss of pigment content in flag leaves, by reducing the chlorophyll a+b content in two wheat cultivars JM 22 and XM 26 by 1.031 and 0.778 mg ml<sup>-1</sup> g FW<sup>-1</sup> (Feng et al., 2014). The results of this study showed that chlorophyll a and b increased along

with plants exposed to higher than ambient temperatures. This is presumably because the sintrong plant is tolerant to high-temperature exposure conditioned in this study. In addition, it is suspected that the sintrong plant is found in the nature so that it has a good level of adaptation to high temperatures as evidenced by the increase in chlorophyll. The low decrease in chlorophyll content, the ratio of chlorophyll a:b, and total chlorophyll indicate the tolerance to high temperatures (Almeselmani and Viswanathan, 2012).

Carotenoids are terpenoid compounds that become photosynthetic pigments with a color effect between red and yellow. Violaxanthin is a member of the carotenoids found in the chloroplast membrane, which causes a yellow color (Zaripheh and Erdman, 2002). High temperatures increase the carotenoid content in sintrong plants. The carotenoid content at  $32.23\pm 0.28$  °C was significantly different with those at  $28.89\pm 0.40$  and  $30.49\pm 0.25$  °C. The increase in carotenoids was at the temperature of  $32.23\pm 0.28$  °C when compared to the ambient temperature of 12.5%. However, in this study, the increase in carotenoids is a form of plant defense against photooxidation damage at high-temperature conditions (Rossi and Huang, 2022).

Anthocyanins belong to the class of flavonoid compounds that are included in the group of natural pigments in plants dissolved in water, which function to give color to flowers, fruits and vegetables (Maulid and Laily, 2015). The anthocyanin content at ambient temperature of  $30.49\pm 0.25$  °C was not significantly different, but at  $32.23\pm 0.28$  °C, it was significantly different. The increase in anthocyanins at a temperature of  $32.23\pm 0.28$  °C was by 60%, compared to those at ambient temperatures. Anthocyanins function as specific light shields that absorb visible and UV radiation in the vacuole and prevent UV rays from penetrating

into the tissue (Subira et al., 2021). High anthocyanin content can increase absorption and tolerance to UV radiation and increase its antioxidant capacity.

The difference in accessions showed a significant effect on chlorophyll a, chlorophyll b, total chlorophyll and carotenoids. Chlorophyll a contained in Bogor 1 and Bogor 2 was higher than that in Cianjur 1. Leaf surface area streamlines the capture of light energy for photosynthesis (Dharmadewi, 2020). It is considered that Bogor 1 and Bogor 2 accessions are able to harvest higher light energy so that the chlorophyll a content is higher because of a wide leaf area. Chlorophyll b contained in Bogor 1 and Cianjur 1 accessions was higher than that in Bogor 2. Bogor 1 and Cianjur 1 accessions had higher chlorophyll b content, presumably because Bogor 1 and Cianjur 1 were accessioned from the lowlands. In the lowlands, light exposure is more optimal than in the highlands. Bogor 2 has low chlorophyll b, which reduces the total chlorophyll. This is so because Bogor 2 was from highland. The total chlorophyll Bogor 1 and Cianjur 1 accessions were higher than Bogor 2. High total chlorophyll will encourage plants to carry out photosynthesis more optimally.

Exposure to high temperature has a significant effect on the rate of photosynthesis, stomata conductance and transpiration (Table 3). Photosynthesis rate at temperature  $32.2\pm 0.28$  °C was 2.52% higher when compared to the rate at ambient temperature  $28.89\pm 0.40$  °C with a photosynthetic rate of  $31.05 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ , while at a temperature of  $30.49\pm 0.25$  °C, the photosynthesis rate increased 8.61%, reaching  $27.16 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ . The rate of photosynthesis is sensitive to the changes in temperature, which will result in an imbalance of energy in the cell. High temperature affects the photochemical reactions in thylakoids and carbon metabolism in the stroma affects each

Table 3. Effect of high temperature and accession on photosynthesis rate, stomatal conductance and transpiration

Treatment	Photosynthesis rate ( $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ )	Stomatal conductance ( $\text{mmol H}_2\text{O m}^{-2} \text{ s}^{-1}$ )	Transpiration rate ( $\text{mmol H}_2\text{O m}^{-2} \text{ s}^{-1}$ )
Temperature (°C)			
T1 ( $28.89\pm 0.40$ )	$24.82\pm 1.09\text{c}$	$1.19\pm 0.36\text{a}$	$3.36\pm 0.50\text{b}$
T2 ( $30.49\pm 0.25$ )	$27.16\pm 0.80\text{b}$	$0.73\pm 0.13\text{b}$	$4.95\pm 0.45\text{a}$
T3 ( $32.23\pm 0.28$ )	$31.05\pm 1.64\text{a}$	$0.69\pm 0.13\text{b}$	$5.05\pm 0.75\text{a}$
Accession			
Bogor 1	$28.21\pm 3.00$	$0.82\pm 0.29$	$4.58\pm 0.60$
Bogor 2	$27.15\pm 2.81$	$0.93\pm 0.32$	$4.48\pm 1.26$
Cianjur 1	$27.66\pm 2.71$	$0.87\pm 0.35$	$4.58\pm 0.92$

Note: Numbers followed by the same letter in the same column show no significant difference based on DMRT at level of 5%

other (Wang et al., 2018). The application of sulfur in the conditions with an increase in high temperature in canola showed a photosynthetic rate of  $6.12 \text{ mol m}^{-2} \text{ s}^{-1}$  (Waraich et al., 2021). High temperature can damage photosynthetic apparatus, such as PS I, PS II, *cytochrome b6f* (*Cytb6f*) complex and rubisco inactivation (Hu et al., 2020). In this study, the opposite applies to high temperatures, in which the rate of photosynthesis increases. This is presumably due to the availability of sufficient water during the growth so that it can compensate for water loss due to increased transpiration.

Stomatal conductance describes the supply of  $\text{CO}_2$  from the atmosphere to the intercellular space of the cell. Stomatal conductance at  $30.49 \pm 0.25$  and  $32.23 \pm 0.28$  °C was not significantly different. Stomatal conductance at temperatures above ambient  $30.49 \pm 0.25$  and  $32.23 \pm 0.28$  °C was  $0.73$  and  $0.69 \text{ } \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ . Stomatal conductance decreases with increasing temperature and this is presumably because at high temperatures, the stomata will close to prevent water loss due to high transpiration (Anggraini et al., 2016). The high temperature inside the leaves results in a decrease in stomatal conductance in olives and this decrease in stomatal conductance aims to minimize the risk of xylem embolism. Similar to the study on *Cyamopsis tetragonoloba* with a high temperature of 42 °C, the stomatal conductance value was reduced to  $45 \text{ mmol m}^{-2} \text{ s}^{-1}$ , lower than the control of  $49 \text{ } \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$  (Alshameri et al., 2019). At high temperatures, low water availability and high evapotranspiration favor a decrease in stomatal conductance because a higher transpiration rate results in lower xylem vessel pressure (Haworth et al., 2018).

The transpiration process starts from the absorption of groundwater by plant roots. The water was transported through the stems to

the atmosphere. Transpiration increases with increasing temperature. Transpiration rate of sintrong plants exposed above ambient temperatures of  $30.49 \pm 0.25$  and  $32.23 \pm 0.28$  °C increased, compared to the ambient temperature with an increase of 32.12% and 33.46%. The water absorbed by plant roots from the soil is not fully used to produce dry matter, because most of the total water absorbed by the roots will be lost through the transpiration process. The main driving force of the transpiration process is the water potential gradient between the space in the stomata and the atmospheric air (Hidayati and Anas, 2016).

Exposure to high temperature had a significant effect on stomatal and trichome density; differences in accessions did not show a significant effect on stomatal and trichome density (Table 4; Figure 1 and 2). Stomata density increased as it increased above ambient temperature at  $30.49 \pm 0.25$  and  $32.23 \pm 0.28$  °C, with a stomata density of  $127 \text{ mm}^{-2}$ . The structure of stomata functions for the exchange of gases  $\text{O}_2$ ,  $\text{CO}_2$ , and water vapor, which are highly important in the processes of photosynthesis, respiration and transpiration. The level of stomatal density is classified as low ( $< 300 \text{ mm}^{-2}$ ), medium ( $300$  to  $500 \text{ mm}^{-2}$ ), and high ( $> 500 \text{ mm}^{-2}$ ) (Marantika et al., 2021). From this category, the sintrong in all treatments belonged to the low-density category (density  $< 300 \text{ mm}^{-2}$ ). Stomata density is related to stomata size; if the density is high, the stomata usually has a small size, or vice versa. High density and number of stomata are the processes of plant adaptation to environmental conditions (Carrera et al., 2021). Trichomes are derived from the epidermis in the form of hair. At high temperatures, the trichomes increase because they function to prevent excessive evaporation of the leaves. This condition is mainly

Table 4. Effect of high temperature and accession on stomatal and trichome density

Treatment (°C)	Density ( $\text{mm}^{-2}$ )	
	Stomata	Trichome
Temperature		
T1 ( $28.89 \pm 0.40$ )	$82 \pm 12\text{b}$	$3 \pm 0.39\text{c}$
T2 ( $30.49 \pm 0.25$ )	$127 \pm 32\text{a}$	$4 \pm 0.54\text{b}$
T3 ( $32.23 \pm 0.28$ )	$127 \pm 30\text{a}$	$5 \pm 0.62\text{a}$
Accession		
Bogor 1	$114 \pm 37$	$4 \pm 1.24$
Bogor 2	$107 \pm 26$	$4 \pm 0.97$
Cianjur 1	$114 \pm 35$	$4 \pm 0.95$

Note: Numbers followed by the same letter in the same column show no significant difference based on DMRT at level of 5%

the leaves and released as water vapor into

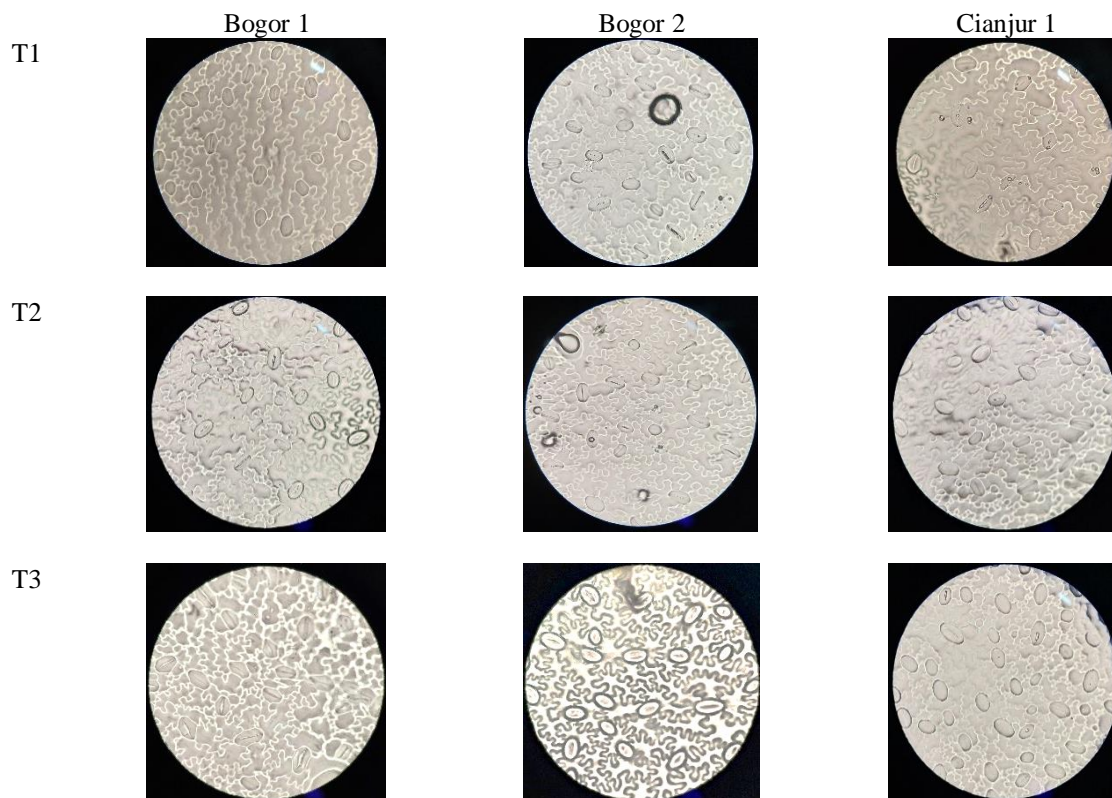


Figure 1. Stomata density of abaxial leaf (T1 =  $28.89 \pm 0.40$  °C, T2 =  $30.49 \pm 0.25$  °C, T3 =  $32.23 \pm 0.28$  °C)

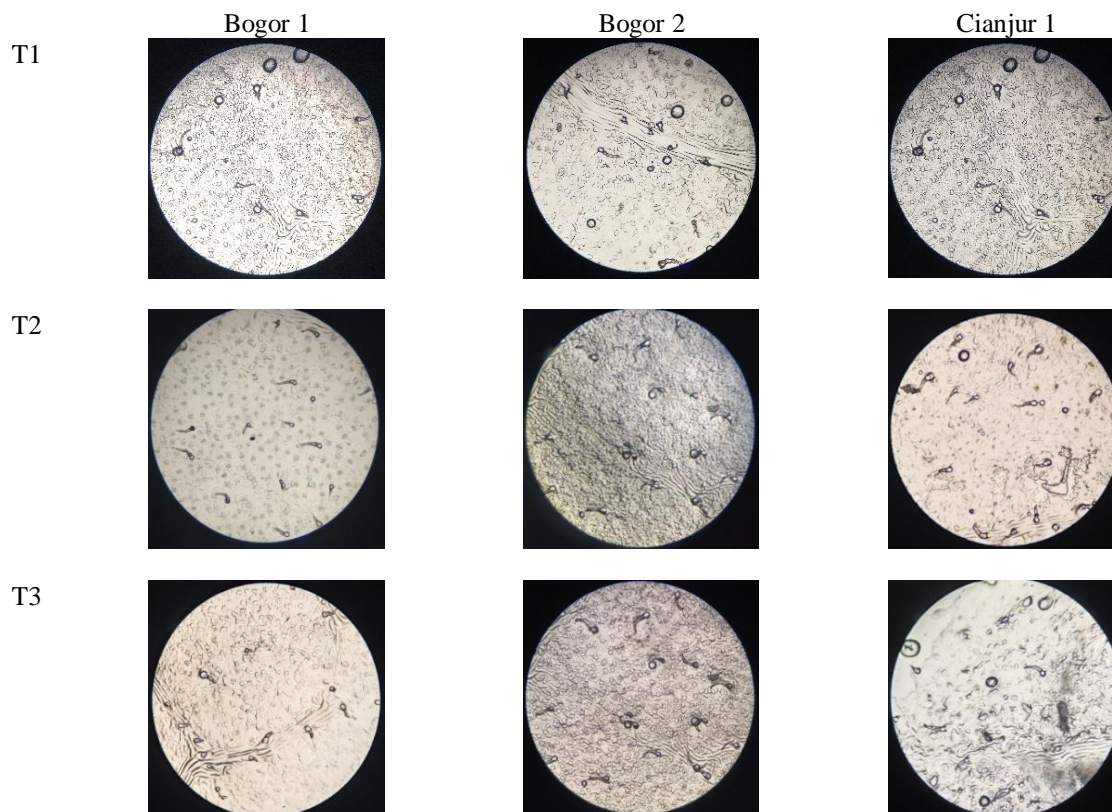


Figure 2. Trichome density of abaxial leaves (T1 =  $28.89 \pm 0.40$  °C, T2 =  $30.49 \pm 0.25$  °C, T3 =  $32.23 \pm 0.28$  °C)

due to the non-linear response of photosynthesis rate to temperature in *Metrosideros polymorpha*. Increasing leaf temperature affects trichomes, which can increase the activity of photosynthetic enzymes; leaf trichomes can increase the rate of photosynthesis (Amada et al., 2020).

The flavonoid content is expressed in mg 100 g<sup>-1</sup> dry matter. Curve standard quercetin showed linear equation with R<sup>2</sup> value = 0.9985 (Figure 4). In this study, exposure to high temperatures up to 32 °C and accession had no effect on the flavonoid content (Figure 3). Flavonoids are part of the phenolic group, which have antioxidant activity and are found in many plants. The content of flavonoids at a temperature higher than the ambient temperature was 30.49±0.25 and 32.23±0.28 °C in a row by

1695.38 and 1834.83 mg 100 g<sup>-1</sup> of dry matter. High temperatures result in an increase in free radicals in the form of reactive oxygen species (ROS), which is reactive in plant tissues and can trigger cell damage. Plants that are tolerant to ROS will adapt to such condition by producing antioxidant compounds (Slimen et al., 2014). In this study, flavonoids did not show any effect due to exposure to high temperatures, presumably because the groundwater status was always maintained at field capacity. In addition, it is suspected that the sintrong plant has a good tolerance for high temperatures. These results are supported by research by Obaid et al. (2016) that the flavonoid 3-monooxygenase and flavonoid 3-dioxygenase enzymes in the flavonoid biosynthetic pathway are inactive

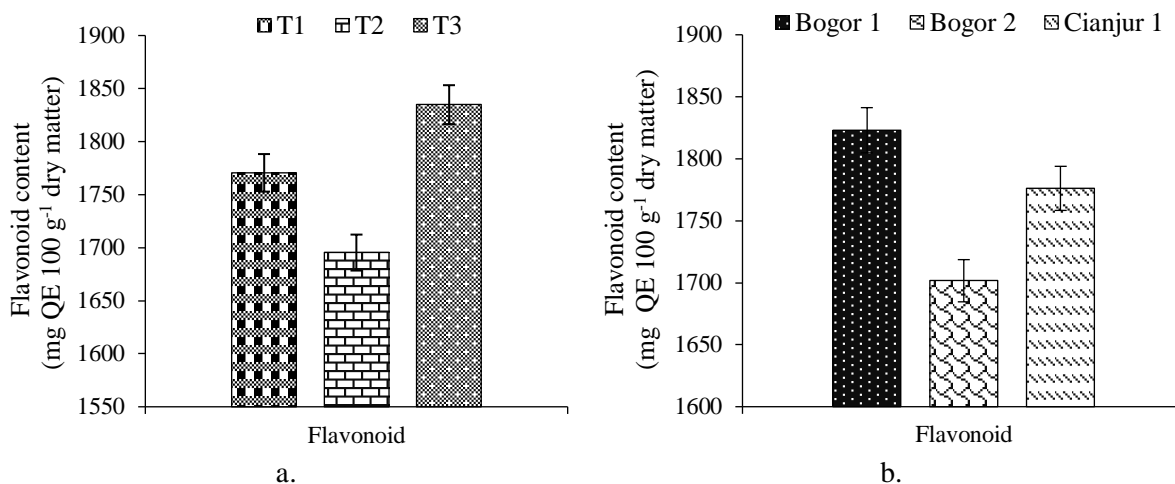


Figure 3. Content of flavonoid, (a) effect of high-temperature T1 = 28.89±0.40 °C, T2 = 30.49±0.25 °C, and T3 = 32.23±0.28 °C; (b) effect of accessions. Bar = standard deviation; n = 4

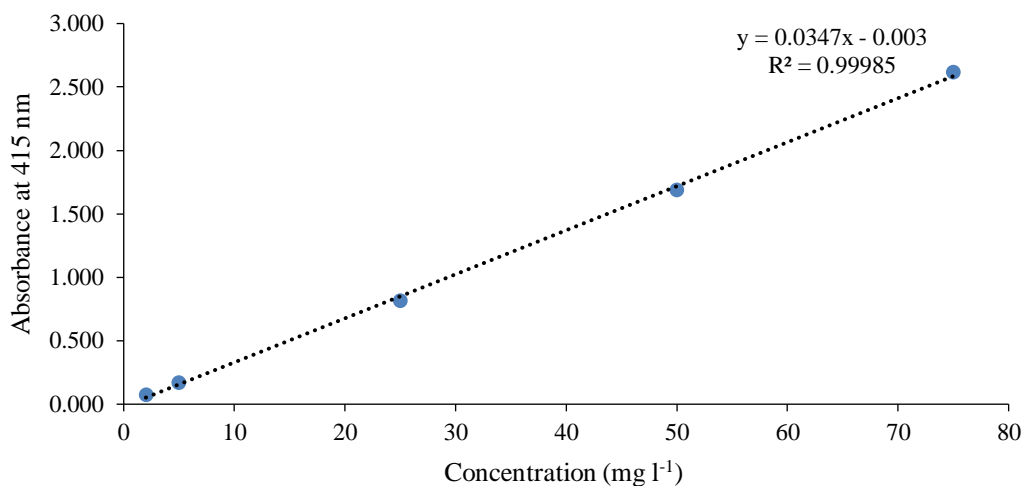


Figure 4. Curve standard of quercetin



Table 5. Pearson correlation test between parameters

Parameters	Flowering age	Wilting rate	LAI	RGR	Photosynthesis rate	Stomata conductance	Transpiration rate	Chlorophyll a	Chlorophyll b	Total chlorophyll	Carotenoids	Anthocyanins	Stomata	Trichome	Flavonoid	
Flowering age	1															
Wilting rate		1														
LAI			1													
RGR				1												
Photosynthesis rate					1											
Stomata conductance						1										
Transpiration rate							1									
Chlorophyll a								1								
Chlorophyll b									1							
Total chlorophyll										1						
Carotenoids											1					
Anthocyanins												1				
Stomata													1			
Trichome														1		
Flavonoid															1	

at 40 to 42 °C in young leaves of *Rhazya stricta*. Inactivity of the enzyme will reduce the total flavonoid content.

Based on Pearson's correlation among all parameters, several parameters were strongly correlated ( $r > 0.5$ ), with positive and negative correlations (Table 5). The leaf area index was positively correlated with the relative growth rate and transpiration rate with the value of  $r > -0.5$ , but negatively correlated with the photosynthesis rate and trichome density. The increasing leaf area index will decrease the rate of photosynthesis. This is because leaves shade each other so that the lower leaves cannot conduct photosynthesis at optimal level (Santrum et al., 2021). The rate of photosynthesis was positively correlated with stomatal conductance, chlorophyll a, chlorophyll b, total chlorophyll, carotenoids, anthocyanins, and trichome density with the  $r$  value of  $> 0.5$ . The transpiration rate was negatively correlated with the density of trichomes with  $r$  value of  $> -0.5$ . This means that the higher the trichome density decreases the transpiration rate. Meanwhile, total chlorophyll was positively correlated with carotenoids, and anthocyanins with values of  $r > 0.5$ . This signifies that higher chlorophyll also contributes to higher anthocyanins and carotenoids. Stomata was positively correlated with trichomes with the  $r$  value of  $> 0.5$ . No correlation ( $r > 0.5$ ) was found between flavonoids with all parameters.

### CONCLUSIONS

Exposure to high temperatures between 30 and 32 °C increases the speed of flowering age, wilting rate, pigment content (chlorophyll a, chlorophyll b, total chlorophyll, anthocyanins and carotenoids), photosynthesis rate, transpiration rate, stomatal density, and trichomes, but decreases leaf area index, relative growth rate and stomatal conductance. The increasing temperature does not affect the flavonoid content in the leaves of sintrong. Therefore, sintrong should be harvested earlier before flowering to obtain high leaf production and good-quality vegetables. Bogor 1 accession has shown the best performance, so it can be developed for functional vegetables and needs further research to determine its nutritional content.

### ACKNOWLEDGEMENT

The authors would like to thank the Ministry of Education, Culture, Research, and Technology

for funding this research through the PTM scheme in 2022 with the contract number of 3831/IT3LI/PT.01.03/P/B/2022.

## REFERENCES

- Abdelhakim, L. O. A., Rosenqvist, E., Wollenweber, B., Spyroglou, I., Ottosen, C. O., & Panzarová, K. (2021). Investigating combined drought- and heat stress effects in wheat under controlled conditions by dynamic image-based phenotyping. *Agronomy*, *11*(2), 364. <https://doi.org/10.3390/agronomy11020364>
- Adjatin, A., Dansi, A., Badoussi, E., Sanoussi, A., Dansi, M., Azokpota, P., Ahissou, H., Akouegninou, A., Akpagana, K., & Sanni, A. (2013). Proximate, mineral and vitamin C composition of vegetable Gbolo [*Crassocephalum rubens* (Juss. ex Jacq.) S. Moore and *C. crepidioides* (Benth.) S. Moore] in Benin. *International Journal of Biology and Chemical Sciences*, *7*(1), 319–331. <https://doi.org/10.4314/ijbcs.v7i1.27>
- Alhaithloul, H. A. S., Galal, G. H., & Seufi, A. E. M. (2021). Effect of extreme temperature changes on phenolic, flavonoid contents and antioxidant activity of tomato seedlings (*Solanum lycopersicum* L.). *PeerJ*, *9*, e11193. <https://doi.org/10.7717/peerj.11193>
- Almeselmani, M., & Viswanathan, P. S. D. (2012). Effects of prolonged high temperature stress on respiration, photosynthesis and gene expression in wheat (*Triticum aestivum* L.) varieties differing in their thermotolerance. *Plant Stress*, *6*(2), 25–32. Retrieved from [http://www.globalsciencebooks.info/Online/GSBOonline/images/2012/PS\\_6\(1\)/PS\\_6\(1\)25-32o.pdf](http://www.globalsciencebooks.info/Online/GSBOonline/images/2012/PS_6(1)/PS_6(1)25-32o.pdf)
- Alshameri, A., Alqurainy, F., Khan, S., Nadeem, M., Gaafar, A. R., Alameri, A., Tarroum M., Alansi, & Ashraf, S. (2019). Morpho-physiological responses of guar [*Cyamopsis tetragonoloba* (L.) Taub.] to multiple stresses of drought, heat and salinity. *Pakistan Journal Botany*, *51*(3). [https://doi.org/10.30848/PJB2019-3\(5\)](https://doi.org/10.30848/PJB2019-3(5))
- Amada, G., Kosugi, Y., Kitayama, Y., & Onoda, Y. (2020). *Roles of leaf trichomes in heat transfers and gas-exchange characteristics across environmental gradients* (Thesis). Kyoto, Japan: Kyoto University. <https://doi.org/10.22541/au.160794364.42389606/v1>
- Anggraini, N., Faridah, E., & Indrioko, S. (2016). Pengaruh cekaman kekeringan terhadap perilaku fisiologis dan pertumbuhan bibit black locust (*Robinia pseudoacacia*). *Jurnal Ilmu Kehutanan*, *9*(1), 40–56. <https://doi.org/10.22146/jik.10183>
- Aniya, Y., Koyama, T., Miyagi, C., Miyahira, M., Inomata, C., Kinoshita, S., & Ichiba, T. (2005). Free radical scavenging and hepatoprotective actions of the medicinal herb, *Crassocephalum crepidioides* from the Okinawa Islands. *Biological and Pharmaceutical Bulletin*, *28*(1), 19–23. <https://doi.org/10.1248/bpb.28.19>
- Bahar, E., Akter, K. M., Lee, G. H., Lee, H. Y., Rashid, H. O., Choi, M. K., Bhattarai, K. R., Hossain, M. M. M., Ara, J., Mazumder, K., Raihan, O., Chae, H. J., & Yoon, H. (2017).  $\beta$ -Cell protection and antidiabetic activities of *Crassocephalum crepidioides* (Asteraceae) Benth. S. Moore extract against alloxan-induced oxidative stress via regulation of apoptosis and reactive oxygen species (ROS). *BMC Complement and Alternative Medicine*, *17*, 179. <https://doi.org/10.1186/s12906-017-1697-0>
- Bashandy, T., & El-Shaieny, A. H. (2021). Morphological and molecular marker screening for drought tolerance in Egyptian Jew's Mallow (*Corchorus olitorius* L.) landraces. *Acta Universitatis Agriculturae et Silviculturae Mendelianae Brunensis*, *69*(1), 79–90. <https://doi.org/10.11118/ACTAUN.2021.009>
- Can, N. M., Thao, D. T. P., & Gil, G. (2020). Wound healing activity of *Crassocephalum crepidioides* (Benth.) S. Moore. leaf hydroethanolic extract. *Oxidative Medicine and Cellular Longevity*, 2483187. <https://doi.org/10.1155/2020/2483187>
- Carrera, C. S., Solís, S. M., Ferrucci, M. S., Vega, C. C. R., Galati, B. G., Ergo, V., Andrade, F. H., & Lascano, R. H. (2021). Leaf structure and ultrastructure changes induced by heat stress and drought during seed filling in field-grown soybean and their relationship with grain yield. *Anais da Academia Brasileira de Ciencias*, *93*(4). <https://doi.org/10.1590/0001-3765202120191388>
- Chen, Y., Zhang, Z., Tao, F., Palosuo, T., & Rötter, R. P. (2017). Field crops research

- impacts of heat stress on leaf area index and growth duration of winter wheat in the North China Plain. *Field Crop Research*, 222, 230–237. <https://doi.org/10.1016/j.fcr.2017.06.007>
- Dharmadewi, I. M. (2020). Analisis kandungan klorofil pada beberapa jenis sayuran hijau sebagai alternatif bahan dasar food suplement. *Jurnal Emasains*, 9(2), 171–176. <https://doi.org/10.5281/zenodo.4299383>
- Du, L., Yang, H., Song, X., Wei, N., Yu, C., Wang, W., & Zhao, Y. (2022). Estimating leaf area index of maize using UAV-based digital imagery and machine learning methods. *Scientific Reports*, 12, 15937. <https://doi.org/10.1038/s41598-022-20299-0>
- Feng, B., Liu, P., Li, G., Dong, S. T., Wang, F. H., Kong, L. A., & Zhang, J. W. (2014). Effect of heat stress on the photosynthetic characteristics in flag leaves at the grain-filling stage of different heat-resistant winter wheat varieties. *Journal Agronomy Crop Science*, 200(2), 143–155. <https://doi.org/10.1111/jac.12045>
- Guilia S., Berrang, F. L., Zavaleta, C. C., Marshall, L., & Sherilee, L. H. (2020). The effect of climatic factors on nutrients in foods: Evidence from a systematic map. *Environmental Research Letters*, 15(11), 113002. <https://doi.org/10.1088/1748-9326/abafd4>
- Guirao M. L., Entrambasaguas, L., Ruiz, J. M., & Procaccini, G. (2019). Heat-stress induced flowering can be a potential adaptive response to ocean warming for the iconic seagrass *Posidonia oceanica*. *Molecular Ecology*, 28(10), 2486–2501. <https://doi.org/10.1111/mec.15089>
- Guo, T., Gull, S., Ali, M. M., Yousef, A. F., Ercisli, S., Kalaji, H. M., Telesiński, A., Auriga, A., Wróbel, J., Radwan, N. S., & Ghareeb, R. Y. (2022). Heat stress mitigation in tomato (*Solanum lycopersicum* L.) through foliar application of gibberellic acid. *Scientific Report*, 12, 11324. <https://doi.org/10.1038/s41598-022-15590-z>
- Handayani, T., Basunanda, P., Murti, H. R., & Sofiari, E. (2013). Pengujian stabilitas membran sel dan kandungan klorofil untuk evaluasi toleransi suhu tinggi pada tanaman kentang. *Jurnal Hortikultura*, 23(1), 28–35. Retrieved from <http://repository.pertanian.go.id/handle/123456789/773>
- Haworth, M., Marino, G., Brunetti, C., Killi, D., De Carlo, A., & Centritto, M. (2018). The impact of heat stress and water deficit on the photosynthetic and stomatal physiology of olive (*Olea europaea* L.) A case study of the 2017 heat wave. *Plants*, 7(4), 76. <https://doi.org/10.3390/plants7040076>
- Hidayati, N., & Anas, I. (2016). Photosynthesis and transpiration rates of rice cultivated under the system of rice intensification and the effects on growth and yield. *Hayati Journal Bioscience*, 23(2), 67–72. <https://doi.org/10.1016/j.hjb.2016.06.002>
- Hoffmann, W. A., & Poorter, H. (2002). Avoiding bias in calculations of relative growth rate. *Annals Botany*, 90(1), 37–42. <https://doi.org/10.1093/aob/mcf140>
- Hu, S., Ding, Y., & Zhu, C. (2020). Sensitivity and responses of chloroplasts to heat stress in plants heat sensitivity of photosynthesis. *Plant Science*, 11, 375. <https://doi.org/10.3389/fpls.2020.00375>
- Intergovernmental Panen on Climate Change (IPCC). (2021). *Climate change 2021*. Retrieved from [www.ipcc.ch](http://www.ipcc.ch)
- Intergovernmental Panen on Climate Change (IPCC). (2015). *Climate change 2014*. Retrieved from [www.ipcc.ch](http://www.ipcc.ch)
- Istiqomah, D. (2020). Kandungan total fenolik, flavonoid dan aktivitas antioksidan ekstrak bunga jengger ayam (*Celosia cristata* L.) [Thesis]. Bogor: IPB University. Retrieved from <https://repository.ipb.ac.id/handle/123456789/104896>
- Jaisyurahman, U., Wirnas, D., Trikoesoemaningtyas, & Purnamawati, H. (2020). Dampak suhu tinggi terhadap pertumbuhan dan hasil tanaman padi. *Jurnal Agronomi Indonesia*, 47(3), 248–254. <https://doi.org/10.24831/jai.v47i3.24892>
- Liu, Q. H., Wu, X., Li, T., Ma, J. Q., & Zhou, X. B. (2013). Effects of elevated air temperature on physiological characteristics of flag leaves and grain yield in rice. *Chilean Journal of Agricultural Research*, 73(2), 85–90. <https://doi.org/10.4067/S0718-58392013000200001>
- Marantika, M., Hiariej, A., & Sahertian, D. E. (2021). Kerapatan dan distribusi stomata

- daun spesies mangrove di Desa Negeri Lama Kota Ambon. *Jurnal Ilmu Alam dan Lingkungan*, 12(1), 1–6. Retrieved from <http://journal.unhas.ac.id/index.php/jai2/article/view/11041>
- Maulid, R., & Laily, N. A. (2015). Kadar total pigmen klorofil dan senyawa antosianin ekstrak kastuba kadar total pigmen klorofil dan senyawa antosianin ekstrak kastuba (*Euphorbia pulcherrima*) berdasarkan umur daun. *Seminar Nasional Konservasi dan Pemanfaat Sumber Daya Alam*, 1(1), 225–230. Retrieved from <https://jurnal.fkip.uns.ac.id/index.php/kpsda/article/view/5379>
- Mouradov, A., Cremer, F., & Coupland, G. (2002). Control of flowering time: Interacting pathways as a basis for diversity. *The Plant Cell*, 14(suppl\_1), S111–S130. <https://doi.org/10.1105/tpc.001362>
- Obaid, A. Y., Sabir, J. S. M., Atef, A., Liu, X., Edris, S., El-Domyati, F. M., Mutwakil, M. Z., Gadalla, N. O., Hajrah, N. H., Al-Kordy, M. A., Hall, N., Bahieldin, A., & Jansen, R. K. (2016). Analysis of transcriptional response to heat stress in *Rhazya stricta*. *BMC Plant Biology*, 16, 252. <https://doi.org/10.1186/s12870-016-0938-6>
- Owokotomo, I. A., & Owokotomo, E. P. (2018). Anti-bacterial and brine shrimps lethality studies of the essential oils of *Crassocephalum crepidioides* (Benth S. More) grown in south west Nigeria. *African Journal of Pure and Applied Chemistry*, 12(1), 1–7. <https://doi.org/10.5897/ajpac2017.0730>
- Pothitirat, W., Chomnawang, M. T., Supabphol, R., & Gritsanapan, W. (2009). Comparison of bioactive compounds content, free radical scavenging and anti-acne inducing bacteria activities of extracts from the mangosteen fruit rind at two stages of maturity. *Fitoterapia*, 80(7), 442–447. <https://doi.org/10.1016/j.fitote.2009.06.005>
- Rossi, S., & Huang, B. (2022). Carotene-enhanced heat tolerance in creeping bentgrass in association with regulation of enzymatic antioxidant metabolism. *Journal of the American Society for Horticultural Science*, 147(3), 145–151. <https://doi.org/10.21273/JASHS05201-22>
- Sailaja, B., Voleti, S. R., Subrahmanyam, D., Sarla, N., Prasanth, V. V., Bhadana, V. P., & Mangrauthia, S. K. (2014). Prediction and expression analysis of miRNAs associated with heat stress in *Oryza sativa*. *Rice Science*, 21(1), 3–12. [https://doi.org/10.1016/S1672-6308\(13\)60164-X](https://doi.org/10.1016/S1672-6308(13)60164-X)
- Santrum, M. J., Tokan, M. K., & Imakulata, M. M. (2021). Estimasi indeks luas daun dan fotosintesis bersih kanopi hutan mangrove di Pantai Salupu Kecamatan Kupang Barat Kabupaten Kupang. *Haumeni Journal of Education*, 1(2), 38–43. Retrieved from <https://ejournal.undana.ac.id/index.php/haumeni/article/view/5402>
- Sims, D. A., & Gamon, J. A. (2002). Relationships between leaf pigment content and spectral reflectance across a wide range of species, leaf structures and developmental stages. *Remote Sense Environment*, 81(2–3), 337–354. [https://doi.org/10.1016/S0034-4257\(02\)00010-X](https://doi.org/10.1016/S0034-4257(02)00010-X)
- Slimen, I. B., Najar, T., Ghram, A., Dabbebi, H., Mrad, M. B., & Abdrabbah, M. (2014). Reactive oxygen species, heat stress and oxidative-induced mitochondrial damage. A review. *International Journal of Hyperthermia*, 30(7), 513–523. <https://doi.org/10.3109/02656736.2014.971446>
- Subira, M. M., Romero, M. P., Moralejo, M., Macia, A., Puig, E., Savin, R., & Romagosa, I. (2021). Post-anthesis thermal stress induces differential accumulation of bioactive compounds in field-grown barley. *Journal of the Science of Food and Agriculture*, 101(15), 6496–6504. <https://doi.org/10.1002/jsfa.11321>
- Suraweera, D. D., Groom, T., & Nicolas, M. E. (2020). Exposure to heat stress during flowering period reduces flower yield and pyrethrins in *Pyrethrum* (*Tanacetum cinerariifolium*). *Journal of Agronomy and Crop Science*, 206(5), 565–578. <https://doi.org/10.1111/jac.12405>
- Tanimonure, V. A., & Naziri, D. (2021). Impact of climate adaptation strategies on the net farm revenue of underutilized indigenous vegetables' (UIVs) production in Southwest Nigeria. *Resources, Environment and Sustainability*, 5, 100029. <https://doi.org/10.1016/j.resenv.2021.100029>
- Tomimori, K., Nakama, S., Kimura, R., Tamaki, K., Ishikawa, C., & Mori, N. (2012). Antitumor activity and macrophage nitric oxide producing action of medicinal

- herb *Crassocephalum crepidioides*. *BMC Complementary Medicine and Therapies*, 12, 8. <https://doi.org/10.1186/1472-6882-12-78>
- Tsoy, O., & Mushegian, A. (2022). Florigen and its homologs of FT/CETS/PEBP/RKIP/YbhB family may be the enzymes of small molecule metabolism: review of the evidence. *BMC Plant Biology*, 22, 56. <https://doi.org/10.1186/s12870-022-03432-z>
- Verma, D., Aghora, T. S., Laxman, R. H., & Dhananjaya, M. V. (2019). Effect of heat stress on growth indices in garden pea (*Pisum sativum* var. *hortense*). *International Journal of Chemical Studies*, 7(6), 1839–1841. Retrieved from <https://www.chemjournal.com/archives/?year=2019&vol=7&issue=6&ArticleId=7917&si=false>
- Vongsak, B., Pongtip, S., Supachoke, M., Suchitra, T., Yuvadee, W., & Wandee, G. (2013). Maximizing total phenolics, total flavonoids content and antioxidant of *Moringa oleifera* leaf extract by the appropriate extraction method. *Industrial crops and products*, 44, 566–571. <https://doi.org/10.1016/j.indcrop.2012.09.021>
- Wang, Q., Chen, J., He, N., & Guo, F. (2018). Metabolic reprogramming in chloroplasts under heat stress in plants. *International Journal of Molecular Sciences*, 19(3), 849. <https://doi.org/10.3390/ijms19030849>
- Waraich, E. A., Hussain, A., Ahmad, Z., Ahmad, M., & Barutcular, C. (2021). Foliar application of sulfur improved growth, yield and physiological attributes of canola (*Brassica napus* L.) under heat stress conditions. *Journal of Plant Nutrition*, 45(3), 369–379. <https://doi.org/10.1080/01904167.2021.1985138>
- Xu, Y. Y., Wang, J., Nie, S. S., Huang, D. Q., Wang, Y., Xu, L., Wang, R. H., Luo, X. B., & Liu, L. W. (2016). Isolation and molecular characterization of the *FLOWERING LOCUS C* gene promoter sequence in radish (*Raphanus sativus* L.). *Journal Integrative Agriculture*, 15(4), 763–774. [https://doi.org/10.1016/S2095-3119\(15\)61295-3](https://doi.org/10.1016/S2095-3119(15)61295-3)
- Zaripheh, S., & Erdman J. W. (2002). Factors that influence the bioavailability of xanthophylls. *Journal of Nutrition*, 132(3), 531S–534S. <https://doi.org/10.1093/jn/132.3.531S>